



UNIVERSIDADE FEDERAL DE UBERLÂNDIA
INSTITUTO DE BIOTECNOLOGIA

PROGRAMA DE PÓS-GRADUAÇÃO EM GENÉTICA E BIOQUÍMICA

Estudo dos efeitos fisiopatológicos e moleculares do *knockdown* do gene CG15105 em cérebro de *Drosophila melanogaster*

Discente: Matheus Henrique Silva

Orientador: Prof. Dr. Carlos Ueira-Vieira

Co-orientadora: Dra. Jessica Regina Costa Silva

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Aos trinta e um dias do mês de janeiro de dois mil e vinte e dois, às 09:20 horas, reuniu-se via web conferência pela plataforma *Cisco Webex*, em conformidade com a Portaria nº 36, de 19 de março de 2020 da Coordenação de Aperfeiçoamento de Pessoal de Nível Superior – CAPES e Resolução de nº 06/2020 do Conselho de Pesquisa e Pós-graduação pela Universidade Federal de Uberlândia, a Banca Examinadora, designada pelo Colegiado do Programa de Pós-graduação em Genética e Bioquímica, assim composta: Professores Doutores: Carlos Ueira Vieira (Orientador), Anderson de Oliveira Souza e Renata Graciele Zanon. A participação dos dois últimos se deu por epístola. Iniciando os trabalhos o (a) presidente Dr (a). Carlos Ueira Vieira apresentou a Comissão Examinadora e o candidato(a), agradeceu a presença dos participantes, e concedeu ao Discente a palavra para a exposição do seu trabalho. A duração da apresentação do Discente e o tempo de arguição e resposta foram conforme as normas do Programa. A seguir o senhor(a) presidente procedeu a leitura das epístolas enviadas pelos membros da banca. Em seguida os membros presentes, passaram a arguir o(a) candidato(a). Ultimada a leitura das epístolas e a arguição, que se desenvolveu dentro dos termos regimentais, a Banca, em sessão secreta, atribuiu o resultado final, considerando o(a) candidato(a):

(A) PROVADO.

Esta defesa de Dissertação de Mestrado é parte dos requisitos necessários à obtenção do título de Mestre. O competente diploma será expedido após cumprimento dos demais requisitos, conforme as normas do Programa, a legislação pertinente e a regulamentação interna da UFU. Nada mais havendo a tratar foram encerrados os trabalhos. Foi lavrada a presente ata que após lida e achada conforme foi assinada pela Banca Examinadora.

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Dedicatória

Dedico este trabalho a memória de meus avós
Sebastião e Geralda.

Epígrafe

*“Juro não faço por mal
Logo você que diz me conhecer tão bem
Devia saber que isso é tão normal
Ando sem tempo pra você, mas amanhã eu passo pra te ver
Sei que ‘ta’ difícil acreditar em mim
Até os meus amigos dizem que eu sumi
Assumo essa é a vida que eu escolhi
A gente não ‘ta’ perto mas ‘to’ longe de esquecer”*

(Lagum)

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Apresentação

O presente trabalho tem como enfoque estudar aspectos genéticos e bioquímicos relacionados a fisiopatologia da neurodegeneração, utilizando estudos *in silico* e *in vivo* (*Drosophila melanogaster*).

O capítulo 1 se trata da fundamentação teórica. Nele constam informações retiradas de acervos de literatura científica a respeito de conhecimentos necessários para a compreensão do presente trabalho. Este capítulo contém informações sobre doenças neurodegenerativas, o modelo animal utilizado (a *D. melanogaster*) e pôr fim a técnica de interferência de RNA.

Já o capítulo 2 é o artigo científico referente a pesquisa realizada. Neste capítulo constam estudos experimentais realizados com a finalidade de compreender o papel do gene CG15105 nos processos fisiopatológicos da neurodegeneração, e também, as interações moleculares e contribuições da mesma para processos relacionados a neurodegeneração.

RESUMO

A neurodegeneração é um processo fisiopatológico que leva ao comprometimento do Sistema Nervoso, é comumente relacionado com o envelhecimento por se tratar de danos cumulativos ao longo do tempo de vida. Com o aumento da expectativa de vida da população mundial o estudo da neurodegeneração se tornou uma prioridade mundial. A doença de Alzheimer é uma das mais comuns dentre as doenças neurodegenerativas, um dos eventos fisiopatológicos dessa doença é acúmulo do peptídeo β -amiloide, devido ao desequilíbrio entre produção e degradação do mesmo. O acúmulo de β -amiloide se relaciona com outras alterações como a redução da expressão do gene CG15105 (*tn*), que codifica uma proteína envolvida em processos de degradação proteica por meio da ubiquitinação. Devido a isso, o presente trabalho teve como objetivo explorar os efeitos do gene CG15105 em cérebro de *Drosophila melanogaster*. Por meio do uso da técnica de *knockdown* genético com iRNA em cérebro de moscas e análises histopatológicas e comportamentais foi observado que o *knockdown* do gene CG15105 reduz a atividade locomotora, e o *knockdown* causa degeneração da morfologia cerebral. E os achados dos estudos *in silico* do gene CG15105, mostraram que sua atividade está relacionada principalmente com a ubiquinação. E alguns dos genes da rede de interação do CG15105 tem atividade no citoesqueleto e apresentam alteração transcricionais em cérebro de moscas relacionadas aos processos de envelhecimento e presença do peptídeo β -amiloide.

Palavras-chave: Neurodegeneração, *Drosophila melanogaster*, iRNA, *tn* e ubiquitinação.

ABSTRACT

Neurodegeneration is a pathophysiological process leading to an impaired nervous system, it is caused by cumulative damage over a lifetime and it is commonly related to aging. With the increase in life expectancy of the world population, the study of neurodegeneration has become a world priority. Alzheimer's disease is one of the most common neurodegenerative diseases. A pathophysiological event of this disease is the accumulation of the β -amyloid peptide, caused by the imbalance between production and degradation. The accumulation of β -amyloid is related to other alterations such as the expression reduction of gene CG15105 (*tn*), which encode proteins involved in protein degradation processes through ubiquitination. In this way, the present work aimed to explore the effects of the CG15105 gene on the brain of *Drosophila melanogaster*.

Through the use of the technique of genetic knockdown with iRNA in the brain of flies and histopathological and behavioral analyses it was observed that the knockdown of CG15105 gene reduce locomotor activity, and the knockdown of CG15105 causes morphological degeneration of brain. And the findings of the *in silico* studies of the CG15105 show that its activity is mainly related to ubiquitination. And some of the genes in the CG15105 interaction network that have cytoskeleton functions present transcriptional alterations in the brain of flies related to aging processes and the presence of the β -amyloid peptide.

Key-words: Neurodegeneration, *Drosophila melanogaster*, iRNA, *tn*, and ubiquitination.

CAPÍTULO 1: Fundamentação teórica

Doenças neurodegenerativas

Nas doenças neurodegenerativas ocorre o comprometimento de funções do Sistema Nervoso (SN) devido a lesões progressivas nas células que compõem este sistema. Essas lesões podem ter como etiologia o acúmulo proteico em células nervosas, levando ao comprometimento funcional e estrutural do SN. Além disso essas doenças podem compartilhar processos fisiopatológicos semelhantes na neuroinflamação, disfunções na ubiquitinação e estresse proteotóxico (DUGGER; DICKSON, 2017).

As doenças neurodegenerativas são classificadas de acordo com as alterações moleculares, anatômicas e manifestações clínicas. Alguns exemplos são: a Doença Frontotemporal, causada por alterações anatômicas nos lóbulos temporais e frontal; a Doença de Parkinson, caracterizada pela desordem motora e presença de corpos de Lewy; e, por fim, a doença de Alzheimer, que apresenta acúmulo proteico de beta-amiloide e proteína Tau e perda de memória (KOVACS, 2018).

Essas doenças podem causar perda progressiva da capacidade cognitiva caracterizando uma demência. Pessoas com demência tem problemas de memória, que dificultam o reconhecimento pessoas e lugares familiares, também apresentam dificuldades de comunicação, aprendizado, controle emocional, alterações de humor e problemas com atividades de autocuidado (WORLD HEALTH ORGANIZATION, 2021).

Em 2017 a Organização Mundial de Saúde reconheceu a demência como uma prioridade da saúde pública mundial, e propôs que os governos fomentassem e realizassem pesquisas para desenvolvimento de tecnologias direcionadas ao diagnóstico e tratamento dos pacientes com demência (WORLD HEALTH ORGANIZATION, 2017).

Dentre as causas de demência a mais comum é a Doença de Alzheimer (DA) sendo responsável por 60 a 70% dos casos (ERKKINEN; KIM; GESCHWIND, 2018; WORLD HEALTH ORGANIZATION, 2021). Foi estimado que cerca de 6,1 milhões de pessoas eram portadoras da doença de Alzheimer no Estados Unidos da América(EUA) em 2011 com prevalência de 14,7% (RAJAN et al., 2019). O censo dos EUA estimou que em 2020 6,07 milhões de pessoas viviam com DA clínica e a perspectiva é que em 2060 este

número seja de 13,85 milhões de portadores da Doença de Alzheimer clínica (RAJAN et al., 2021). Quando analisado os índices da DA em escala mundial Brasil é o segundo país com maior prevalência da doença (1037 por 100.000) (NICHOLS et al., 2019).

O principal sintoma da DA é a amnesia, que se manifesta com esquecimentos de memória recente e evolui para perdas mais severas. Outras manifestações clínicas também podem ser observadas, dentre elas: problemas cognitivos relacionados a criatividade, ansiedade, alterações sensoriais e alterações de humor (ATRI, 2019).

A Sociedade Americana de Psiquiatria definiu que os critérios para o diagnóstico da doença de Alzheimer são através da sintomatologia e do histórico familiar do paciente. Alguns biomarcadores também podem ser utilizados, como a presença de atrofia cortical e presença da placa senil, detectada em exames histopatológicos pós-morte, ou através de exames de tomografia cerebral por emissão de pósitrons (PET), e também, os níveis da peptídeo β -amiloide 42 ou da Tau fosfato no líquido cérebro-espinhal (LCS). Marcados genéticos, também, podem ser usados para auxiliar no diagnóstico como os genes da proteína precursora de amiloide (APP) e presenilina 1 ou 2 (PSEN1 ou PSEN2). Entretanto, esses marcadores biológicos não estão validados ainda, o que acarreta em diagnósticos com certo grau de incerteza, por isso há possibilidade do diagnóstico ser “provável” ou “possível” doença de Alzheimer (AMERICAN PSYCHIATRIC ASSOCIATION, 2014)

A principal característica da fisiopatologia da doença é formação da placa senil, que consiste em aglomeração do peptídeo β -amiloide (β a) e emaranhados neurofibrilares de Tau. A formação do β a ocorre quando a proteína precursora de amiloide (PPA) é clivada pelas enzimas β -secretase e γ -secretase, essa via de clivagem é uma via alternativa e é chamada de via amiloidogênica, comumente esta clivagem ocorre pelas enzimas α -secretase e γ -secretase e leva a formação de peptídeos solúveis que são degradados. O peptídeo β a possui entre 36 e 42 aminoácidos, e tem funções fisiológicas na sinapse e sobrevivência de neurônios, entretanto o desbalanço entre produção e clearance é tóxico e dá origem a formação das placas de β a, o peptídeo β a de 42 aminoácidos é a isoforma mais citotóxica e com maior potencial patogênico. Esse evento ocorre especificamente nos astrócitos, levando a morte destas células e causando a desmelinização dos neurônios (WANG et al., 2017).

A via amiloidogênica interfere na fosforilação da proteína Tau. Quando há a formação de β a ocorre a hiperfosforilação da Tau, que se liga as estruturas do citoesqueleto celular e constituem os emaranhados neurofibrilares (KUMAR; ABBAS; ASTER, 2015).

A fisiopatologia da doença está relacionada, ainda, com a desregulação do metabolismo da glicose. O cérebro possui uma alta demanda de glicose em relação aos demais órgãos do corpo, e tal metabolismo corrobora para a formação de memórias, uma vez que os astrócitos convertem glicogênio em lactato e transportam para os neurônios durante o processo de consolidação de memórias de longo prazo (ABOLHASSANI et al., 2017).

Além disso, alguns aspectos genéticos associados a doença de Alzheimer, como os genes da proteína precursora de amiloide, presenilina 1 e 2 que, estão relacionados processos fisiopatológicos que levam ao desenvolvimento do Alzheimer precoce, ou familiar, que se dá pela herança genética de mutações nestes três genes (LANE; HARDY; SCHOTT, 2018). Outro associado a etiologia da doença é o gene da APOE. Esse gene possui três variantes alélicas, ϵ 2, ϵ 3 e ϵ 4. A presença do alelo ϵ 4 está associada a fatores de risco para o desenvolvimento e progressão da doença, enquanto que o alelo ϵ 2 em homozigose está associado a um baixo risco de desenvolvimento da DA (SERRANO-POZO; DAS; HYMAN, 2021).

Um trabalho de nosso grupo de pesquisa, que visava avaliar as alterações transcricionais durante a via amiloidogênica, observou que o CG15105 tem sua transcrição reduzida com o envelhecimento e com a presença do acúmulo de β a. Desta forma ambos os genes podem estar relacionados com processos de degeneração do Sistema Nervos Central, seja esse processo ligado ao envelhecimento ou com a Doença de Alzheimer (DA COSTA SILVA et al., 2022; SILVA, 2019). Este trabalho utilizou como animal modelo a *Drosophila melanogaster*, que é um modelo muito utilizado na área da genética.

Modelo animal: *Drosophila melanogaster*

Em 1910, Morgan publicou um estudo que descreveu uma mutação que causava alteração fenotípica da cor dos olhos da *D. melanogaster*. Neste estudo, Morgan observou a ocorrência de mutações espontâneas que deixavam as moscas com os olhos da cor branca, originalmente da cor vermelha. E ao avaliar a herdabilidade de tal fenótipo ele

chegou à conclusão: a mutação ocorria no cromossomo X e era recessiva (STEPHENSON; METCALFE, 2013).

O estudo de Morgan foi um precursor de vários trabalhos que revelaram mecanismos genéticos até então desconhecidos. E dessa forma o uso da *D. melanogaster* como modelo animal de estudo foi ampliado possibilitando encontrar grupos de pesquisa, bancos de dados e bancos de linhagens com diferentes linhagens da mosca como o Estoque Central de Bloomington (banco de linhagens de *Drosophila*) e o Flybase (banco de dados científicos com informações desde ciclo de vida e alimentação até informações genéticas complexas sobre a mosca) (“Bloomington Drosophila Stock Center”, [s.d.]; LARKIN et al., 2021).

A *D. melanogaster* é um modelo animal amplamente utilizado na genética devido ao ciclo de vida curto e por possuir um genoma pequeno em relação a outros animais como humanos e murinos. A *D. melanogaster* possui quatro pares de cromossomos sendo o primeiro o par de cromossomos sexuais e os outros três, os autossômicos. O dimorfismo sexual ocorre devido a quantidade de cromossomos X que a mosca possui, sendo dois (XX) para fêmeas e um (XY) para machos (YAMAGUCHI; YOSHIDA, 2018).

Os primeiros estudos utilizando este modelo revelaram mecanismo genéticos importantes em eucariotos. Com a otimização do uso do modelo foram criadas moscas humanizadas com a finalidade de mimetizar as condições de doenças humanas, permitindo a aplicação das moscas como modelo de estudo. Além de apresentar um ciclo de vida curto, levando cerca de 9 a 10 dias para o desenvolvimento do estágio de ovo até o adulto (FLATT, 2020).

Uma das metodologias de construção de moscas transgênicas é o P-elemento. Ele é um elemento de transposição do DNA que ocorre naturalmente na mosca e em suas extremidades 3' e 5' possui sequências repetitivas que vão se ligar ao DNA genômico com o auxílio de uma transposase que cliva o DNA onde é possível a ligação das sequências repetitivas. Para ser utilizada como ferramenta de edição gênica basta incluir a sequência heteróloga entre as sequências repetitivas, e com o auxílio da transposase e da maquinaria de reparo de DNA a sequência heteróloga será inserida no DNA genômico da mosca (RIO, 1991; WEILGUNY et al., 2020).

Na sequência introduzida é necessário inserir algum marcador fenotípico para avaliar o sucesso da técnica, na maioria das vezes é utilizado a marcação da cor dos olhos.

Para isso são utilizadas moscas de olhos brancos e é introduzido uma sequência que contém o gene para dar a coloração vermelha as moscas. E desta forma é possível separar as moscas que realmente possuem a sequência heteróloga das moscas em que a técnica falhou e não houve introdução da sequência de interesse (KLEMENZ; WEBER; GEHRING, 1987; OSANAI-FUTAHASHI et al., 2012).

Para inserir essa ferramenta de edição gênica é utilizada a técnica de microinjeção em ovos de *D. melanogaster*. Nesta técnica são coletados ovos de moscas e é aplicada uma microinjeção, contendo o elemento P e a transposase, na porção distal dos ovos. Desta forma, as células germinativas das moscas serão afetadas, e quando a mosca eclodir ela carregará a sequência transgênica em suas células germinativas e passará os genes transgênicos para sua prole. Portanto, as moscas transgênicas que serão utilizadas são a prole da mosca que recebeu a microinjeção (LIM et al., 2018; RINGROSE, 2009).

Existem algumas maquinarias que permitem a regulação da expressão heteróloga de genes aplicáveis em *D. melanogaster* transgênicas, dentre elas o sistema UAS-GAL4. Esse sistema é natural de fungos e tem a função de regulação da transcrição de maneira temporal e espacial, ou seja, ele regula em qual fase da vida haverá a transcrição e em qual (quais) tecido (s) será expresso. A GAL4 é uma enzima ligante a sequência UAS para iniciar a transcrição. O transgênico que carrega a sequência GAL4 é chamada de linhagem *Driver*, pois essa linhagem carrega um enhancer a 3' da sequência GAL4 que irá controlar quando e onde a enzima GAL4 será produzida. E o transgênico com a sequência UAS é a linhagem *responder*, por carregar a sequência a ser expressa a 5' da sequência UAS, e é transcrita quando GAL4 se liga a UAS (CAYGILL; BRAND, 2016).

Desta forma, é possível realizar cruzamentos entre linhagens *Driver* e *responder* a fim de obter moscas com expressão de genes em momentos específicos do ciclo de vida e em determinados tecidos, permitindo a análise do efeito de genes de acordo com o tecido e fase do desenvolvimento de sua expressão.

Também é possível avaliar o efeito da não expressão de genes por meio do *knockdown* utilizando a técnica de interferência de RNA. Essa técnica consiste em inserir sequências de DNA codificadoras de dsRNA, que irão se ligar a RNA e levar a sua degradação (MOHR, 2014).

Interferência de RNA

A interferência de RNA é um mecanismo de silenciamento genético pós-transcricional, ou seja, um processo impede a tradução de determinado mRNA. Isso ocorre por meio da ação dos dsRNA (double stand RNA), que são moléculas de RNA em fita dupla capazes de circularem no meio extracelular, e quando entram no citoplasma das células são clivados pela enzima Dicer e dando origem aos siRNA. A proteína Argo se liga aos siRNA para formar o complexo de silenciamento por interferência de RNA, este complexo se liga a uma sequência específica do mRNA para que ele seja clivado e, dessa forma, impossibilita a tradução (KANASTY, 2013).

Os dsRNA tem tamanho maior ou igual a 100 pares de base e possuem propriedades que não são observadas em outros tipos de RNAs, como por exemplo a ligação a proteínas específicas e não formação de estruturas tridimensionais de RNA (REICH; BASS, 2019). Em *D. melanogaster*, os dsRNA se ligam a sequências de RNAs virais como objetivo de realizar a degradação do mesmo, e impedir a ocorrência a replicação viral (SWEVERS; LIU; SMAGGHE, 2018).

Essa técnica pode ser aplicada para terapias gênicas e para pesquisa científica. A terapia gênica pode ocorrer através da administração de sequências de dsRNAs que irão impedir a tradução e produção de proteínas relacionadas a doenças genéticas. O grande desafio dessa aplicabilidade é encontrar um vetor que seja eficiente e capaz de direcionar o dsRNA para células sem que o mesmo sofra degradação pelas enzimas degradadoras de RNAs (SETTEN; ROSSI; HAN, 2019; WENG et al., 2019).

Para a pesquisa científica é possível a produção de animais transgênicos modelos, como *D. melanogaster*, que carregam sequências em seu DNA que irão produzir dsRNA. E ainda, o uso do sistema UAS-GAL4 pode auxiliar nesse processo e permitir o knockdown genético em tecidos e estágios específicos (PERKINS et al., 2015).

Devido a importâncias dessas ferramentas o presente trabalho as utilizou para compreender a contribuição do gene CG15105 para o funcionamento do Sistema Nervoso. Utilizando a ferramenta de RNAi em *Drosophila melanogaster* para compreender como o knockdown de tais genes influência no funcionamento e morfologia cerebral. E por meio de análises de bioinformática busca compreender as interações moleculares relacionadas aos processos neuronais.

Referências

- ABOLHASSANI, N. et al. Molecular pathophysiology of impaired glucose metabolism, mitochondrial dysfunction, and oxidative DNA damage in Alzheimer's disease brain. **Mechanisms of Ageing and Development**, v. 161, p. 95–104, jan. 2017.
- ALBERS, M. W. et al. At the interface of sensory and motor dysfunctions and Alzheimer's Disease. **Alzheimer's & dementia : the journal of the Alzheimer's Association**, v. 11, n. 1, p. 70–98, jan. 2015.
- AMERICAN PSYCHIATRIC ASSOCIATION. **Manual diagnóstico e estatístico de transtornos mentais**. 5. ed. Porto Alegre: Artmed, 2014.
- ATRI, A. The Alzheimer's Disease Clinical Spectrum: Diagnosis and Management. **Medical Clinics of North America**, Neurology for the Non-Neurologist. v. 103, n. 2, p. 263–293, 1 mar. 2019.
- Bloomington Drosophila Stock Center**. Disponível em: <<https://bdsc.indiana.edu/about/index.html>>. Acesso em: 30 nov. 2021.
- BUCHMAN, A. S.; BENNETT, D. A. Loss of motor function in preclinical Alzheimer's disease. **Expert review of neurotherapeutics**, v. 11, n. 5, p. 665–676, maio 2011.
- CAYGILL, E. E.; BRAND, A. H. The GAL4 System: A Versatile System for the Manipulation and Analysis of Gene Expression. Em: DAHMANN, C. (Ed.). **Drosophila: Methods and Protocols**. Methods in Molecular Biology. New York, NY: Springer, 2016. p. 33–52.
- CG17754**. Disponível em: <<https://www.alliancegenome.org/gene/FB:FBgn0030114>>. Acesso em: 7 fev. 2020.
- DA COSTA SILVA, J. R. et al. Differential gene expression by RNA-seq during Alzheimer's disease-like progression in the *Drosophila melanogaster* model. **Neuroscience Research**, fev. 2022.
- DUGGER, B. N.; DICKSON, D. W. Pathology of Neurodegenerative Diseases. **Cold Spring Harbor Perspectives in Biology**, v. 9, n. 7, p. a028035, 7 jan. 2017.
- ERKKINEN, M. G.; KIM, M.-O.; GESCHWIND, M. D. Clinical Neurology and Epidemiology of the Major Neurodegenerative Diseases. **Cold Spring Harbor Perspectives in Biology**, v. 10, n. 4, p. a033118, abr. 2018.
- FLATT, T. Life-History Evolution and the Genetics of Fitness Components in *Drosophila melanogaster*. **Genetics**, v. 214, n. 1, p. 3–48, 1 jan. 2020.
- KANASTY, R. Delivery materials for siRNA therapeutics. **NATURE MATERIALS**, v. 12, p. 11, 2013.
- KLEMENZ, R.; WEBER, U.; GEHRING, W. J. The white gene as a marker in a new P-element vector for gene transfer in *Drosophila*. **Nucleic Acids Research**, v. 15, n. 10, p. 3947–3959, 26 maio 1987.

- KOVACS, G. G. Concepts and classification of neurodegenerative diseases. Em: **Handbook of Clinical Neurology**. [s.l.] Elsevier, 2018. v. 145p. 301–307.
- KUMAR, V.; ABBAS, A.; ASTER, J. C. **Robbins and Cotran Patologia - Bases Patológicas das Doenças**. Philadelphia: Elsevier Health Sciences, 2015.
- LANE, C. A.; HARDY, J.; SCHOTT, J. M. Alzheimer's disease. **European Journal of Neurology**, v. 25, n. 1, p. 59–70, 2018.
- LARKIN, A. et al. FlyBase: updates to the *Drosophila melanogaster* knowledge base. **Nucleic Acids Research**, v. 49, n. D1, p. D899–D907, 8 jan. 2021.
- LIM, B. et al. Visualization of Transvection in Living *Drosophila* Embryos. **Molecular Cell**, v. 70, n. 2, p. 287–296.e6, 19 abr. 2018.
- MOHR, S. E. RNAi screening in *Drosophila* cells and in vivo. **Methods (San Diego, Calif.)**, v. 68, n. 1, p. 82–88, 15 jun. 2014.
- NICHOLS, E. et al. Global, regional, and national burden of Alzheimer's disease and other dementias, 1990–2016: a systematic analysis for the Global Burden of Disease Study 2016. **The Lancet Neurology**, v. 18, n. 1, p. 88–106, jan. 2019.
- OSANAI-FUTAHASHI, M. et al. A visible dominant marker for insect transgenesis. **Nature Communications**, v. 3, p. 1295, 2012.
- PERKINS, L. A. et al. The Transgenic RNAi Project at Harvard Medical School: Resources and Validation. **Genetics**, v. 201, n. 3, p. 843–852, 1 nov. 2015.
- RAJAN, K. B. et al. Prevalence and Incidence of Clinically Diagnosed Alzheimer's Disease Dementia from 1994 to 2012 in a Population Study. **Alzheimer's & dementia : the journal of the Alzheimer's Association**, v. 15, n. 1, p. 1–7, jan. 2019.
- RAJAN, K. B. et al. Population estimate of people with clinical Alzheimer's disease and mild cognitive impairment in the United States (2020-2060). **Alzheimer's & Dementia: The Journal of the Alzheimer's Association**, 27 maio 2021.
- REICH, D. P.; BASS, B. L. Mapping the dsRNA World. **Cold Spring Harbor Perspectives in Biology**, v. 11, n. 3, p. a035352, mar. 2019.
- RINGROSE, L. Transgenesis in *Drosophila melanogaster*. Em: CARTWRIGHT, E. J. (Ed.). **Transgenesis Techniques: Principles and Protocols**. Methods in Molecular Biology. Totowa, NJ: Humana Press, 2009. p. 3–19.
- RIO, D. C. Regulation of *Drosophila* P element transposition. **Trends in Genetics**, v. 7, n. 9, p. 282–287, 1 set. 1991.
- SERRANO-POZO, A.; DAS, S.; HYMAN, B. T. APOE and Alzheimer's Disease: Advances in Genetics, Pathophysiology, and Therapeutic Approaches. **The Lancet. Neurology**, v. 20, n. 1, p. 68–80, jan. 2021.

SETTEN, R. L.; ROSSI, J. J.; HAN, S.-P. The current state and future directions of RNAi-based therapeutics. **Nature Reviews. Drug Discovery**, v. 18, n. 6, p. 421–446, jun. 2019.

SILVA, J. C. **Uso de Drosophila melanogaster como modelo de doenças neurodegenerativas: de análises transcrpcionais à avaliação comportamental.** [s.l.] Universidade Federal de Uberlândia, 17 jul. 2019.

STEPHENSON, R.; METCALFE, N. Drosophila melanogaster: a fly through its history and current use. **The Journal of the Royal College of Physicians of Edinburgh**, v. 43, n. 1, p. 70–75, 21 mar. 2013.

SWEVERS, L.; LIU, J.; SMAGGHE, G. Defense Mechanisms against Viral Infection in Drosophila: RNAi and Non-RNAi. **Viruses**, v. 10, n. 5, p. 230, maio 2018.

WANG, J. et al. A systemic view of Alzheimer disease — insights from amyloid- β metabolism beyond the brain. **Nature Reviews Neurology**, v. 13, n. 10, p. 612–623, out. 2017.

WEILGUNY, L. et al. Reconstructing the Invasion Route of the P-Element in Drosophila melanogaster Using Extant Population Samples. **Genome Biology and Evolution**, v. 12, n. 11, p. 2139–2152, 10 set. 2020.

WENG, Y. et al. RNAi therapeutic and its innovative biotechnological evolution. **Biotechnology Advances**, v. 37, n. 5, p. 801–825, out. 2019.

WORLD HEALTH ORGANIZATION. **Global action plan on the public health response to dementia 2017–2025**. Disponível em: <<https://www.who.int/publications-detail-redirect/9789241513487>>. Acesso em: 10 nov. 2021.

WORLD HEALTH ORGANIZATION. **Dementia**. Disponível em: <<https://www.who.int/news-room/fact-sheets/detail/dementia>>. Acesso em: 10 nov. 2021.

WU, Q. et al. KLHL5 Is a Prognostic-Related Biomarker and Correlated With Immune Infiltrates in Gastric Cancer. **Frontiers in Molecular Biosciences**, v. 7, p. 599110, 10 dez. 2020.

XU, J. et al. Characterization of a novel splicing variant of KLHL5, a member of the kelch protein family. **Molecular Biology Reports**, v. 30, n. 4, p. 239–242, dez. 2003.

YAMAGUCHI, M.; YOSHIDA, H. Drosophila as a Model Organism. Em: YAMAGUCHI, M. (Ed.). **Drosophila Models for Human Diseases**. Advances in Experimental Medicine and Biology. Singapore: Springer Singapore, 2018. v. 1076p. 1–10.

1 **CAPÍTULO 2- The knockdown of CG15105 on *Drosophila***
2 ***melanogaster* makes damage on brain and it has network with genes**
3 **crucial to synaptic and cytoskeletal functions**

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6

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16

17 **Abstract**

18 Background: The CG15105 gene is neurodegeneration-related protein-encoding
19 gene. The present study aimed to explore the pathophysiology effects of knockdown
20 CG15015 on the brain of *Drosophila melanogaster* and the relationship between
21 molecular interaction and neurodegeneration.

22 Methodology: Crosses were performed with transgenic *D. melanogaster* to obtain
23 flies with dsRNA expression in the brain, submitted to the climbing test and
24 histopathological analyses. In *silico* analyses were performed to understand how was the
25 network of the interaction of one of the genes can act on neurodegeneration.

26 Results: The results showed that the knockdown of the CG15105 gene in the brain
27 reduced the rate of climbing of flies to the ages evaluated. Significant histopathological
28 changes were observed, there was presence of vacuoles on knockdown brain. The *in*
29 *silico* analysis was performed to understand interaction network in motor deficit. The
30 results showed that the gene encodes a protein called thin, which is orthologue to the
31 human protein TRIM2 and has a binding site of the ubiquitin E3 and zinc finger (RING),
32 and 46.03% of the genes in the interaction network of this protein are involved with
33 ubiquitin E1 and E2. Thus, this protein may be related to the process of ubiquitination of
34 proteins in which it establishes physical interaction, which is mostly proteins related to
35 the cytoskeleton. This protein also has interaction with other proteins that perform
36 functions in the response to hormonal stimuli and cycle and regulation of development.
37 And finally, an analysis was made using brain transcriptome data from Alzheimer's model
38 flies, and it was observed that transcriptional changes occur in some of the genes of the
39 interaction network of the CG15105 gene, but until then such a relationship between these
40 genes has been described only in muscle tissue.

41 Conclusion: The CG15105 is involved in brain development and functioning, and
42 the knockdown causes a reduction of motor activity and morphological damage on brain..

43 **Keywords:** Neurodegeneration, TRIM, RNAi, ubiquitination.

44 **1. Introduction**

45 The CG15105 gene also called *tn*, has the function of connecting the ubiquitin-
46 protein E3 and has the zinc finger domain. E3 proteins bind stems from binding to other
47 proteins to mediate the ubiquitination process, whose function is to degrade proteins
48 (DAHL-HALVARSSON et al., 2020). When ubiquitination processes are comprised
49 change the proteolytic balance, this happened in neurodegenerative diseases, such as
50 Alzheimer's disease (KURTISHI et al., 2019).

51 Studies show that the *TRIM2* gene plays a crucial function in the nervous system.
52 Mutations in the *TRIM2* gene may lead to the development of axonal neuropathy (LI et
53 al., 2020). In addition, the gene acts in the process of regulating neuron polarization
54 through the process of ubiquitination of light chain filaments and has a key function
55 during neurodevelopment (KHAZAEI et al., 2011).

56 A study developed by our research group showed that there is a reduction in the
57 expression of the CG15105 on brain aging with the presence of β-amyloid accumulation
58 using *Drosophila melanogaster* as an animal model (SILVA, 2019). Thus, the present
59 study aims to revalidate the findings of our group and investigate the contributions of
60 CG15105 to neurodegeneration, through the genetic knockdown of gene in the brain and
61 *in silico* assay to understand molecular interactions and relation to Alzheimer disease.

62

63 **2. Methods**

64 **2.1. *Drosophila melanogaster***

65 For the development of the present study, transgenic strains of *Drosophila*
66 *melanogaster* were utilized, which were obtained by the *Drosophila* Bloomington Stock
67 Center. They were kept in the Genetics Laboratory at the Federal University of
68 Uberlândia a 25°C, with a 12h/12h light/dark cycle. They were fed ad libitum with
69 standard Bloomington culture medium (soy powder 0,01%, glucose 7,2%, agar 0,6%,
70 cornmeal 0,073%, yeast 0,018%, nipagin 0,06% and acid solution 0,05% m/v)

71 Females flies from the elav-Gal4 strain (BL#458) were mated with males of the
72 *w¹¹¹⁸* (BL#3605) and UAS-dsRNACG1505 (BL#67287) strains to obtain negative control
73 and knockdown flies for the CG15105 gene on brian to obtain flies with the genotypes of
74 interest. The elav-Gal4 is a driver strain that will direct the expression of the responder to
75 the brain of the flies, and the UAS-dsRNACG15105 are the responder strains that carry
76 the dsRNA sequence for the mRNA of interest and will perform RNA interference and
77 lead to knockdown of the selected genes. The *w¹¹¹⁸* strain is the mutant strain used as the
78 genetic background for transgene construction, thus this strain was used to obtain flies
79 from the negative control group that had only one copy of the drive transgene, and no
80 responder.

81 Mating was made by anesthetizing flies with carbon dioxide. Female virgin flies
82 - identified by visible meconium - were collected and then placed with male flies of the
83 other strains. The offspring, having the genotype of interest, were collected 10 to 13 days
84 after mating, to obtain flied ranging from 0h to 72h.

85 **2.2. Locomotor activity**

86 The behavior of flies was evaluated using the rapid interactive negative geotaxis
87 test, which is based on the negative geotaxis behavior of *D. melanogaster* and can assess
88 the locomotor ability of the flies. The performance of the flies in this test can be
89 influenced by age and nervous system impairment. Thus, such as test allowed us to assess
90 the influence of genes of interest on neurodegeneration over time (GARGANO et al.,
91 2005).

92 Flies were placed in transparent tubes 9.5 cm high by 2.3 cm in diameter, placed
93 in 12-seat racks, and positioned 40 cm away from a film camera and a fluorescent light.

94 The flies were left in the environment for 20 minutes, and then the pad was tapped three
95 times, and the number of flies able to climb to a height of 5 cm within 4 seconds of the
96 last tap of the pad was evaluated. Flies were submitted to this test at ages of 1-4, 3-6 and
97 8-11 days after eclosion (d.a.e.). The environmental conditions were controlled, the
98 experiment was performed in a place with little noise, with an ambient temperature of
99 25°C ± 2, and always performed at similar times. Three tubes with 20 to 30 flies were
100 used.

101 **2.3. Histopathological analyses**

102 Light microscopy was performed to evaluate morphological changes in the brain
103 caused by the knockdown of the CG15105 gene. Ten adult flies were collected from each
104 experimental group, and anesthetized on ice, and fixed with Carnoy's solution (6:3:1 of
105 99% ethanol, chloroform, and glacial acetic acid) with a solution volume of 10x the
106 sample volume for 24 hours or more, and then the flies were decapitated and the heads
107 were dehydrated with ethyl alcohol in increasing concentrations (70%, 80%, 90%, and
108 100%) for 15 minutes each. Then diaphanized in two xylols for 15 minutes each. And
109 embedded in paraffin by passing through a liquid paraffin bath (57°C to 60°C) for 30
110 minutes, to make the paraffin blocks. The blocks were cut with a microtome to a thickness
111 of approximately 4 µm. The selected sections were placed on glass slides and left to dry
112 in an oven at 40°C for 12 hours.

113 The slides were then deparaffinized with two xylols baths (40 minutes each batch)
114 and hydrated with ethyl alcohol in decreasing concentrations (100%, 90% 80%, and 70%)
115 and then water, stained with Harris' hematoxylin and yellow eosin, dehydrated with ethyl
116 alcohol in increasing concentrations (70%, 80%, 90%, and 100%). Then the slides were
117 sealed with Teflon and coverslips. Images were captured using a light microscope (Leica)
118 coupled with an image capture system (LAS EZ software) at 100x and 1000x
119 magnification.

120 Analysis of the images was done using ImageJ software, to quantify the degree of
121 neurodegeneration, for we used images at 1000x magnification of the eye lobe of the brain
122 of the flies, the images were cut equally to exclude the intercellular spaces of the fly's
123 brain, and leaving only the medullary region of the eye lobe. The total and proportional
124 area occupied by nerve cells was then quantified. The intercellular space rate was

125 calculated according to the percentage of areas with gaps, that is, interstitial space,
126 vacuolar lesions, and fissures.

127 **2.4. *In silico* analyses**

128 The *in silico* tests were performed to understand the structure, function, and
129 interaction network of the protein encoded by the CG15105 gene. For this, databases and
130 software were used to obtain the desired information.

131 To understand the structure queries to the Uniprot, Swiss-model, and Interprot
132 databases were used. To evaluate protein function the Uniprot, Flybase, and NCBI
133 databases were used. Finally, to evaluate the protein interaction network, EsyN, MIST,
134 STRING, and ClueGO software were used.

135 In the Uniprot platform, there were data of five isoforms of the thin protein from
136 *D. melanogaster*, given the similarities of the information, isoform C
137 (B7YZK8_DROME) was used as a guide. The same isoform was considered for the
138 evaluation in Swiss-model and Interprot software.

139 Flybase information was obtained by analysis of the *tn* gene, whose id is
140 FBgn0265356 data obtained from version FB2021_03, released June 15, 2021.

141 For the analysis in the software MIST @ Harvard Medical School v5.0 (April
142 2020), the *D. melanogaster* species was used as a parameter, using a filter that removed
143 the lowest results in the rank.

144 In the analysis using the String database version 11.0 the results of the *tn* protein,
145 C isoforms from *D. melanogaster* were queried, and the results were not separated into
146 groups.

147 For the ClueGO analysis, version v2.5.8 was used. In this software, it was
148 evaluated which genes relate to the CG15105 gene and paired with the database generated
149 in the transcriptome analysis of COSTA-SILVA (2019), and a correlation of their
150 contributions to cellular processes was made.

151 **2.5. Statistical analyses**

152 Results were analyzed using Excel 2016 and GraphPad Prism 8 software. The
153 results of the scaling were analyzed by applying the D'Agostino-Pearson normality test
154 to assess whether the data had a Gaussian distribution, the results pointed out that all

155 groups were with normal distribution, then two-way ANOVA with Tukey's post hoc test
156 was used to assess the statistically significant difference between the treated group and
157 the control group. The histology results were analyzed by applying the D'Agostino-
158 Pearson normality test and the results indicated that the data had a Gaussian (normal)
159 distribution, and then T test was applied.

160 **3. Results**

161 **3.1. The knockdown flies decrease the climb rate compared to**
162 **control group**

163 The knockdown flies of gene CG15105 (elav-GAL4/+; UAS-dsRNACG1505/+)
164 show reduction of climb rate when it compared to control group (elav-GAL4/+; +/+). This
165 reduction could be observed in flies with age 1 to 4 days after eclosion (d.a.e) p=0.0061,
166 and age 3 to 6 d.a.e. (p=0.0014) and 8 to 11 d.a.e. (p=0.0271), Figure 1.

167 **3.2. The knockdown of CG15105 causes morphological alterations**
168 **on brain**

169 The histopathological analysis shows presence of short and abundant vacuoles on
170 knockdown flies' brain (figure2-a), and quantitative analyses on ImageJ show statically
171 differences and increase of intercellular space (p=0.023) in flies' brains caused by the
172 knockdown of gene CG15105 on age 8 to 11 d.a.e. (Figure 2-b).

173 **3.3. Molecular network of gene CG15105 and the relation with**
174 **aging and β-amylid peptide**

175 To understand how these genes, contribute to the neurodegenerative process we
176 choose the gene CG15105 for make *in silico* analyses to realize how the network of this
177 gene operates neuronal process.

178 The gene CG15105, also call by *tn*, encodes THIN protein (Figure 3). This protein
179 is a Tripartite Motif (TRIM) and it has RING domain on N ends. RING's function is
180 binding on zinc and makes transcription regulation. THIN, also, have an ubiquitin E3
181 ligase domain and there is homology with the six-bladed beta-propeller domain.

182 The data obtained with software Uniprot, Flybase and NCBI showed that this
183 protein has activity on translation suppression, binding ubiquitin-protein, and zinc. THIN

184 plays a role in homeostasis and development of muscle cells, and positive regulation of
185 the glycolytic process. The main location on body is at the Z disc on muscle cells.

186 After the structure and role analysis *in silico*, network analyses are performed to
187 understand how this gene can be involved in the neuronal process. Then we observed a
188 diversity network with different genes with varied roles (Table 1).

189 The software EsyN showed the CG15105 has a physical interaction with genes
190 *ald1*, *tm2*, *mib2*, and *pglym78* (Supplementary Figure 1). Also, have a genetic interaction
191 by transcription regulation. The genes *sallimus (sis)* and *estrogen-related receptor (err)*
192 suppress *tn* gene, while *α actin* enhances *tn* expression (Supplementary Figure 2).

193 On the other hand, the *tn* gene could change the expression of other genes. It
194 makes suppression expression of *muscle liim protein at 84b (mlp84p)*, *pdgf- and vegf-*
195 *receptor related (pvr)* and enhance expression of *cin85* and *cd2ap related (cindr)*.
196 Moreover, *tn* could suppress and enhance the expression of *myosin heavy chain (mhc)*.

197 The software MIST could show a varied network of genes that results from
198 predictions and data from others research (Supplementary Figure 3). It is possible to see
199 that are genetic, proteic, and phenotypic interactions.

200 The thin protein has a physics interaction with proteins encodes by *pdha*,
201 *semaphorin 1 a (sema1a)*, *fas2*, and *spn*. These proteins have an important role in the
202 development and function of the nervous system.

203 There is an expression relation between *tn* and genes: *l(2)efl*, *mlp84*, *lectin-galc1*,
204 *cpn*, CG45078, *scla*, *sclb*, *mlc1*, *mp20*, *smyda-9*, CG15247, *prm*, *clect27*, CG5177,
205 CG17107, *aqp*, *kah*, *zasp66*, *neurochondrin*, CG9626, CG9297, CG5023, *mtnc*, *tm2*, *up*,
206 and *sals*. Some of these genes have roles specifics of muscle, while others have roles on
207 carbohydrates metabolism, genetic regulation, membranes, metal binding, and
208 embryological development, that contribute to neurological damage caused by *tn*
209 knockdown.

210 Interologs interactions are protein interactions described in other species, and that
211 this interaction can occur in orthologous proteins in the species in question due to the
212 preservation of biochemical characteristics throughout evolution. The *tn* gene presents
213 interolog interactions with proteins encoded by the *ubc2*, *brat*, *didum*, CG7220, *hsp83*,
214 *ubc4*, and *eff* genes. The *brat* gene (*brain tumor*) encodes a tumor suppressor that

215 regulates the proliferation of cells in the brain. This protein inhibits the translation of
216 proteins, prevents self-renewal, and induces differentiation during the process of dividing
217 neural stem cells (EDWARDS et al., 2003).

218 CG15105 also establishes a phenotypic correlation with other genes, meaning that
219 these genes are similar or phenotypically opposed, these genes may be part of the same
220 protein complex or be involved in similar cellular processes. The genes that establish this
221 correlation with tn are CG31224, *eIF1A*, *Nup358*, *Rpt5*, *mts*, *tinc*, *wdb*, and *E(bx)* (Table
222 1).

223 The analyses on STRING software indicated some genes that had already
224 appeared in the results of previous analyses, such as genes *actin*, *sls*, *mib2*, *mlp84B*, and
225 other genes that does not had previously been observed, such as *dcr-1*, *dcr-2*, *lin-28*, *bub-1*,
226 *ken* and *min* (Supplementary Figure 4).

227 After the analyses described so far, a list of all genes that interact with CG15105
228 and the encoders of proteins that interact with thin (Table 1), an analysis was made in
229 ClueGO with the objective of understanding in which biological processes the genes in
230 question acted (Figure 7). It was observed that 46.3% of the genes worked in the process
231 of active ubiquitin synthesis, mainly E1 and E2, 23.81% of genes act in the process of
232 contraction of muscle fibers, 7.94% act in the responses to hormonal stimuli, 6.35% has
233 the function of negatively regulating cell development, and 6.35% are involved with
234 muscle system functions, 3.17% regulate the JACK-STAT pathway that acts in signaling
235 receptors. Other metabolic functions appeared in lower percentages, such as plasma
236 mRNA concentration, pre-mRNA processing, pyruvate metabolism, and myoblast fusion.

237 To understand how the transcription of genes in this interaction network is
238 affected in a brain by aging with and without the accumulation of the β -amyloid peptide,
239 a comparison between data obtained with bioinformatics analyses and the results obtained
240 in SILVA (2019) was performed. After pairing the data, it was observed that not all genes
241 found in the predictions had their transcriptions altered throughout age, and none of the
242 genes that interact with CG15105 have their transcription altered when comparing the
243 control groups and β -amyloid group. Then, the genes whose transcription is altered over
244 age in both groups were selected and this interaction was evaluated using ClueGO
245 software.

246 It was observed that the genes *tn*, *prm*, *mp20*, *tm2*, *mlc1* and *mhc* had their
247 transcriptions increased with aging without the presence of accumulation of the peptide
248 β-amyloid, while the *l(2)efl* gene has reduced transcription with aging without the
249 presence of accumulation of the β-amyloid peptide in *D. melanogaster* brains (Figure
250 6). Analyses using database information showed that the interaction between the genes
251 mentioned above was described only in specific functions of the muscular system, and
252 now we describe these alterations in *D. melanogaster* heads.

253 The genes *tn*, *prm*, *up*, *tm2*, and *mlc1* had their transcriptions increased with aging
254 with the presence of accumulation of the β-amyloid peptide, while the *l(2)efl* gene has
255 reduced transcription and aging with the presence of accumulation of the β-amyloid
256 peptide in the brains of *D. melanogaster* (Figure 7). Analyses using database information
257 showed that the interaction between the genes mentioned above was described only in
258 specific functions of the muscular system, however this data showed these genes have
259 alterations of transcription on heads.

260 We can observe that there is a difference in the transcription pattern of genes that
261 interact with the *tn* gene throughout aging with and without accumulation of the β-
262 amyloid peptide in *D. melanogaster* heads. The *tn*, *Prm*, *Tm2*, *Mlc1*, and *l(2)efl* genes
263 follow a similar transcription pattern in both aging processes. However, the *mp20* and
264 *mhc* genes have their expression reduced only in aging without the presence of
265 accumulation of the β-amyloid peptide, while the gene *up* has its expression reduced only
266 in aging with the presence of accumulation of the β-amyloid peptide.

267 **4. Discussion**

268 The knockdown on brain of the CG15105 gene caused a reduction in locomotor
269 activity and did not cause significant motor changes. The results of the *in silico* analyses
270 showed that the CG15105 gene and 46.3% of its interaction network are directly related
271 to ubiquitin E1 and E2. Other functions also appeared in significant proportions such as
272 muscle contraction process, hormonal stimuli, and cell development. These results
273 suggest that knockdown CG15105 changes physiology of brain and maybe this process
274 is because of ubiquitination.

275 Among the proteins that establish physical interaction with TRIM are some
276 proteins that act in neurological processes, such as *pdha*, *mib2*, *fas2*, *sema1a*, and *spn*.
277 And in this way, the *tn* protein may be involved with the ubiquitination process of these

278 proteins that perform plasticity and synaptic signaling functions. The *pdha* encodes a
279 protein that has dehydrogenase pyruvate functions and contributes to acetyl-CoA
280 synthesis (WISIDAGAMA; THUMMEL, 2019). And the *sema1a* encodes a protein
281 transmembrane with the role to control dendrites and the axon direction, this gene is
282 essential to motor neurons functions, and also, have an interation with the receptor
283 encodes by *plexa* (HONG et al., 2020; JEONG, 2017). In this way, there is another one
284 in this network involved in synaptic activity, it is *spn* that regulates the signal of neurexin
285 and neuroligin, that active presynaptic zone (RAMESH et al., 2021). And the *fas2*
286 encodes a protein that regulates nerve fascicles using molecular connexion with axon
287 MP1 pathway and play roles on motor and memories formation (CHENG et al., 2001;
288 KIM; WEN; JAN, 2009).

289 The TRIM2 protein was already with the function of ubiquitination of light chain
290 neurofilament, which led to the contribution to neuronal polarization during neuronal
291 development (KHAZAEI et al., 2011). Also, the deficient of TRIM2 cause to
292 accumulation of light chain neurofilaments on brain, and this cause neurodegeneration,
293 how we showed with climb test, the behavior can change (BALASTIK et al., 2008).

294 In addition, the TRIM2 protein is related to cellular plasticity in the early stages
295 of neurological development, and suppression of the TRIM2 coding gene leads to cellular
296 apoptosis (LOKAPALLY et al., 2020).

297 In the last analysis in which it was considerations, specific transcriptions of
298 *Drosophila* heads showed genes that until then had important interactions for muscle
299 functions. This may have occurred because most of them are related to cytoskeletons, and
300 the findings related to functions are more related to muscle contraction. However, our
301 results suggest that these genes are associated with brain functions, and transcriptional
302 changes are associated with neurodegeneration, aging, and motor deficit.

303 **5. Conclusion**

304 The *tn* (CG15105) gene is involved in genetic and biochemical processes related
305 to brain development and functioning and has an interaction network capable of
306 regulating different neurodegeneration-related cellular processes, such as ubiquity and
307 synaptic regulation. And more studies are needed to track in more detail the genetic and
308 protein changes caused by the knockdown of this gene, including in a pathophysiological
309 situation of β-amylid peptide accumulation.

310 **Declaration of Competing Interest**

311 The authors declare that they have no known competing financial interests or
312 personal relationships that could have appeared to influence the work reported in this
313 paper.

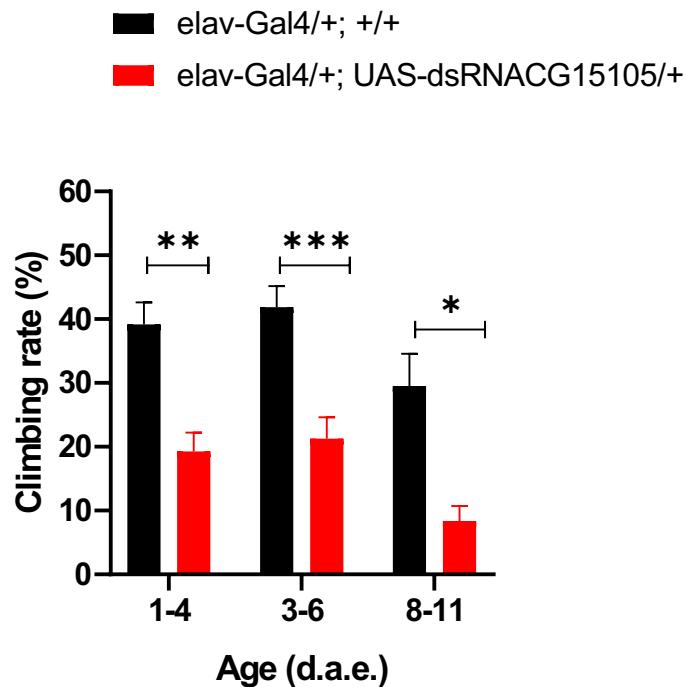
314 **Acknowledgment**

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316 Institute of Biotechnology for supporting and funding the research. And Fundação de
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321 **References**

- 322 BALASTIK, M. et al. Deficiency in ubiquitin ligase TRIM2 causes accumulation of
323 neurofilament light chain and neurodegeneration. **Proceedings of the National**
324 **Academy of Sciences**, 6 ago. 2008.
- 325 CHENG, Y. et al. Drosophila fasciclinII is required for the formation of odor memories
326 and for normal sensitivity to alcohol. **Cell**, v. 105, n. 6, p. 757–768, 15 jun. 2001.
- 327 DAHL-HALVARSSON, M. et al. Impaired muscle morphology in a Drosophila model
328 of myosin storage myopathy was suppressed by overexpression of an E3 ubiquitin
329 ligase. **Disease Models & Mechanisms**, v. 13, n. 12, p. dmm047886, 29 dez. 2020.
- 330 EDWARDS, T. A. et al. Model of the Brain Tumor–Pumilio translation repressor
331 complex. **Genes & Development**, v. 17, n. 20, p. 2508–2513, 15 out. 2003.
- 332 HONG, Y. G. et al. Identification of cis-Regulatory Region Controlling Semaphorin-1a
333 Expression in the Drosophila Embryonic Nervous System. **Molecules and Cells**, v. 43,
334 n. 3, p. 228–235, 31 mar. 2020.
- 335 JEONG, S. Visualization of the Axonal Projection Pattern of Embryonic Motor Neurons
336 in Drosophila. **Journal of Visualized Experiments : JoVE**, n. 124, p. 55830, 16 jun.
337 2017.
- 338 KIM, M. D.; WEN, Y.; JAN, Y.-N. Patterning and organization of motor neuron
339 dendrites in the Drosophila larva. **Developmental Biology**, v. 336, n. 2, p. 213–221, 15
340 dez. 2009.
- 341 KURTISHI, A. et al. Cellular Proteostasis in Neurodegeneration. **Molecular**
342 **Neurobiology**, v. 56, n. 5, p. 3676–3689, maio 2019.

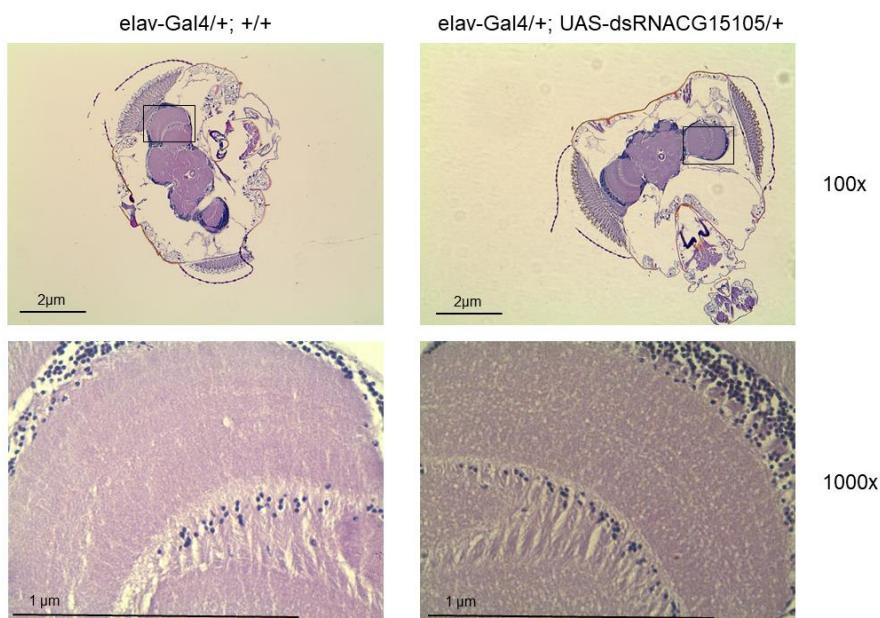
- 343 LOKAPALLY, A. et al. Interplay of TRIM2 E3 Ubiquitin Ligase and ALIX/ESCRT
344 Complex: Control of Developmental Plasticity During Early Neurogenesis. **Cells**, v. 9,
345 n. 7, p. 1734, jul. 2020.
- 346 RAMESH, N. et al. Antagonistic interactions between two Neuroligins coordinate pre-
347 and postsynaptic assembly. **Current biology: CB**, v. 31, n. 8, p. 1711- 1725.e5, 26 abr.
348 2021.
- 349 SILVA, J. C. **Uso de Drosophila melanogaster como modelo de doenças**
350 **neurodegenerativas: de análises transpcionais à avaliação comportamental.** [s.l.]
351 Universidade Federal de Uberlândia, 17 jul. 2019.
- 352 WISIDAGAMA, D. R.; THUMMEL, C. S. Regulation of Drosophila Intestinal Stem
353 Cell Proliferation by Enterocyte Mitochondrial Pyruvate Metabolism. **G3:**
354 **Genes|Genomes|Genetics**, v. 9, n. 11, p. 3623–3630, 5 set. 2019.
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- 356

Figures

359 **Figure 1- Evaluation of the climb rate in different days after the eclosion of**
 360 **knockdown flies for the CG15105 gene and negative control.** Percentage of flies that
 361 climbed height equal to or greater than 5 cm after four seconds. Legend represents the
 362 genotype of the flies of each group. The bars represent the arithmetic mean \pm standard
 363 error. Two-way ANOVA was applied with Tukey *post hoc* test to evaluate the
 364 significance of the difference of the knockdown group about the control in each age. It
 365 was represented with * $p \leq 0.05$, ** $p \leq 0.01$, and *** $p \leq 0.001$.

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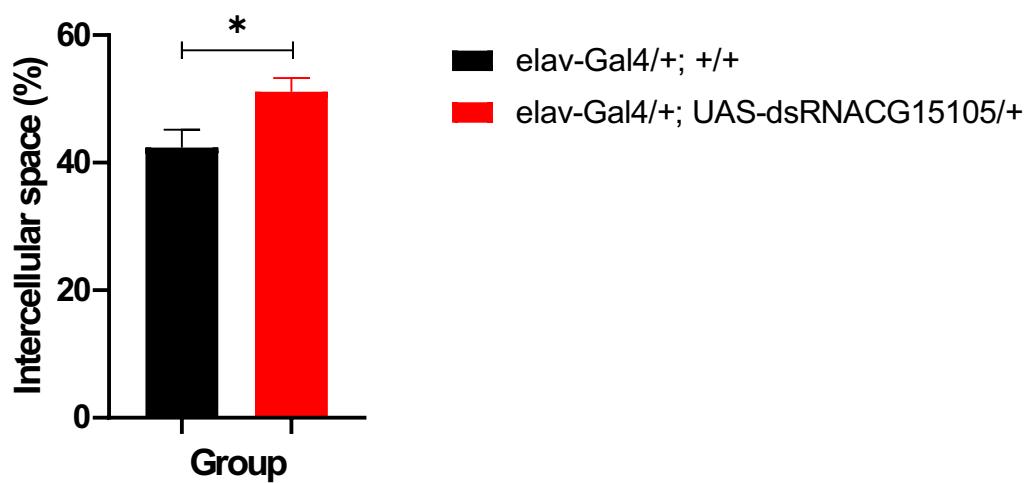
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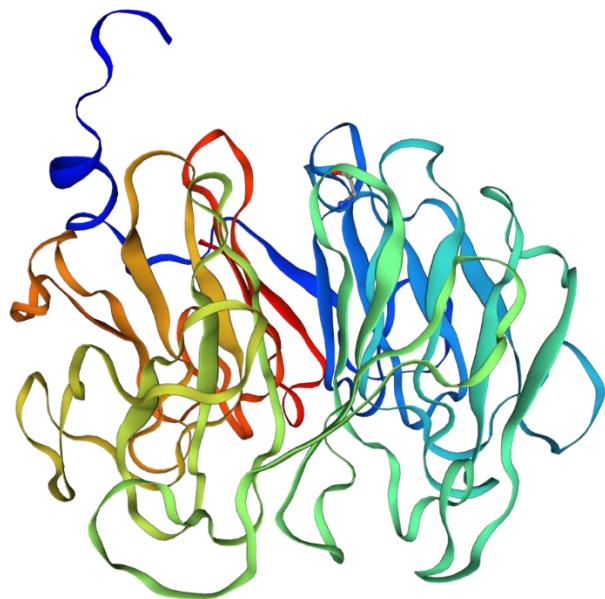
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371 **Figure 2- Evaluation of histopathological changes induced by knockdown of**
 372 **CG1505 genes in the brain of *D. melanogaster*. A-** Images of histopathological slice of
 373 heads of *D. melanogaster* at 8 to 11 d.a.e.. Samples smeared with hematoxylin and eosin
 374 and images captured using Leica light microscope with the aid of LAS EZ software with
 375 an increase of 100x and 1000x.B- Graph representing the brain intercellular space of each
 376 group at the age of 8 to 11 days. The T test with was performed and found p=0.023.

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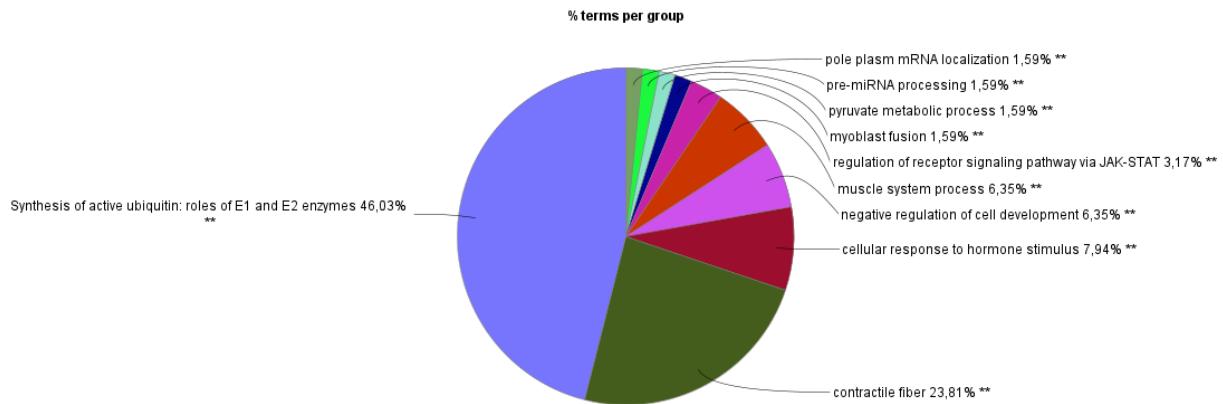
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379 **Figure 3 - 3D structure of the thin protein.** Image from Uniprot

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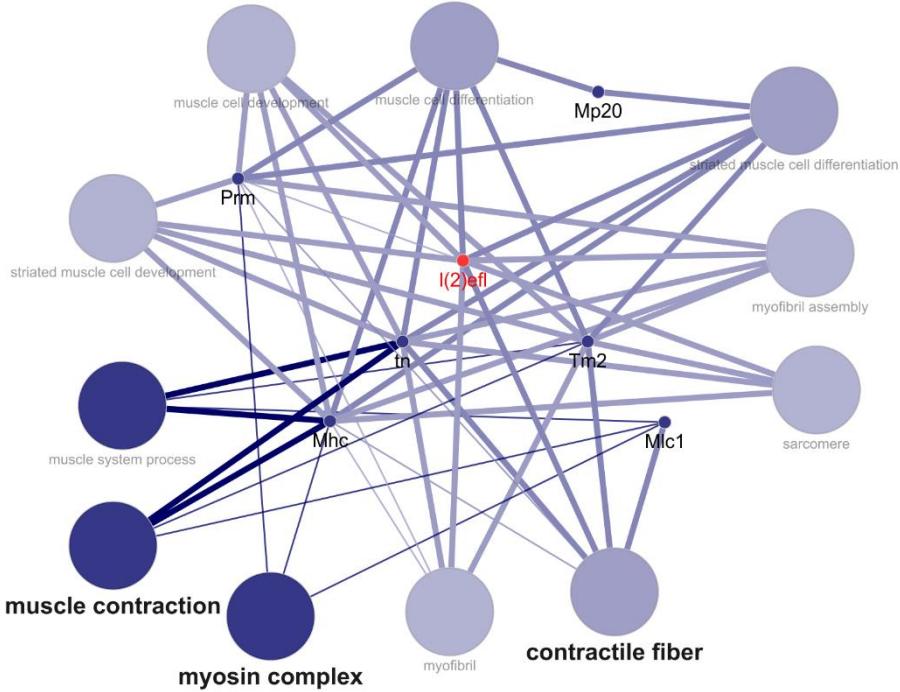
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384 **Figure 4- Functions of genes of CG15105 network.** Image obtained with the analyses
385 performed by the ClueGO bioinformatic tool.

386

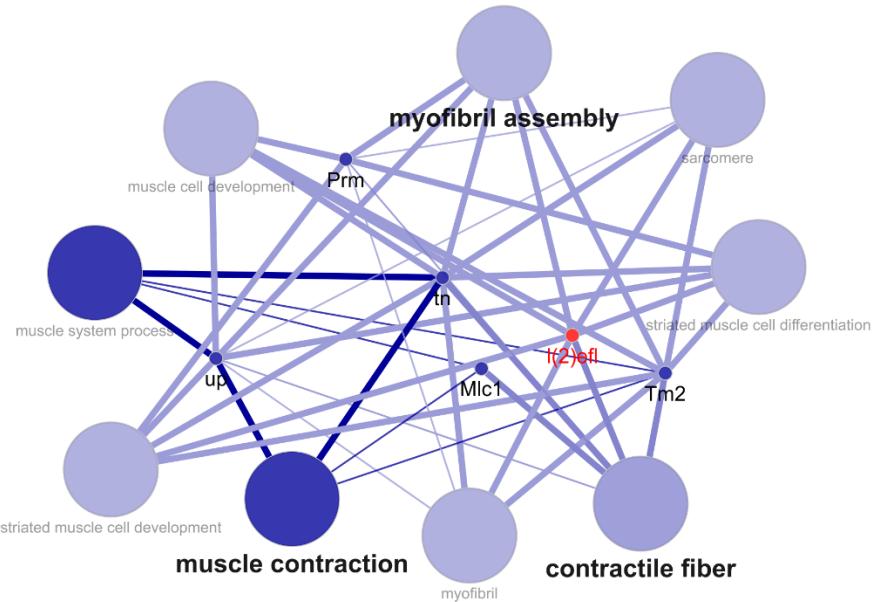


387

388 **Figure 5- Results of the analysis of the transcripts of the negative control**
 389 **group with 0-3 d.a.e. and 10 d.a.e.** In blue are the genes that have increased their
 390 transcription with aging and in red the gene that had its transcription reduced. This
 391 network showed genes that are related to aging on brain and are on network on CG15105
 392 gene, and the cell process that was describe.

393

394



395

396 **Figure 6- Results of the analysis of the transcripts of the group with the**
397 **accumulation of β -amyloid with 0-3 d.a.e. and 10 d.a.e.** In blue are the genes that have
398 increased their transcription with aging and in red the gene that had its transcription
399 reduced. This network showed genes that are related to aging an accumulation of β -
400 amyloid on brain and are on network on CG15105 gene, and the cell process that was
401 descrete.

402

403

Table 1

404

Table of *tn* network- Data from analyses and data bases of softwares: EsyN, MIST, STRING, FlyBase and NCBI.

| REFERENCE | GENE | NAME | PROTEIN | INTERACTION WITH CG15105 | FUNCTION |
|---------------|---------|---------------------------|---------|--|---|
| <i>tn</i> | CG15105 | Thin | Tn | - | The protein of the TRIM/RBC family plays a role in the structure of myofibrils. |
| <i>sls</i> | CG1915 | Sallimus | sls | Suppresses expression/ Co-expression | The protein binds to myosin filaments on the Z disc. |
| <i>mhc</i> | CG17927 | Myosin Heavy chain | MHC | It has the expression regulated | Plays a role in muscle contraction. |
| <i>actn</i> | CG4376 | α -actin | Actn | Increases expression/ Co-expression | Gene encodes specific actin proteins of muscle cells and actin from other tissues that form the cytoskeleton of the cell. |
| <i>mlp84b</i> | CG1019 | Muscle LIM protein at 84B | MLP84B | It has the expression suppress/ Coexpression | Encodes a protein that makes up the cytoskeleton of muscle cells. |
| <i>ald1</i> | CG6058 | Aldolase 1 | Ald1 | Physics | Encodes an enzyme that acts on two targets, fructose-1,6- |

| | | | | | |
|----------------|---------|---|---------|------------------------|--|
| <i>tm2</i> | CG4843 | Tropomyosin 2 | TM2 | Physics | biphosphatase and fructose-1-phosphatase It is associated with the troponin complex and plays an important role in regulating calcium-dependent muscle contraction. |
| <i>mib2</i> | CG17492 | Mind Bomb 2 | Mib2 | Physics/ Co-expression | Encodes a protein that acts on synaptic plasticity and muscle integrity. |
| <i>pglym78</i> | CG1721 | Phosphoglyceromutase 78 | pglym78 | Physics | Encodes an enzyme that performs activities in the fusion of myofibers and the somatic development of muscle cells. |
| <i>fas2</i> | CG3665 | Fasciclin 2 | Fas2 | Physical interaction | It is involved with the development of the nerve fascicle, through molecular reconnection with the MP1 pathway of the axon. |
| <i>pdha</i> | CG7010 | Pyruvate dehydrogenase E1 alpha subunit | Pdha | Physical interaction | Pyruvate dehydrogenase activity, more specifically acetyl-transferase, and participates in the biosynthesis of acetyl-CoA |

| | | | | | |
|---------------------|---------|----------------------------------|--------------|----------------------|---|
| <i>sem1a</i> | CG18405 | Semaphorin 1a | sem1a | Physical interaction | It is a transmembrane protein and plays a receptor role that regulates the target of dendrites and the direction of axons. It also establishes interactions with the receiver encoded by PlexA. |
| <i>spn</i> | CG16757 | Spinophilin | Spn | Physical interaction | It is a protein that regulates neurixin/neuroligin signaling that activates the pre-synaptic zone. |
| <i>l(2)efl</i> | CG4533 | lethal (2) essential for life | L(2)EFL | Expression | It has a vital function during embryological development |
| <i>lectin-galC1</i> | CG9976 | Galactose-specific C-type lectin | Lectin-galC1 | Expression | Encodes a protein, it binds to galactose and is involved with cell-cell adhesion. |
| <i>cpn</i> | CG4795 | Calphotin | Cpn | Expression | It has Ca^{2+} buffer function in photoreceptor microvilli, this occurs due to strong negative charge, and protein composition rich in glutamic acid and highly hydrophobic amino acids. |

| | | | | | |
|----------------|---------|----------------|----------------|------------|---|
| | | | | | |
| CG45076 | CG45076 | No information | No information | Expression | acids. With this, calphotin protects photoreceptor cells from Ca^{2+} overload and degeneration caused by light. |
| CG45078 | CG45078 | No information | No information | Expression | It is involved with the transport of proteins, it has been predicted that it may be present in the plasma membrane and is expressed in the head and heart in adult flies. |
| <i>scla</i> | CG45090 | Sarcolamban A | SclA | Expression | Encodes a protein and is expressed in the head and heart in adult flies. |
| <i>sclb</i> | CG45091 | Sarcolamban B | SclB | Expression | Encodes a protein that plays a central role in regulating calcium transport in the sarcoplasmic reticulum and regulating muscle contraction. |

| | | | | | |
|----------------|---------|--|----------------|------------|---|
| <i>mlc1</i> | CG5596 | Myosin alkali light chain | Mlc1 | Expression | Encodes a protein that binds to calcium ions located in the cytosol and acts on the development of mesoderm. It's part of the myosin complex of muscle cells. |
| <i>mp20</i> | CG4696 | Muscle protein 20 | Mp20 | Expression | Encodes a protein that acts in the regulation of myoblast fusion. And it can be found in the cytoplasm. |
| <i>smyda-9</i> | CG12119 | SET and MYND domain-containing, arthropod-specific, member 9 | SmydA-9 | Expression | It binding activity to histone deacetylase and lysine-N-methyltransferase protein, which negatively regulate gene expression |
| <i>CG15247</i> | CG15247 | No information | No information | Expression | Encodes a protein, the expression was observed in the embryonic, larval and adult male stages. No more information about its expression and function. |
| <i>prm</i> | CG5939 | Paramyosin | Prm | Expression | Encodes a specific structural protein of muscle cells of |

| | | | | | |
|----------------|---------|------------------------------|-----------------------------------|------------|---|
| | | | | | invertebrates, can phosphorylate and this acts on an important muscle function. |
| <i>clect27</i> | CG3244 | Schlaff | SLF | Expression | Encodes a chitin-binding protein that is involved in the formation of the asa. |
| CG5177 | CG5177 | enzyme Trehalose-phosphatase | Trehalose 6-phosphate phosphatase | Expression | Participates in the process of biosynthesis of trehalose, is located in the extracellular medium, and is expressed in adult heads and other organs of the fly body. |
| CG17107 | CG17107 | No information | No information | Expression | The gene encodes a protein that is expressed in the middle intestine in the embryonic and larval stages. |
| <i>aqp</i> | CG12251 | aquaporin | AQP | Expression | Encodes a protein that has channel function, is active in cytoplasmic, and is expressed in adult fly heads. |
| <i>kah</i> | CG17181 | Kahuli | Kah | Expression | Regulates transcription by connecting to E-box |

| | | | | | |
|----------------------|--------|---|----------------|------------|---|
| <i>zasp66</i> | CG6416 | Z band alternatively spliced PDZ-motif protein 66 | Zasp66 | Expression | Encodes that protein that binds actin, is located in the Z disk of the muscle fiber. |
| <i>neurochondrin</i> | CG2330 | Neurochondrin | Neurochondrin | Expression | It has the role of up-regulation of the MHC, is involved in the process of regulation of synaptic plasticity and development of neurons. |
| CG9626 | CG9626 | No information | No information | Expression | No information |
| CG9297 | CG9297 | No information | No information | Expression | Encodes a protein that is involved with the process of endocytosis and transport by binding to organelle membranes and the plasma membrane. |
| CG5023 | CG5023 | No information | No information | Expression | It binds to actin and has the function of organizing the actinomyosin complex and is active in the cytoskeleton. |
| <i>mtnc</i> | CG5097 | Metallothionein C | MtnC | Expression | It connected to metal ions |
| <i>up</i> | CG7107 | upheld | Troponin T | Expression | It encodes a protein that regulates tropomyosin and is found to thin |

| | | | | | |
|--------------|---------|------------------------------------|-------------------------------|------------|--|
| | | | | | filaments in the muscle sarcomere. |
| <i>sals</i> | CG31374 | sarcomere length short | Sals | Expression | Encodes a protein that binds actin and acts on the growth of muscle fiber. |
| <i>ubc2</i> | CG6720 | Ubiquitin-conjugating enzyme 2 | Ubc2 | Interolog | Catalyze the covalent binding of ubiquitin to other proteins |
| <i>brat</i> | CG10719 | brain tumor | Brat | Interolog | Encodes a tumor suppressor that regulates the proliferation of cells in the brain. This protein inhibits the translation of proteins, prevents self-renewal, and induces differentiation during the process of dividing neural stem cells. |
| <i>didum</i> | CG2146 | dilute class unconventional myosin | Didum | Interolog | It has mitochondrial transport function in neuronal cells. |
| CG7220 | CG7220 | CG7220 | Conjugate enzyme ubiquitin E2 | Interolog | It is involved with the activity of proteases that mediate the process of protein catabolism, monoubiquitination, and protein polyubiquitination. |

| | | | | | |
|--------------|--------|-----------------------|-------|-----------|--|
| <i>hsp83</i> | CG1242 | Heat shock protein 83 | HSP83 | Interolog | Encodes chaperones that promote maturation, structural maintenance, and regulation of the properties of specific proteins that have the function of controlling the cell cycle. The gene goes through a functional cycle that is linked to ATPase activity, this cycle induces conformational changes in proteins leading to its activation. And it interacts dynamically with co-chaperones that recognize the substrate, ATPase cycle, and functional chaperones. Together with Hop and piwi, they support the channelization and development via epigenetic silencing of genetic variants and suppression of genetic variants induced by transposons. Required by |
|--------------|--------|-----------------------|-------|-----------|--|

| | | | | | |
|-------------|---------|---------------------------------------|------|------------------------|--|
| | | | | | piRNA biogenesis and facilitated the loading of piRNA inside piwi proteins. |
| <i>ubc4</i> | CG8284 | Ubiquitin-conjugating enzyme 4 | UBC4 | Interolog | Catalyze the covalent binding of ubiquitin to other proteins |
| <i>eff</i> | CG7425 | Effete | Eff | Interolog | Encodes the conserved part of the enzymes conjugated to ubiquitin E2 class I which functions of ubiquitination and degradation and is related to the regulation of apoptosis and chromatin organization. |
| <i>rpt5</i> | CG10370 | Regulatory particle triple-A ATPase 5 | Rpt5 | phenotypic correlation | Encodes a component of proteasome 26S, which degrades polyubiquitinated proteins in the cytoplasm and nucleus. |
| <i>tinc</i> | CG31247 | tincar | Tinc | phenotypic correlation | Encodes a transmembrane protein that is involved with regulating the development of cellular omatides. |

| | | | | | |
|---------------|---------|---------------------------|--------|------------------------|--|
| <i>e(bx)</i> | CG32346 | Enhancer of bithorax | E(bx) | phenotypic correlation | <p>It is a component of binding to <i>histone remodeling factor (NURF)</i>, which is an ATP-dependent catalysis complex of nucleosome slippage and transcription facilitator</p> |
| <i>elf1a</i> | CG8280 | Elongation factor 1 alpha | ElF1A | phenotypic correlation | <p>Encodes proteins necessary for mRNA transcription, is involved with the 43S pre-initiation complex, ensuring the selection of the aug codon start and its dissociation by the ribosome subunit 40S.</p> |
| <i>nup358</i> | CG11856 | Nucleoporin 358kD | Nup358 | phenotypic correlation | <p>Encodes a protein that makes up the nuclear pore complex, which regulates transport through the nuclear envelope and plays a crucial role in the transport of mRNA by recruiting the mRNA transport complex composed of Nxt1 and sbr/Nxf1</p> |

| | | | | | |
|----------------|---------|------------------|----------------|------------------------|--|
| <i>wdb</i> | CG5643 | widerborst | wdb | phenotypic correlation | Encodes a subunit B' regulatory of the serine/threonine phosphatase PP2A complex, which regulates Hedgehog, a protein kinase B, and the insulin receptor signaling pathway. |
| <i>mts</i> | CG7109 | star microtubule | mts | phenotypic correlation | Encodes the a2 phosphatase subunit, which is involved in various development processes and signaling pathways. |
| CG31224 | CG31224 | No information | No information | phenotypic correlation | It is a protein-coding gene that has the function of linking to DNA and suppressing transcription. It is involved with the regulation of transcription by RNA polymerase II. |
| <i>bub2</i> | CG14030 | Bub1 kinase | Bub2 | Co-expression | Encodes a protein that participates in a checkpoint and is involved with the formation of the |

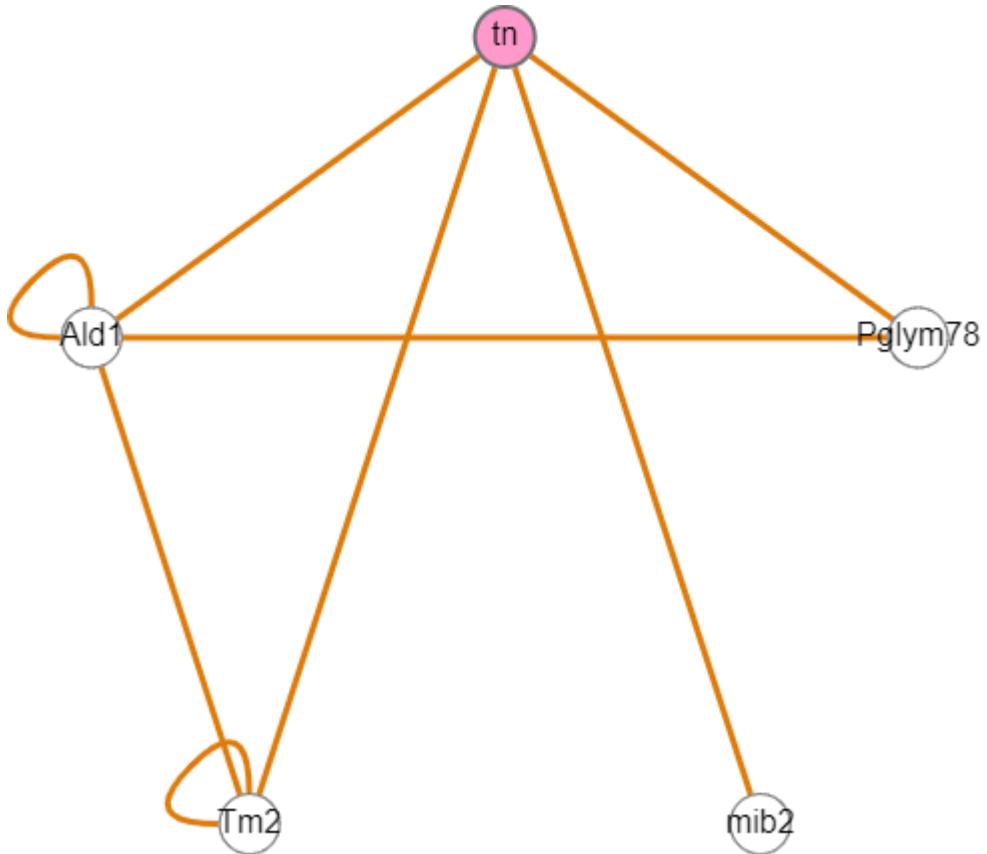
| | | | | | |
|---------------|---------|-----------------------|--------|---------------|--|
| <i>dcr-1</i> | CG4792 | Dicer-1 | Dcr-1 | Co-expression | cytolytic spindle and the endocytosis of parasites. It encodes an enzyme that participates in the RNA interference process, the enzyme catalyzes the cleavage of dsRNA. |
| <i>dcr-2</i> | CG6493 | Dicer-2 | Dcr-2 | Co-expression | It encodes an enzyme that participates in the RNA interference process, the enzyme catalyzes the cleavage of dsRNA, specifically cleave viral RNA. |
| <i>ken</i> | CG5575 | ken and barbie | Ken | Co-expression | Encodes a protein that is a transcription factor and participates in the process of genital formation. |
| <i>lin-28</i> | CG17334 | Lin-28 | Lin-28 | Co-expression | Encodes an RNA-binding protein that participates in the micro RNA maturation process. |
| <i>me</i> | CG33558 | missing-in-metastasis | me | Textmining | Encodes a protein involved in guided cell migration by inhibiting the endocytosis of the |

product encoded by
Cortactin.

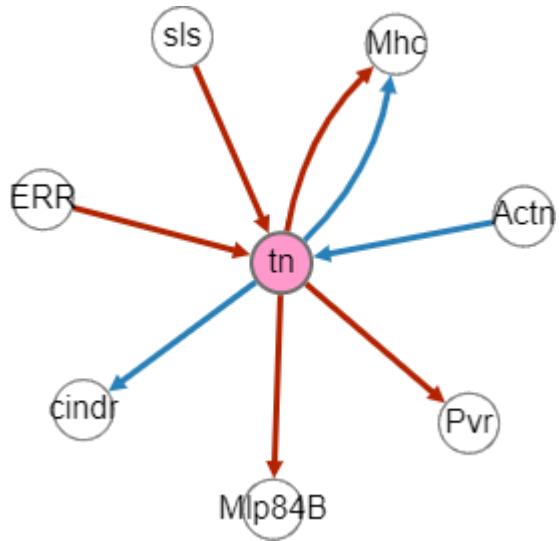
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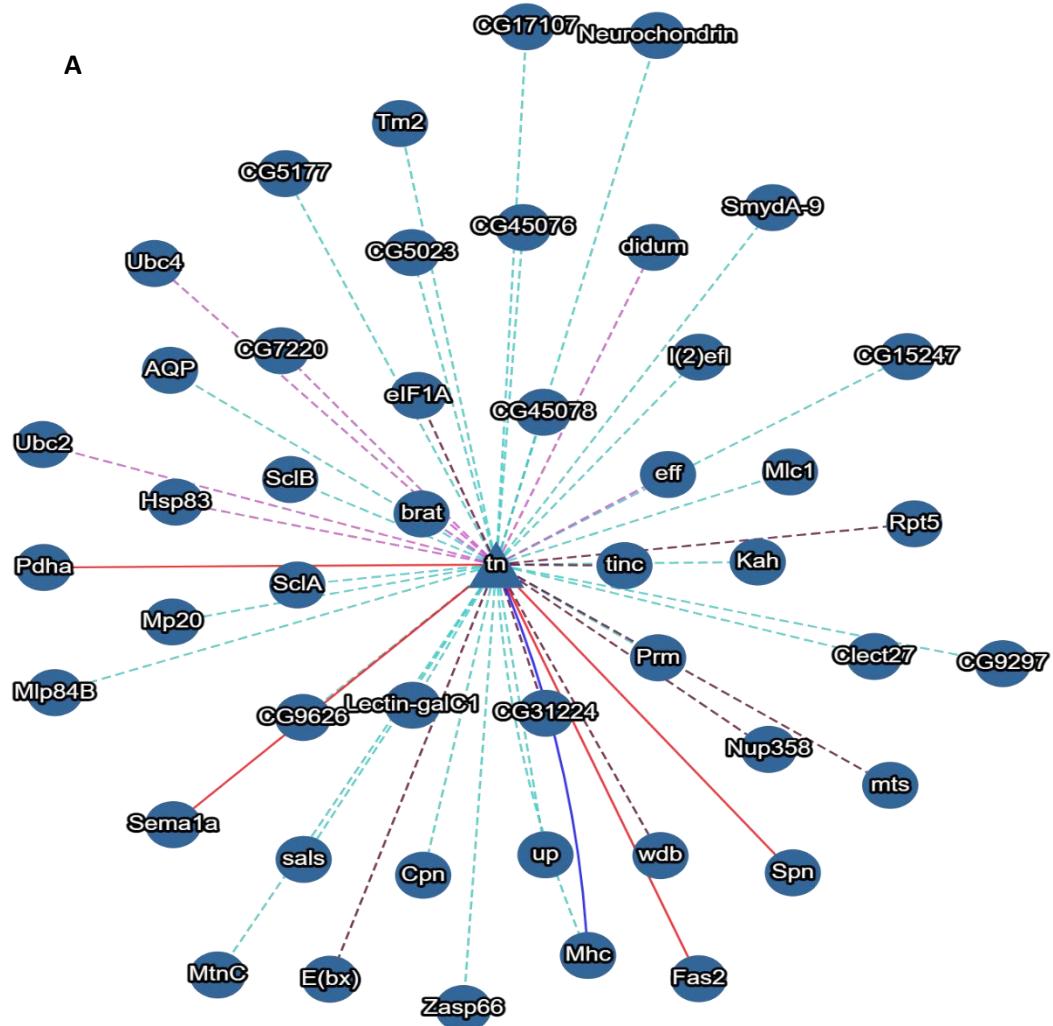
Supplementary figures



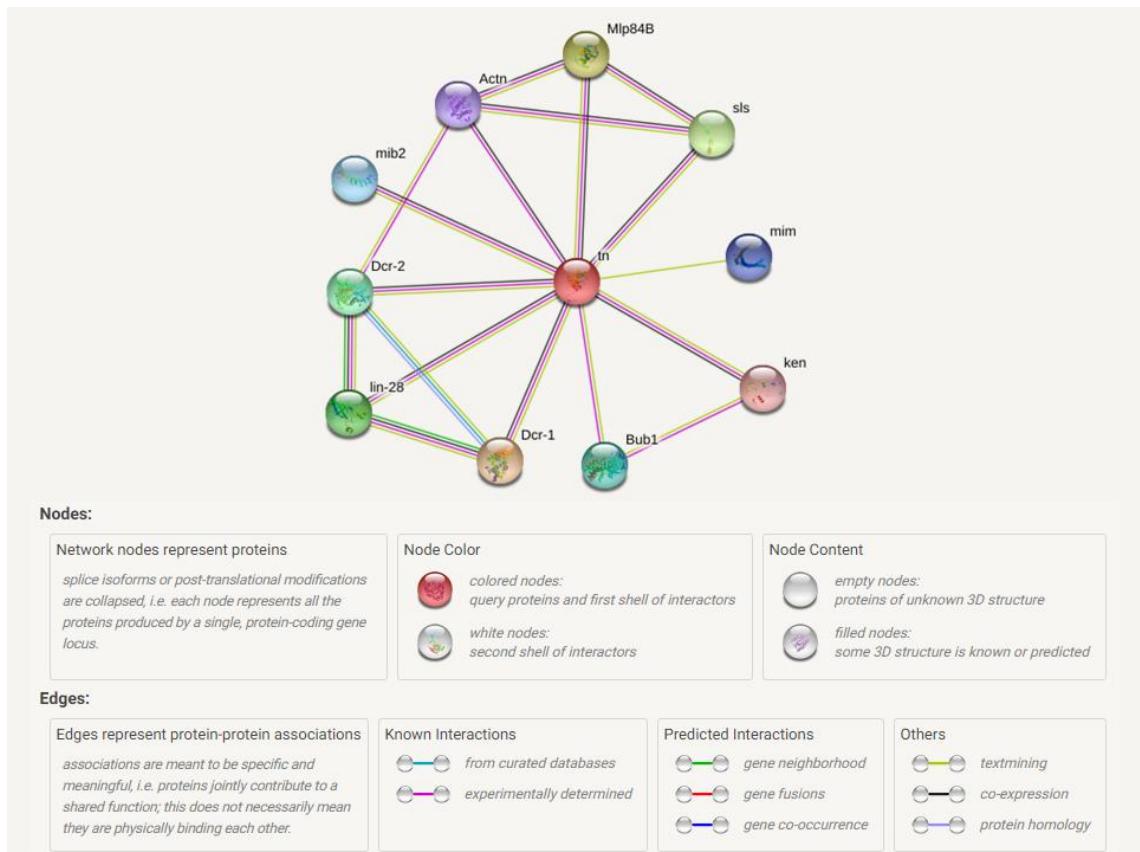
Supplementary Figure 1- Physical interaction network. Software esyN.



Supplementary Figure 2 – Genetic interaction network. Software esyN. Red arrow represents suppression of expression and blue arrow represents increased expression.



Supplementary Figure 3 – Interactions with the thin protein, represented with tn. The red solid line represents the protein-protein interactions, the blue solid line represents the genetic interactions, the blue, green, light purple and dark dotted lines represent the expression correlation interactions, interlog genetics, interlogs, and phenotypic correlations, respectively. Software MIST



Supplementary Figure 4 - Protein-protein associations. Software STRING