



UNIVERSIDADE FEDERAL DE UBERLÂNDIA
INSTITUTO DE BIOTECNOLOGIA
PÓS-GRADUAÇÃO EM GENÉTICA E BIOQUÍMICA



**Sinalização de Anexina A1: Papel das Vias De EGFR E IL-6 nos Cânceres de
Próstata e Mama**

Aluna: Sara Teixeira Soares Mota

Orientador: Dr^a. Thaise Gonçalves de Araújo

Co-Orientador: Dr^a. Lara Vecchi

UBERLÂNDIA – MG
2021



UNIVERSIDADE FEDERAL DE UBERLÂNDIA
INSTITUTO DE BIOTECNOLOGIA
PÓS-GRADUAÇÃO EM GENÉTICA E BIOQUÍMICA



**SINALIZAÇÃO DE ANEXINA A1: PAPEL DAS VIAS DE EGFR E IL-6 NOS
CÂNCERES DE PRÓSTATA E MAMA**

Aluna: Sara Teixeira Soares Mota

Orientador: Dr^a. Thaise Gonçalves de Araújo

Co-Orientador: Dr^a. Lara Vecchi

**Tese apresentada à Universidade
Federal de Uberlândia como parte
dos requisitos para obtenção do
Título de Doutora em Genética e
Bioquímica (Área Genética)**

UBERLÂNDIA – MG

2021



UNIVERSIDADE FEDERAL DE UBERLÂNDIA
 Coordenação do Programa de Pós-Graduação em Genética e Bioquímica
 Av. Pará 1720, Bloco 2E, Sala 244 - Bairro Umuarama, Uberlândia-MG, CEP 38400-902
 Telefone: +55 (34) 3225-8438 - www.ppggb.ibtec.ufu.br - ppggb@ufu.br



ATA DE DEFESA - PÓS-GRADUAÇÃO

Programa de Pós-Graduação em:	Genética e Bioquímica				
Defesa de:	Doutorado Acadêmico - 02/2021 - PPGGB.				
Data:	Quatro de novembro de dois mil e vinte e um	Hora de início:	08:30h	Hora de encerramento:	13:00
Matrícula do Discente:	11723GBI009				
Nome do Discente:	Sara Teixeira Soares Mota				
Título do Trabalho:	Sinalização de Anexina A1: papéis das vias de EGFR e IL-6 nos cânceres de próstata e mama.				
Área de concentração:	Genética				
Linha de pesquisa:	Biologia Molecular e Celular.				
Projeto de Pesquisa de vinculação:	Identificação e caracterização de biomarcadores nos cânceres de próstata e de mama.				

Aos quatro dias do mês de novembro de dois mil e vinte e um, às 08:30 horas, reuniu-se via web conferência pela Plataforma *Microsoft Teams*, em conformidade com a Portaria nº 36, de 19 de março de 2020 da Coordenação de Aperfeiçoamento de Pessoal de Nível Superior – CAPES e Resolução de nº 06/2020 do Conselho de Pesquisa e Pós-graduação pela Universidade Federal de Uberlândia, a Banca Examinadora, designada pelo Colegiado do Programa de Pós-graduação em Genética e Bioquímica, assim composta: Prof^ª. Dr^ª. Tatiana Martins Tilli, Prof^ª. Dr^ª. Adriana Freitas Neves, Prof^ª. Dr^ª. Fernanda de Assis Araújo, Prof^ª. Dr^ª. Vivian Alonso Goulart e Prof^ª. Dr^ª. Thaise Gonçalves de Araújo, orientadora da candidata e demais convidados. Iniciando os trabalhos a presidente da mesa, Prof^ª. Dr^ª. Thaise Gonçalves de Araújo, apresentou a Comissão Examinadora e a candidata, agradeceu a presença do público, e concedeu à Discente a palavra para a exposição do seu trabalho. A duração da apresentação da Discente e o tempo de arguição e resposta foram conforme as normas do Programa de Pós-graduação em Genética e Bioquímica. A seguir a senhora presidente concedeu a palavra, pela ordem sucessivamente, aos examinadores, que passaram a arguir a candidata. Ultimada a arguição, que se desenvolveu dentro dos termos regimentais, a Banca, em sessão secreta, atribuiu os conceitos finais. Em face do resultado obtido, a Banca Examinadora considerou a candidata:

APROVADA.

Esta defesa de Tese de Doutorado é parte dos requisitos necessários à obtenção do título de Doutor. O competente diploma será expedido após cumprimento dos demais requisitos, conforme as normas do Programa, a legislação pertinente e a regulamentação interna da UFU. Nada mais havendo a tratar foram

encerrados os trabalhos. Foi lavrada a presente ata que após lida e achada conforme foi assinada pela Banca Examinadora.



Documento assinado eletronicamente por **Thaise Gonçalves de Araújo, Presidente**, em 04/11/2021, às 13:12, conforme horário oficial de Brasília, com fundamento no art. 6º, § 1º, do [Decreto nº 8.539, de 8 de outubro de 2015](#).



Documento assinado eletronicamente por **Fernanda de Assis Araujo, Professor(a) do Magistério Superior**, em 04/11/2021, às 13:12, conforme horário oficial de Brasília, com fundamento no art. 6º, § 1º, do [Decreto nº 8.539, de 8 de outubro de 2015](#).



Documento assinado eletronicamente por **Vivian Alonso Goulart, Professor(a) do Magistério Superior**, em 04/11/2021, às 13:12, conforme horário oficial de Brasília, com fundamento no art. 6º, § 1º, do [Decreto nº 8.539, de 8 de outubro de 2015](#).



Documento assinado eletronicamente por **Tatiana Martins Tilli, Usuário Externo**, em 04/11/2021, às 13:12, conforme horário oficial de Brasília, com fundamento no art. 6º, § 1º, do [Decreto nº 8.539, de 8 de outubro de 2015](#).



Documento assinado eletronicamente por **ADRIANA FREITAS NEVES, Usuário Externo**, em 04/11/2021, às 13:12, conforme horário oficial de Brasília, com fundamento no art. 6º, § 1º, do [Decreto nº 8.539, de 8 de outubro de 2015](#).



A autenticidade deste documento pode ser conferida no site https://www.sei.ufu.br/sei/controlador_externo.php?acao=documento_conferir&id_orgao_acesso_externo=0, informando o código verificador **3006798** e o código CRC **34750B2A**.

Dados Internacionais de Catalogação na Publicação (CIP)
Sistema de Bibliotecas da UFU, MG, Brasil.

M917s Mota, Sara Teixeira Soares, 1993-
2021 Sinalização de Anexina A1 [recurso eletrônico] : papel das vias de
EGFR e IL-6 nos cânceres de próstata e mama / Sara Teixeira Soares
Mota. - 2021.

Orientadora: Thaise Gonçalves de Araújo.

Coorientadora: Lara Vecchi.

Tese (Doutorado) - Universidade Federal de Uberlândia, Programa
de Pós-Graduação em Genética e Bioquímica.

Disponível em: <http://doi.org/10.14393/ufu.te.2022.5005>

Inclui bibliografia.

1. Genética. I. Araújo, Thaise Gonçalves de, 1984-, (Orient.). II.
Vecchi, Lara, 1981-, (Coorient.). III. Universidade Federal de Uberlândia.
Programa de Pós-Graduação em Genética e Bioquímica. IV. Título.

CDU:575

André Carlos Francisco
Bibliotecário – CRB-6/3408



UNIVERSIDADE FEDERAL DE UBERLÂNDIA
INSTITUTO DE BIOTECNOLOGIA
PÓS-GRADUAÇÃO EM GENÉTICA E BIOQUÍMICA



**SINALIZAÇÃO DE ANEXINA A1: PAPEL DAS VIAS DE EGFR E IL-6 NOS
CÂNCERES DE PRÓSTATA E MAMA**

Aluna: Sara Teixeira Soares Mota

COMISSÃO EXAMINADORA

Presidente: Profa. Dr^a. Thaise Gonçalves de Araújo

Examinadores: Profa. Dr^a. Vivian Alonso Goulart
Profa. Dr^a. Fernanda de Assis Araújo
Profa. Dr^a. Adriana Freitas Neves
Profa. Dr^a. Tatiana Martins Tilli

Data da defesa:

As sugestões da Comissão Examinadora e as Normas PPGGB para o formato da Tese foram contempladas.

Orientador

“Por isso mesmo, empenhem-se para acrescentar à sua fé a virtude; à virtude o conhecimento; ao conhecimento o domínio próprio; ao domínio próprio a perseverança; à perseverança a piedade; à piedade a fraternidade; e à fraternidade o amor.”

2 Pedro 1:5-7.

Dedicatória

A Deus.

A gente sonha Deus realiza!

A todos que me ajudaram ao longo desta caminhada.

Agradecimentos

Muito obrigado, meu Deus, por tudo de bom e de mau que é colocado no meu caminho, pois aquilo que não me traz felicidade me permite reconhecê-la quando ela chega em minha vida.

Aos meus pais, irmãos, marido e todos da minha família, que me incentivaram nos momentos difíceis e compreenderam a minha ausência enquanto eu me dedicava à realização deste trabalho.

Agradeço à minha orientadora, Profª. Drª. Thaise Gonçalves de Araújo, pela sua disponibilidade e incentivo que foram fundamentais para realizar e prosseguir este estudo. Saliento o apoio incondicional prestado, a forma interessada, extraordinária e pertinente como acompanhou a realização deste trabalho. As suas críticas construtivas, as discussões e reflexões foram fundamentais ao longo de todo o percurso. Não posso esquecer a sua grande contribuição para o meu crescimento como pesquisadora, desde os tempos da faculdade. Eternamente grata por todo o apoio.

Agradeço a minha co-orientadora, Drª. Lara Vecchi, obrigada pela confiança no meu trabalho, pelo respeito, por me ensinar, pela paciência, pelos sábios conselhos e por prontamente me ajudar sempre que a procurei. Pela orientação e compreensão. Eu realmente aprendi muito com você.

Aos amigos, que sempre estiveram ao meu lado, pela amizade incondicional e pelo apoio demonstrado ao longo de todo o período em que me dediquei a este trabalho.

Agradeço as técnicas Luciana, Valéria e Natássia, e ao Prof. Dr. Luiz Ricardo Goulart, e todos do Laboratório de Nanobiotecnologia pelo suporte técnico e ensinamentos e

paciência e pelas boas conversas e risadas. Me sinto orgulhosa e privilegiada por fazer parte dessa equipe.

Agradeço aos membros da banca examinadora, pelo interesse e disponibilidade.

Muito obrigada aos membros do Programa de Pós-graduação em Genética e Bioquímica pelo suporte, em especial aos professores do programa. A secretária Janaína, pelo pronto atendimento sempre que solicitada.

A CNPq, CAPES e FAPEMIG, pelo apoio financeiro durante todo o meu doutorado.

Dizer obrigada, às vezes, não é suficiente para agradecer a tão amável e gentil pessoa que nos momentos das nossas vidas, aqueles mais difíceis, nos estende a mão amiga e nos oferece amparo.

Muito obrigada a todos que fazem ou fizeram parte da minha trajetória. Essa realização é nossa!

SUMÁRIO

APRESENTAÇÃO.....	11
-------------------	----

CAPÍTULO I

Annexin A1 as a Regulator of Immune Response in Cancer.....	01
Abstract.....	01
1. Introduction.....	01
2. Anticancer Immune Response and Cancer Immune Escape.....	01
3. The Immunosuppressive Properties of the TME.....	03
4. Structure and Functions of Annexin A1.....	06
4.1. AnxA1 in Cancer.....	08
4.2. Immunosuppressive Functions of AnxA1 in Physiological and Cancer Contexts.....	10
5. Concluding Remarks.....	13
Abbreviations.....	14
References.....	15

CAPÍTULO II

Annexin A1 promotes the nuclear localization of the epidermal growth factor receptor in castration-resistant prostate cancer.....	01
Abstract.....	01
1. Introduction.....	01
2. Materials and methods.....	03
3. Results.....	04
3.1. Correlation between EGFR and AnxA1 expression in PCa sample.....	04
3.2. CRPC cell lines express high levels of cytoplasmic and nuclear EGFR and AnxA1.....	06
3.3. AnxA1 knockdown inhibits the EGFR translocation to nuclei.....	08

3.4. AnxA1 knockdown inhibits the internalization of EGFR and its downstream signaling.....	09
3.5. AnxA1 elicits an autocrine signaling through FPR1.....	09
3.6. AnxA1 autocrine signaling is responsible for EGFR nuclear localization.....	09
3.7. AnxA1 autocrine signaling through FPR1 is essential for EGFR internalization.....	09
4. Discussion.....	14
5. Conclusions...	16
References.....	17

CAPÍTULO III

Interleukin-6 signaling in triple negative breast cancer cells elicits the Annexin A1/Formyl peptide receptor 1 axis and affects the tumor microenvironment.....	01
Abstract.....	01
1. Introduction.....	01
2. Materials and Methods.....	03
3. Results.....	04
3.1. Correlation between AnxA1 and IL-6 expression in BC samples.....	04
3.2. TNBC cell lines express high levels of AnxA1 and IL-6.....	09
3.3. AnxA1 autocrine signaling leads to a decrease of IL-6 expression.....	13
3.4. IL-6 inhibition is involved in tumor growth, metastasis and tumor microenvironment.....	16
4. Discussion.....	18
5. Conclusions.....	20
References.....	21

APRESENTAÇÃO

A Anexina A1 (AnxA1) é uma proteína de 37 KDa que se liga a fosfolipídios de membrana de maneira cálcio-dependente, sendo responsável por regular a inflamação, a organização de membranas celulares e o tráfego vesicular. AnxA1 é expressa principalmente no citoplasma, onde apresenta uma atividade anti-inflamatória. Já no núcleo está envolvida em regular a transcrição e, portanto, o funcionamento gênico. AnxA1 também pode ser clivada proteolicamente gerando um fragmento de 33-KDa e liberando seu peptídeo clivado N-terminal, que é biologicamente ativo. Tanto o peptídeo quanto a proteína de comprimento total podem ser secretados para o meio extracelular, onde são capazes de ativar Receptores de Peptídeos Formilados (FPRs), desencadeando, assim uma sinalização autócrina.

O eixo de sinalização autócrina AnxA1/FPR1 está envolvida nas propriedades pró-invasivas e pró-tumorais, principalmente de tumores em estágios mais avançados, como o câncer de mama triplo negativo (CMTN) e o câncer de próstata resistente à castração (CRPC). AnxA1 desempenha papéis relevantes na transformação maligna, ativando oncogenes e inativando genes supressores de tumor, induzindo a proliferação e invasão celulares e a formação de metástases. Por isso, investigar a via sinalização da AnxA1, bem com a interação com outras moléculas nos diversos tipos de tumores, se mostra imprescindível para elucidar as particularidades dessa proteína e, assim, desenvolver novas estratégias de diagnóstico e terapêuticas para as diferentes neoplasias malignas.

Em tumores, a Anxa1 desempenha atividade pró-inflamatória, induzindo um microambiente tumoral (TME) favorável com a promoção do crescimento e circulação de células malignas. Portanto, no Capítulo I, revisamos o papel da AnxA1 no estabelecimento do microambiente tumoral, com foco nas atividades imunossupressora e imunomoduladora. Primeiramente, descrevemos como células cancerosas suprimem as atividades do sistema imunológico, favorecendo a malignidade e a disseminação do câncer. Em seguida, apresentamos as principais características do TME e que a presença de algumas células imunológicas em sua composição cria um ambiente altamente imunossupressor, o que favorece o crescimento e a progressão do tumor. Retratamos a estrutura e características da

AnxA1, bem como os seus papéis na transformação maligna. Por fim, evidenciamos como a AnxA1 promove a supressão tumoral pela interação com o receptor do fator de crescimento epidérmico (EGFR), desencadeando um mecanismo de escape imunológico pelas células do câncer.

No segundo capítulo, mostramos que a AnxA1 ativa uma sinalização autócrina em células CRPC por meio do FPR1. Além disso, no CRPC, observamos que a sinalização autócrina de AnxA1 promove a localização nuclear no EGFR. Já foi descrito previamente que, uma vez no núcleo, EGFR é associado à progressão do câncer de próstata para o fenótipo resistente à castração. Quando inibimos o FPR1 utilizando Ciclosoprina H, a localização nuclear do EGFR e sua sinalização a jusante nas linhagens CRPC foram bloqueadas. Sugerimos, então, que a inibição da sinalização autócrina de AnxA1 e, conseqüentemente, da localização nuclear de EGFR pode ser uma estratégia relevante para o tratamento de CRPC

No último Capítulo, demonstramos uma interação entre as vias de sinalização de AnxA1/FPR1 e da Interleucina-6 (IL-6) e STAT3 (Fator de transcrição ativado por tirosinas quinases 3) no CMTN. Os nossos resultados mostraram que nas linhagens de CMTN, MDA-MB-231 e MDA-MB-157, a cascata de sinalização de IL-6 modula a ativação do eixo autócrino AnxA1/FPR1. Ao tratar camundongos fêmeas atímicas transplantadas com células MDA-MB-231 com Tocilizumabe, um anticorpo monoclonal humanizado que atua bloqueando os receptores de IL-6, observamos a redução do crescimento do tumor e a inibição da formação de metástases. Por último, mostramos que as vias IL-6 /STAT3 e AnxA1/FPR1 influenciam o TME, alterando a motilidade de fibroblastos. Sugerimos que a inibição de IL-6 pode ser usada como uma possível intervenção terapêutica e/ou uma nova terapia adjuvante para melhorar o resultado clínico de pacientes com CMTN ao bloquear a sinalização autócrina de AnxA1, retardando a progressão tumoral.

Por fim, demonstramos que a AnxA1 estimula um microambiente tumoral favorável o que leva o escape imunológico pelas células do câncer. E em tumores de próstata mais agressivos, como CRPC, a via de sinalização autócrina da AnxA1 e FPR1 promove a localização nuclear de EGFR, e que esta localização desempenha um papel importante na sobrevivência celular e agressividade das células CRPC. Já em tumores de mama triplo negativos a via de IL-6 mostrou-se um mecanismo compensatório da sinalização de do eixo AnxA1 e FPR1 e essas

vias além de apresentarem um papel importante na agressividade do CMTN, também influenciam a TME, reduzindo a motilidade de fibroblastos. Portanto, este trabalho contribuiu de forma eficaz para a comunidade científica demonstrando que o Anexina A1 desempenha diferentes papéis na progressão tumoral e que estudos mais aprofundados sobre essa molécula, e, em especial na via de sinalização autócrina AnxA1/FPR1, mostram se relevantes para uma maior compreensão dos tumores mais agressivos, bem como, possíveis estratégias terapêuticas.

Palavras-chave: Anexina A1, câncer de mama, câncer de próstata, microambiente tumoral, EGFR, IL-6, STAT3, ciclosporina e Tocilizumab.

Capítulo 1

Annexin A1 as a regulator of immune response in cancer

Annexin A1 as a Regulator of Immune Response in Cancer

Thaise Gonçalves Araújo ^{1,2} , Sara Teixeira Soares Mota ^{1,2}, Helen Soares Valença Ferreira ¹,
Matheus Alves Ribeiro ¹, Luiz Ricardo Goulart ²  and Lara Vecchi ^{2,*} 

¹ Laboratory of Genetics and Biotechnology, Federal University of Uberlândia, Patos de Minas 387400-128, MG, Brazil; tgaraujo@ufu.br (T.G.A.); saratsm.s@hotmail.com (S.T.S.M.); helensvalenca@gmail.com (H.S.V.F.); matheusribeiromarketing@gmail.com (M.A.R.)

² Laboratory of Nanobiotechnology, Federal University of Uberlândia, Uberlândia 38400-902, MG, Brazil; lrgoulart@ufu.br

* Correspondence: laravecchi7@yahoo.it

Abstract: Annexin A1 is a 37 kDa phospholipid-binding protein that is expressed in many tissues and cell types, including leukocytes, lymphocytes and epithelial cells. Although Annexin A1 has been extensively studied for its anti-inflammatory activity, it has been shown that, in the cancer context, its activity switches from anti-inflammatory to pro-inflammatory. Remarkably, Annexin A1 shows pro-invasive and pro-tumoral properties in several cancers either by eliciting autocrine signaling in cancer cells or by inducing a favorable tumor microenvironment. Indeed, the signaling of the N-terminal peptide of AnxA1 has been described to promote the switching of macrophages to the pro-tumoral M2 phenotype. Moreover, AnxA1 has been described to prevent the induction of antigen-specific cytotoxic T cell response and to play an essential role in the induction of regulatory T lymphocytes. In this way, Annexin A1 inhibits the anti-tumor immunity and supports the formation of an immunosuppressed tumor microenvironment that promotes tumor growth and metastasis. For these reasons, in this review we aim to describe the role of Annexin A1 in the establishment of the tumor microenvironment, focusing on the immunosuppressive and immunomodulatory activities of Annexin A1 and on its interaction with the epidermal growth factor receptor.

Keywords: Annexin A1; tumor microenvironment; immune-suppression; cancer aggressiveness



Citation: Araújo, T.G.; Mota, S.T.S.; Ferreira, H.S.V.; Ribeiro, M.A.; Goulart, L.R.; Vecchi, L. Annexin A1 as a Regulator of Immune Response in Cancer. *Cells* **2021**, *10*, 2245.

<https://doi.org/10.3390/cells10092245>

Academic Editor: Antonello Petrella

Received: 30 June 2021

Accepted: 7 August 2021

Published: 30 August 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Neoplastic transformation and progression encompass multiple unknown or poorly described events. In this context, Hanahan and Weinberg proposed different hallmarks that, despite being interrelated, define the principles of tumorigenesis. These hallmarks include the resistance to cell death mechanisms, uncontrolled proliferation, induction of angiogenesis, invasion potential, metastasis formation and immune escape [1]. Cancer cells acquire these hallmarks by modifying the extracellular matrix (ECM) and modulating the behavior of different cell types that surround the tumor. In this way, cancer cells create a unique environment, called the tumor microenvironment (TME), that strongly favors malignancy and cancer dissemination [2]. Unfortunately, the appearance of metastasis profoundly worsens the prognoses of patients, since no efficient therapies are available [3]. It is therefore essential to study the factors that are involved in the establishment of the TME in order to design new therapeutic strategies that could help to avoid cancer spreading and manage metastatic cancer patients. Of particular interest is the understanding of how cancer cells create a TME that favors immune evasion. In this context, it is important to unravel the immunomodulatory role of the protein Annexin A1 (AnxA1), which is the purpose of this review.

2. Anticancer Immune Response and Cancer Immune Escape

The immune system recognizes antigens expressed on the surface of transformed cells, leading to their destruction, thus hampering tumor progression. Remarkably, both

Treg	regulatory T cell	1
Trp	tryptophane	2
VCAM 1	vascular cell adhesion molecule 1	3
VEGF	vascular endothelial growth factor	4

References

1. Hanahan, D., and R. A. Weinberg. "Hallmarks of Cancer: The Next Generation." *Cell* 144, no. 5 (2011): 646-74. <https://doi.org/10.1016/j.cell.2011.02.013>.
2. Brassart-Pasco, S., S. Brezillon, B. Brassart, L. Ramont, J. B. Oudart, and J. C. Monboisse. "Tumor Microenvironment: Extracellular Matrix Alterations Influence Tumor Progression." *Front Oncol* 10 (2020): 397. <https://doi.org/10.3389/fonc.2020.00397>.
3. Chaffer, C. L., and R. A. Weinberg. "A Perspective on Cancer Cell Metastasis." *Science* 331, no. 6024 (2011): 1559-64. <https://doi.org/10.1126/science.1203543>.
4. Blankenstein, T., P. G. Coulie, E. Gilboa, and E. M. Jaffee. "The Determinants of Tumour Immunogenicity." *Nat Rev Cancer* 12, no. 4 (2012): 307-13. <https://doi.org/10.1038/nrc3246>.
5. Pradeu, T., and E. D. Carosella. "On the Definition of a Criterion of Immunogenicity." *Proc Natl Acad Sci U S A* 103, no. 47 (2006): 17858-61. <https://doi.org/10.1073/pnas.0608683103>.
6. Kitamura, T., B. Z. Qian, and J. W. Pollard. "Immune Cell Promotion of Metastasis." *Nat Rev Immunol* 15, no. 2 (2015): 73-86. <https://doi.org/10.1038/nri3789>.
7. Crusz, S. M., and F. R. Balkwill. "Inflammation and Cancer: Advances and New Agents." *Nat Rev Clin Oncol* 12, no. 10 (2015): 584-96. <https://doi.org/10.1038/nrclinonc.2015.105>.
8. Colotta, F., P. Allavena, A. Sica, C. Garlanda, and A. Mantovani. "Cancer-Related Inflammation, the Seventh Hallmark of Cancer: Links to Genetic Instability." *Carcinogenesis* 30, no. 7 (2009): 1073-81. <https://doi.org/10.1093/carcin/bgp127>.
9. Gonzalez, H., C. Hagerling, and Z. Werb. "Roles of the Immune System in Cancer: From Tumor Initiation to Metastatic Progression." *Genes Dev* 32, no. 19-20 (2018): 1267-84. <https://doi.org/10.1101/gad.314617.118>.
10. Kim, R., M. Emi, and K. Tanabe. "Cancer Cell Immune Escape and Tumor Progression by Exploitation of Anti-Inflammatory and Pro-Inflammatory Responses." *Cancer Biol Ther* 4, no. 9 (2005): 924-33. <https://doi.org/10.4161/cbt.4.9.2101>.
11. Gaudino, S. J., and P. Kumar. "Cross-Talk between Antigen Presenting Cells and T Cells Impacts Intestinal Homeostasis, Bacterial Infections, and Tumorigenesis." *Front Immunol* 10 (2019): 360. <https://doi.org/10.3389/fimmu.2019.00360>.
12. Waldman, A. D., J. M. Fritz, and M. J. Lenardo. "A Guide to Cancer Immunotherapy: From T Cell Basic Science to Clinical Practice." *Nat Rev Immunol* 20, no. 11 (2020): 651-68. <https://doi.org/10.1038/s41577-020-0306-5>.
13. Matsuzaki, J., T. Tsuji, I. Luescher, L. J. Old, P. Shrikant, S. Gnjjatic, and K. Odunsi. "Nonclassical Antigen-Processing Pathways Are Required for Mhc Class Ii-Restricted Direct Tumor Recognition by Ny-Eso-1-Specific Cd4(+) T Cells." *Cancer Immunol Res* 2, no. 4 (2014): 341-50. <https://doi.org/10.1158/2326-6066.CIR-13-0138>.
14. Knutson, K. L., M. L. Disis, and L. G. Salazar. "Cd4 Regulatory T Cells in Human Cancer Pathogenesis." *Cancer Immunol Immunother* 56, no. 3 (2007): 271-85. <https://doi.org/10.1007/s00262-006-0194-y>.
15. Dardalhon, V., T. Korn, V. K. Kuchroo, and A. C. Anderson. "Role of Th1 and Th17 Cells in Organ-Specific Autoimmunity." *J Autoimmun* 31, no. 3 (2008): 252-6. <https://doi.org/10.1016/j.jaut.2008.04.017>.

16. Bretscher, Peter. "On Analyzing How the Th1/Th2 Phenotype of an Immune Response Is Determined: Classical Observations Must Not Be Ignored." 10, no. 1234 (2019). <https://doi.org/10.3389/fimmu.2019.01234>.
17. Emens, L. A. "Breast Cancer Immunobiology Driving Immunotherapy: Vaccines and Immune Checkpoint Blockade." *Expert Rev Anticancer Ther* 12, no. 12 (2012): 1597-611. <https://doi.org/10.1586/era.12.147>.
18. Burkholder, B., R. Y. Huang, R. Burgess, S. Luo, V. S. Jones, W. Zhang, Z. Q. Lv, C. Y. Gao, B. L. Wang, Y. M. Zhang, and R. P. Huang. "Tumor-Induced Perturbations of Cytokines and Immune Cell Networks." *Biochim Biophys Acta* 1845, no. 2 (2014): 182-201. <https://doi.org/10.1016/j.bbcan.2014.01.004>.
19. Zambrano-Zaragoza, J. F., E. J. Romo-Martinez, J. Duran-Avelar Mde, N. Garcia-Magallanes, and N. Vibanco-Perez. "Th17 Cells in Autoimmune and Infectious Diseases." *Int J Inflam* 2014 (2014): 651503. <https://doi.org/10.1155/2014/651503>.
20. Kryczek, I., S. Wei, W. Szeliga, L. Vatan, and W. Zou. "Endogenous IL-17 Contributes to Reduced Tumor Growth and Metastasis." *Blood* 114, no. 2 (2009): 357-9. <https://doi.org/10.1182/blood-2008-09-177360>.
21. Zou, W., and N. P. Restifo. "T(H)17 Cells in Tumour Immunity and Immunotherapy." *Nat Rev Immunol* 10, no. 4 (2010): 248-56. <https://doi.org/10.1038/nri2742>.
22. Guery, L., and S. Hugues. "Th17 Cell Plasticity and Functions in Cancer Immunity." *Biomed Res Int* 2015 (2015): 314620. <https://doi.org/10.1155/2015/314620>.
23. Cvetanovich, G. L., and D. A. Hafler. "Human Regulatory T Cells in Autoimmune Diseases." *Curr Opin Immunol* 22, no. 6 (2010): 753-60. <https://doi.org/10.1016/j.coi.2010.08.012>.
24. Martin-Orozco, E., M. Norte-Munoz, and J. Martinez-Garcia. "Regulatory T Cells in Allergy and Asthma." *Front Pediatr* 5 (2017): 117. <https://doi.org/10.3389/fped.2017.00117>.
25. Wu, H., P. Li, N. Shao, J. Ma, M. Ji, X. Sun, D. Ma, and C. Ji. "Aberrant Expression of Treg-Associated Cytokine IL-35 Along with IL-10 and Tgf-Beta in Acute Myeloid Leukemia." *Oncol Lett* 3, no. 5 (2012): 1119-23. <https://doi.org/10.3892/ol.2012.614>.
26. Ohue, Y., and H. Nishikawa. "Regulatory T (Treg) Cells in Cancer: Can Treg Cells Be a New Therapeutic Target?" *Cancer Sci* 110, no. 7 (2019): 2080-89. <https://doi.org/10.1111/cas.14069>.
27. Andersen, M. H., D. Schrama, P. Thor Straten, and J. C. Becker. "Cytotoxic T Cells." *J Invest Dermatol* 126, no. 1 (2006): 32-41. <https://doi.org/10.1038/sj.jid.5700001>.
28. Mills, C. D. "M1 and M2 Macrophages: Oracles of Health and Disease." *Crit Rev Immunol* 32, no. 6 (2012): 463-88. <https://doi.org/10.1615/CritRevImmunol.v32.i6.10>.
29. Cai, J., W. Zhang, P. Yang, Y. Wang, M. Li, C. Zhang, Z. Wang, H. Hu, Y. Liu, Q. Li, J. Wen, B. Sun, X. Wang, T. Jiang, and C. Jiang. "Identification of a 6-Cytokine Prognostic Signature in Patients with Primary Glioblastoma Harboring M2 Microglia/Macrophage Phenotype Relevance." *PLoS One* 10, no. 5 (2015): e0126022. <https://doi.org/10.1371/journal.pone.0126022>.
30. Qi, L., H. Yu, Y. Zhang, D. Zhao, P. Lv, Y. Zhong, and Y. Xu. "IL-10 Secreted by M2 Macrophage Promoted Tumorigenesis through Interaction with Jak2 in Glioma." *Oncotarget* 7, no. 44 (2016): 71673-85. <https://doi.org/10.18632/oncotarget.12317>.
31. Arlauckas, S. P., S. B. Garren, C. S. Garriss, R. H. Kohler, J. Oh, M. J. Pittet, and R. Weissleder. "Arg1 Expression Defines Immunosuppressive Subsets of Tumor-Associated Macrophages." *Theranostics* 8, no. 21 (2018): 5842-54. <https://doi.org/10.7150/thno.26888>.
32. Hao, N. B., M. H. Lu, Y. H. Fan, Y. L. Cao, Z. R. Zhang, and S. M. Yang. "Macrophages in Tumor Microenvironments and the Progression of Tumors." *Clin Dev Immunol* 2012 (2012): 948098. <https://doi.org/10.1155/2012/948098>.

33. Solinas, G., G. Germano, A. Mantovani, and P. Allavena. "Tumor-Associated Macrophages (Tam) as Major Players of the Cancer-Related Inflammation." *J Leukoc Biol* 86, no. 5 (2009): 1065-73. <https://doi.org/10.1189/jlb.0609385>.
34. Leek, R. D., C. E. Lewis, R. Whitehouse, M. Greenall, J. Clarke, and A. L. Harris. "Association of Macrophage Infiltration with Angiogenesis and Prognosis in Invasive Breast Carcinoma." *Cancer Res* 56, no. 20 (1996): 4625-9.
35. Cruvinel Wde, M., D. Mesquita, Jr., J. A. Araujo, T. T. Catelan, A. W. de Souza, N. P. da Silva, and L. E. Andrade. "Immune System - Part I. Fundamentals of Innate Immunity with Emphasis on Molecular and Cellular Mechanisms of Inflammatory Response." *Rev Bras Reumatol* 50, no. 4 (2010): 434-61. <https://doi.org/10.1590/S0482-50042010000400008>.
36. Hubo, M., B. Trinschek, F. Kryczanowsky, A. Tuettenberg, K. Steinbrink, and H. Jonuleit. "Costimulatory Molecules on Immunogenic Versus Tolerogenic Human Dendritic Cells." *Front Immunol* 4 (2013): 82. <https://doi.org/10.3389/fimmu.2013.00082>.
37. Domogalla, M. P., P. V. Rostan, V. K. Raker, and K. Steinbrink. "Tolerance through Education: How Tolerogenic Dendritic Cells Shape Immunity." *Front Immunol* 8 (2017): 1764. <https://doi.org/10.3389/fimmu.2017.01764>.
38. Bharadwaj, U., M. Li, C. Chen, and Q. Yao. "Mesothelin-Induced Pancreatic Cancer Cell Proliferation Involves Alteration of Cyclin E Via Activation of Signal Transducer and Activator of Transcription Protein 3." *Mol Cancer Res* 6, no. 11 (2008): 1755-65. <https://doi.org/10.1158/1541-7786.MCR-08-0095>.
39. Jarnicki, A. G., J. Lysaght, S. Todryk, and K. H. Mills. "Suppression of Antitumor Immunity by Il-10 and Tgf-Beta-Producing T Cells Infiltrating the Growing Tumor: Influence of Tumor Environment on the Induction of Cd4+ and Cd8+ Regulatory T Cells." *J Immunol* 177, no. 2 (2006): 896-904. <https://doi.org/10.1158/1541-7786.MCR-08-0095>.
40. Wiguna, A. P., and P. Walden. "Role of Il-10 and Tgf-Beta in Melanoma." *Exp Dermatol* 24, no. 3 (2015): 209-14. <https://doi.org/10.1111/exd.12629>.
41. Bonetti, M. I., L. Pieri, L. Domenici, S. Urbani, G. Romano, A. Aldinucci, C. Ballerini, M. Monici, R. Saccardi, V. Basile, A. Bosi, and P. Romagnoli. "Dendritic Cells with Lymphocyte-Stimulating Activity Differentiate from Human Cd133 Positive Precursors." *Blood* 117, no. 15 (2011): 3983-95. <https://doi.org/10.1182/blood-2010-08-299735>.
42. Saibil, S. D., E. K. Deenick, and P. S. Ohashi. "The Sound of Silence: Modulating Anergy in T Lymphocytes." *Curr Opin Immunol* 19, no. 6 (2007): 658-64. <https://doi.org/10.1016/j.coi.2007.08.005>.
43. Candolfi, M., K. Yagiz, D. Foulad, G. E. Alzadeh, M. Tesarfreund, A. K. Muhammad, M. Puntel, K. M. Kroeger, C. Liu, S. Lee, J. F. Curtin, G. D. King, J. Lerner, K. Sato, Y. Mineharu, W. Xiong, P. R. Lowenstein, and M. G. Castro. "Release of Hmgb1 in Response to Proapoptotic Glioma Killing Strategies: Efficacy and Neurotoxicity." *Clin Cancer Res* 15, no. 13 (2009): 4401-14. <https://doi.org/10.1158/1078-0432.CCR-09-0155>.
44. Olkhanud, P. B., B. Damdinsuren, M. Bodogai, R. E. Gress, R. Sen, K. Wejsza, E. Malchinkhuu, R. P. Wersto, and A. Biragyn. "Tumor-Evoked Regulatory B Cells Promote Breast Cancer Metastasis by Converting Resting Cd4(+) T Cells to T-Regulatory Cells." *Cancer Res* 71, no. 10 (2011): 3505-15. <https://doi.org/10.1158/0008-5472.CAN-10-4316>.
45. Matsushita, T. "Regulatory and Effector B Cells: Friends or Foes?" *J Dermatol Sci* 93, no. 1 (2019): 2-7. <https://doi.org/10.1016/j.jdermsci.2018.11.008>.

46. Veglia, F., M. Perego, and D. Gabrilovich. "Myeloid-Derived Suppressor Cells Coming of Age." *Nat Immunol* 19, no. 2 (2018): 108-19. <https://doi.org/10.1038/s41590-017-0022-x>.
47. Sendo, S., J. Saegusa, and A. Morinobu. "Myeloid-Derived Suppressor Cells in Non-Neoplastic Inflamed Organs." *Inflamm Regen* 38 (2018): 19. <https://doi.org/10.1186/s41232-018-0076-7>.
48. Beury, D. W., K. A. Carter, C. Nelson, P. Sinha, E. Hanson, M. Nyandjo, P. J. Fitzgerald, A. Majeed, N. Wali, and S. Ostrand-Rosenberg. "Myeloid-Derived Suppressor Cell Survival and Function Are Regulated by the Transcription Factor Nrf2." *J Immunol* 196, no. 8 (2016): 3470-8. <https://doi.org/10.4049/jimmunol.1501785>.
49. Anani, Waseem, and Michael R. Shurin. "Targeting Myeloid-Derived Suppressor Cells in Cancer." In *Tumor Immune Microenvironment in Cancer Progression and Cancer Therapy*, edited by Pawel Kalinski, 105-28. Cham: Springer International Publishing, 2017. https://doi.org/10.1007/978-3-319-67577-0_8.
50. Purdy, A. K., and K. S. Campbell. "Natural Killer Cells and Cancer: Regulation by the Killer Cell Ig-Like Receptors (Kir)." *Cancer Biol Ther* 8, no. 23 (2009): 2211-20. <https://doi.org/10.4161/cbt.8.23.10455>.
51. Jost, S., and M. Altfeld. "Control of Human Viral Infections by Natural Killer Cells." *Annu Rev Immunol* 31 (2013): 163-94. <https://doi.org/10.1146/annurev-immunol-032712-100001>.
52. Vivier, E., D. H. Raulet, A. Moretta, M. A. Caligiuri, L. Zitvogel, L. L. Lanier, W. M. Yokoyama, and S. Ugolini. "Innate or Adaptive Immunity? The Example of Natural Killer Cells." *Science* 331, no. 6013 (2011): 44-9. <https://doi.org/10.1126/science.1198687>.
53. Morvan, M. G., and L. L. Lanier. "Nk Cells and Cancer: You Can Teach Innate Cells New Tricks." *Nat Rev Cancer* 16, no. 1 (2016): 7-19. <https://doi.org/10.1038/nrc.2015.5>.
54. Zhang, Z., S. Liu, B. Zhang, L. Qiao, Y. Zhang, and Y. Zhang. "T Cell Dysfunction and Exhaustion in Cancer." *Front Cell Dev Biol* 8 (2020): 17. <https://doi.org/10.3389/fcell.2020.00017>.
55. DeVito, N. C., M. P. Plebanek, B. Theivanthiran, and B. A. Hanks. "Role of Tumor-Mediated Dendritic Cell Tolerization in Immune Evasion." *Front Immunol* 10 (2019): 2876. <https://doi.org/10.3389/fimmu.2019.02876>.
56. Correia, A. L., and M. J. Bissell. "The Tumor Microenvironment Is a Dominant Force in Multidrug Resistance." *Drug Resist Updat* 15, no. 1-2 (2012): 39-49. <https://doi.org/10.1016/j.drug.2012.01.006>.
57. Yuan, Y., Y. C. Jiang, C. K. Sun, and Q. M. Chen. "Role of the Tumor Microenvironment in Tumor Progression and the Clinical Applications (Review)." *Oncol Rep* 35, no. 5 (2016): 2499-515. <https://doi.org/10.3892/or.2016.4660>.
58. Spill, F., D. S. Reynolds, R. D. Kamm, and M. H. Zaman. "Impact of the Physical Microenvironment on Tumor Progression and Metastasis." *Curr Opin Biotechnol* 40 (2016): 41-48. <https://doi.org/10.1016/j.copbio.2016.02.007>.
59. Reina-Campos, M., J. Moscat, and M. Diaz-Meco. "Metabolism Shapes the Tumor Microenvironment." *Curr Opin Cell Biol* 48 (2017): 47-53. <https://doi.org/10.1016/j.ceb.2017.05.006>.
60. Kashyap, A. S., M. Schmittnaegel, N. Rigamonti, D. Pais-Ferreira, P. Mueller, M. Buchi, C. H. Ooi, M. Kreuzaler, P. Hirschmann, A. Guichard, N. Rieder, R. Bill, F. Herting, Y. Kienast, S. Dirnhofer, C. Klein, S. Hoves, C. H. Ries, E. Corse, M. De Palma, and A. Zippelius. "Optimized Antiangiogenic Reprogramming of the Tumor Microenvironment Potentiates Cd40 Immunotherapy." *Proc Natl Acad Sci U S A* 117, no. 1 (2020): 541-51. <https://doi.org/10.1073/pnas.1902145116>.
61. Anderson, N. M., and M. C. Simon. "The Tumor Microenvironment." *Curr Biol* 30, no. 16 (2020): R921-R25. <https://doi.org/10.1016/j.cub.2020.06.081>.
62. Wang, M., J. Zhao, L. Zhang, F. Wei, Y. Lian, Y. Wu, Z. Gong, S. Zhang, J. Zhou, K. Cao, X. Li, W. Xiong, G. Li, Z. Zeng, and C. Guo. "Role of Tumor Microenvironment in Tumorigenesis." *J Cancer* 8, no. 5 (2017): 761-73. <https://doi.org/10.7150/jca.17648>.

63. Huang, W., S. Luo, R. Burgess, Y. H. Yi, G. F. Huang, and R. P. Huang. "New Insights into the Tumor Microenvironment Utilizing Protein Array Technology." *Int J Mol Sci* 19, no. 2 (2018). <https://doi.org/10.3390/ijms19020559>.
64. Frantz, C., K. M. Stewart, and V. M. Weaver. "The Extracellular Matrix at a Glance." *J Cell Sci* 123, no. Pt 24 (2010): 4195-200. <https://doi.org/10.1242/jcs.023820>.
65. Theocharis, A. D., S. S. Skandalis, C. Gialeli, and N. K. Karamanos. "Extracellular Matrix Structure." *Adv Drug Deliv Rev* 97 (2016): 4-27. <https://doi.org/10.1016/j.addr.2015.11.001>.
66. Deryugina, E. I., and J. P. Quigley. "Tumor Angiogenesis: Mmp-Mediated Induction of Intravasation- and Metastasis-Sustaining Neovasculature." *Matrix Biol* 44-46 (2015): 94-112. <https://doi.org/10.1016/j.matbio.2015.04.004>.
67. Shiga, K., M. Hara, T. Nagasaki, T. Sato, H. Takahashi, and H. Takeyama. "Cancer-Associated Fibroblasts: Their Characteristics and Their Roles in Tumor Growth." *Cancers (Basel)* 7, no. 4 (2015): 2443-58. <https://doi.org/10.3390/cancers7040902>.
68. Wang, X., W. Zhang, X. Sun, Y. Lin, and W. Chen. "Cancer-Associated Fibroblasts Induce Epithelial-Mesenchymal Transition through Secreted Cytokines in Endometrial Cancer Cells." *Oncol Lett* 15, no. 4 (2018): 5694-702. <https://doi.org/10.3892/ol.2018.8000>.
69. Lei, X., Y. Lei, J. K. Li, W. X. Du, R. G. Li, J. Yang, J. Li, F. Li, and H. B. Tan. "Immune Cells within the Tumor Microenvironment: Biological Functions and Roles in Cancer Immunotherapy." *Cancer Lett* 470 (2020): 126-33. <https://doi.org/10.1016/j.canlet.2019.11.009>.
70. Mayer, A., M. Haist, C. Loquai, S. Grabbe, M. Rapp, W. Roth, P. Vaupel, and H. Schmidberger. "Role of Hypoxia and the Adenosine System in Immune Evasion and Prognosis of Patients with Brain Metastases of Melanoma: A Multiplex Whole Slide Immunofluorescence Study." *Cancers (Basel)* 12, no. 12 (2020). <https://doi.org/10.3390/cancers12123753>.
71. Nachmany, I., Y. Bogoch, G. Friedlander-Malik, O. Amar, E. Bondar, N. Zohar, S. Hantisteanu, O. Fainaru, N. Lubezky, J. M. Klausner, and N. Pencovich. "The Transcriptional Profile of Circulating Myeloid Derived Suppressor Cells Correlates with Tumor Development and Progression in Mouse." *Genes Immun* 20, no. 7 (2019): 589-98. <https://doi.org/10.1038/s41435-019-0062-3>.
72. Magnuson, A. M., E. Kiner, A. Ergun, J. S. Park, N. Asinovski, A. Ortiz-Lopez, A. Kilcoyne, E. Paoluzzi-Tomada, R. Weissleder, D. Mathis, and C. Benoist. "Identification and Validation of a Tumor-Infiltrating Treg Transcriptional Signature Conserved across Species and Tumor Types." *Proc Natl Acad Sci U S A* 115, no. 45 (2018): E10672-E81. <https://doi.org/10.1073/pnas.1810580115>.
73. Gabrilovich, D. "Mechanisms and Functional Significance of Tumour-Induced Dendritic-Cell Defects." *Nat Rev Immunol* 4, no. 12 (2004): 941-52. <https://doi.org/10.1038/nri1498>.
74. Melief, C. J. "Cancer Immunotherapy by Dendritic Cells." *Immunity* 29, no. 3 (2008): 372-83. <https://doi.org/10.1016/j.immuni.2008.08.004>.
75. Chen, J., A. Ganguly, A. D. Mucsi, J. Meng, J. Yan, P. Detampel, F. Munro, Z. Zhang, M. Wu, A. Hari, M. D. Stenner, W. Zheng, P. Kubes, T. Xia, M. W. Amrein, H. Qi, and Y. Shi. "Strong Adhesion by Regulatory T Cells Induces Dendritic Cell Cytoskeletal Polarization and Contact-Dependent Lethargy." *J Exp Med* 214, no. 2 (2017): 327-38. <https://doi.org/10.1084/jem.20160620>.
76. Wing, K., Y. Onishi, P. Prieto-Martin, T. Yamaguchi, M. Miyara, Z. Fehervari, T. Nomura, and S. Sakaguchi. "Ctla-4 Control over Foxp3+ Regulatory T Cell Function." *Science* 322, no. 5899 (2008): 271-5. <https://doi.org/10.1084/jem.20160620>.

77. Fife, B. T., and J. A. Bluestone. "Control of Peripheral T-Cell Tolerance and Autoimmunity Via the Ctlα-4 and Pd-1 Pathways." *Immunol Rev* 224 (2008): 166-82. <https://doi.org/10.1111/j.1600-065X.2008.00662.x>
78. Peng, L. S., J. Y. Zhang, Y. S. Teng, Y. L. Zhao, T. T. Wang, F. Y. Mao, Y. P. Lv, P. Cheng, W. H. Li, N. Chen, M. Duan, W. Chen, G. Guo, Q. M. Zou, and Y. Zhuang. "Tumor-Associated Monocytes/Macrophages Impair Nk-Cell Function Via Tgfbeta1 in Human Gastric Cancer." *Cancer Immunol Res* 5, no. 3 (2017): 248-56. <https://doi.org/10.1158/2326-6066.CIR-16-0152>.
79. Gordon, S., and P. R. Taylor. "Monocyte and Macrophage Heterogeneity." *Nat Rev Immunol* 5, no. 12 (2005): 953-64. <https://doi.org/10.1038/nri1733>.
80. Ruffell, B., N. I. Affara, and L. M. Coussens. "Differential Macrophage Programming in the Tumor Microenvironment." *Trends Immunol* 33, no. 3 (2012): 119-26. <https://doi.org/10.1016/j.it.2011.12.001>.
81. Chen, J. J., Y. C. Lin, P. L. Yao, A. Yuan, H. Y. Chen, C. T. Shun, M. F. Tsai, C. H. Chen, and P. C. Yang. "Tumor-Associated Macrophages: The Double-Edged Sword in Cancer Progression." *J Clin Oncol* 23, no. 5 (2005): 953-64. <https://doi.org/10.1200/JCO.2005.12.172>.
82. Jiang, M., J. Chen, W. Zhang, R. Zhang, Y. Ye, P. Liu, W. Yu, F. Wei, X. Ren, and J. Yu. "Interleukin-6 Trans-Signaling Pathway Promotes Immunosuppressive Myeloid-Derived Suppressor Cells Via Suppression of Suppressor of Cytokine Signaling 3 in Breast Cancer." *Front Immunol* 8 (2017): 1840. <https://doi.org/10.3389/fimmu.2017.01840>.
83. Shvedova, A. A., E. R. Kisin, N. Yanamala, A. V. Tkach, D. W. Gutkin, A. Star, G. V. Shurin, V. E. Kagan, and M. R. Shurin. "Mdsc and Tgfbeta Are Required for Facilitation of Tumor Growth in the Lungs of Mice Exposed to Carbon Nanotubes." *Cancer Res* 75, no. 8 (2015): 1615-23. <https://doi.org/10.1158/0008-5472.CAN-14-2376>.
84. Huang, B., P. Y. Pan, Q. Li, A. I. Sato, D. E. Levy, J. Bromberg, C. M. Divino, and S. H. Chen. "Gr-1+ Cd115+ Immature Myeloid Suppressor Cells Mediate the Development of Tumor-Induced T Regulatory Cells and T-Cell Anergy in Tumor-Bearing Host." *Cancer Res* 66, no. 2 (2006): 1123-31. <https://doi.org/10.1158/0008-5472.CAN-05-1299>.
85. Armstrong, D., C. Y. Chang, D. R. Lazarus, D. Corry, and F. Kheradmand. "Lung Cancer Heterogeneity in Modulation of Th17/IL17a Responses." *Front Oncol* 9 (2019): 1384. <https://doi.org/10.3389/fonc.2019.01384>.
86. Mucida, D., Y. Park, G. Kim, O. Turovskaya, I. Scott, M. Kronenberg, and H. Cheroutre. "Reciprocal Th17 and Regulatory T Cell Differentiation Mediated by Retinoic Acid." *Science* 317, no. 5835 (2007): 256-60. <https://doi.org/10.1126/science.1145697>.
87. Garrido-Mesa, N., F. Algieri, A. Rodriguez Nogales, and J. Galvez. "Functional Plasticity of Th17 Cells: Implications in Gastrointestinal Tract Function." *Int Rev Immunol* 32, no. 5-6 (2013): 493-510. <https://doi.org/10.3109/08830185.2013.834899>.
88. Vecchi, Lara, Thaise Gonçalves Araújo, Fernanda Van Petten de Vasconcelos Azevedo, Sara Teixeira Soares Mota, Veridiana de Melo Rodrigues Ávila, Matheus Alves Ribeiro, and Luiz Ricardo Goulart. "Phospholipase A2 Drives Tumorigenesis and Cancer Aggressiveness through Its Interaction with Annexin A1." 10, no. 6 (2021): 1472. <https://doi.org/10.3390/cells10061472>
89. Parente, L., and E. Solito. "Annexin 1: More Than an Anti-Phospholipase Protein." *Inflamm Res* 53, no. 4 (2004): 125-32. <https://doi.org/10.1007/s00011-003-1235-z>.
90. Gerke, Volker, Carl E. Creutz, and Stephen E. Moss. "Annexins: Linking Ca²⁺ Signalling to Membrane Dynamics." *Nature Reviews Molecular Cell Biology* 6, no. 6 (2005): 449-61. <https://doi.org/10.1038/nrm1661>.

91. Sheikh, M. H., and E. Solito. "Annexin A1: Uncovering the Many Talents of an Old Protein." *Int J Mol Sci* 19, no. 4 (2018). <https://doi.org/10.3390/ijms19041045>. 251 252
92. Pepinsky, R. B., L. K. Sinclair, J. L. Browning, R. J. Mattaliano, J. E. Smart, E. P. Chow, T. Falbel, A. Ribolini, J. L. Garwin, and B. P. Wallner. "Purification and Partial Sequence Analysis of a 37-Kda Protein That Inhibits Phospholipase A2 Activity from Rat Peritoneal Exudates." *J Biol Chem* 261, no. 9 (1986): 4239-46. [https://doi.org/10.1016/S0021-9258\(17\)35653-3](https://doi.org/10.1016/S0021-9258(17)35653-3). 253 254 255 256
93. Perretti, M., and F. D'Acquisto. "Annexin A1 and Glucocorticoids as Effectors of the Resolution of Inflammation." *Nat Rev Immunol* 9, no. 1 (2009): 62-70. <https://doi.org/10.1038/nri2470>. 257 258
94. Arcone, R., G. Arpaia, M. Ruoppolo, A. Malorni, P. Pucci, G. Marino, A. Ialenti, M. Di Rosa, and G. Ciliberto. "Structural Characterization of a Biologically Active Human Lipocortin 1 Expressed in Escherichia Coli." *Eur J Biochem* 211, no. 1-2 (1993): 347-55. <https://doi.org/10.1111/j.1432-1033.1993.tb19904.x>. 259 260 261
95. Flower, R. J. "Eleventh Gaddum Memorial Lecture. Lipocortin and the Mechanism of Action of the Glucocorticoids." *Br J Pharmacol* 94, no. 4 (1988): 987-1015. <https://doi.org/10.1111/j.1476-5381.1988.tb11614.x>. 262 263
96. Sugimoto, M. A., J. P. Vago, M. M. Teixeira, and L. P. Sousa. "Annexin A1 and the Resolution of Inflammation: Modulation of Neutrophil Recruitment, Apoptosis, and Clearance." *J Immunol Res* 2016 (2016): 8239258. <https://doi.org/10.1155/2016/8239258>. 264 265 266
97. Purvis, G. S. D., E. Solito, and C. Thiemermann. "Annexin-A1: Therapeutic Potential in Microvascular Disease." *Front Immunol* 10 (2019): 938. <https://doi.org/10.3389/fimmu.2019.00938>. 267 268
98. Shen, X., S. Zhang, Z. Guo, D. Xing, and W. Chen. "The Crosstalk of Abca1 and Anxa1: A Potential Mechanism for Protection against Atherosclerosis." *Mol Med* 26, no. 1 (2020): 84. <https://doi.org/10.1186/s10020-020-00213-y>. 269 270 271
99. Hall, S. C., D. M. Smith, F. R. Masiarz, V. W. Soo, H. M. Tran, L. B. Epstein, and A. L. Burlingame. "Mass Spectrometric and Edman Sequencing of Lipocortin I Isolated by Two-Dimensional Sds/Page of Human Melanoma Lysates." *Proc Natl Acad Sci U S A* 90, no. 5 (1993): 1927-31. <https://doi.org/10.1073/pnas.90.5.1927>. 272 273 274
100. Bai, F., P. Zhang, Y. Fu, H. Chen, M. Zhang, Q. Huang, D. Li, B. Li, and K. Wu. "Targeting Anxa1 Abrogates Treg-Mediated Immune Suppression in Triple-Negative Breast Cancer." *J Immunother Cancer* 8, no. 1 (2020). <https://doi.org/10.1136/jitc-2019-000169>. 275 276 277
101. Rosengarth, A., and H. Luecke. "A Calcium-Driven Conformational Switch of the N-Terminal and Core Domains of Annexin A1." *J Mol Biol* 326, no. 5 (2003): 1317-25. [https://doi.org/10.1016/S0022-2836\(03\)00027-5](https://doi.org/10.1016/S0022-2836(03)00027-5). 278 279
102. Rosengarth, A., V. Gerke, and H. Luecke. "X-Ray Structure of Full-Length Annexin 1 and Implications for Membrane Aggregation." *J Mol Biol* 306, no. 3 (2001): 489-98. <https://doi.org/10.1006/jmbi.2000.4423>. 280 281
103. Glenney, J. R., Jr., and B. F. Tack. "Amino-Terminal Sequence of P36 and Associated P10: Identification of the Site of Tyrosine Phosphorylation and Homology with S-100." *Proc Natl Acad Sci U S A* 82, no. 23 (1985): 7884-8. <https://doi.org/10.1073/pnas.82.23.7884>; 282 283 284
104. Donnelly, S. R., and S. E. Moss. "Functional Analysis of the Human Annexin I and Vi Gene Promoters." *Biochem J* 332 (Pt 3), no. Pt 3 (1998): 681-7. <https://doi.org/10.1042/bj3320681>. 285 286
105. Boudhraa, Z., B. Bouchon, C. Viallard, M. D'Incan, and F. Degoul. "Annexin A1 Localization and Its Relevance to Cancer." *Clin Sci (Lond)* 130, no. 4 (2016): 205-20. <https://doi.org/10.1042/CS20150415>. 287 288
106. Gerke, V., and S. E. Moss. "Annexins: From Structure to Function." *Physiol Rev* 82, no. 2 (2002): 331-71. <https://doi.org/10.1152/physrev.00030.2001>. 289 290
107. Moss, S. E., and R. O. Morgan. "The Annexins." *Genome Biol* 5, no. 4 (2004): 219. <https://doi.org/10.1186/gb-2004-5-4-219> 291 292

108. Weyd, H. "More Than Just Innate Affairs - on the Role of Annexins in Adaptive Immunity." *Biol Chem* 397, no. 10 (2016): 1017-29. <https://doi.org/10.1515/hsz-2016-0191>.
109. Grewal, T., M. Hoque, J. R. W. Conway, M. Reverter, M. Wahba, S. S. Beevi, P. Timpson, C. Enrich, and C. Rentero. "Annexin A6-a Multifunctional Scaffold in Cell Motility." *Cell Adh Migr* 11, no. 3 (2017): 288-304. <https://doi.org/10.1080/19336918.2016.1268318>.
110. Boudhraa, Z., C. Merle, D. Mazzocut, J. M. Chezal, C. Chambon, E. Miot-Noirault, M. Theisen, B. Bouchon, and F. Degoul. "Characterization of Pro-Invasive Mechanisms and N-Terminal Cleavage of Anxa1 in Melanoma." *Arch Dermatol Res* 306, no. 10 (2014): 903-14. <https://doi.org/10.1007/s00403-014-1517-z>.
111. Gerdes, H. H. "Membrane Traffic in the Secretory Pathway." *Cell Mol Life Sci* 65, no. 18 (2008): 2779-80. <https://doi.org/10.1007/s00018-008-8348-z>.
112. Han, G., Y. Tian, B. Duan, H. Sheng, H. Gao, and J. Huang. "Association of Nuclear Annexin A1 with Prognosis of Patients with Esophageal Squamous Cell Carcinoma." *Int J Clin Exp Pathol* 7, no. 2 (2014): 751-9.
113. Futter, C. E., and I. J. White. "Annexins and Endocytosis." *Traffic* 8, no. 8 (2007): 951-8. <https://doi.org/10.1111/j.1600-0854.2007.00590.x>.
114. Lim, L. H., and S. Pervaiz. "Annexin 1: The New Face of an Old Molecule." *Faseb j* 21, no. 4 (2007): 968-75. <https://doi.org/10.1096/fj.06-7464rev>.
115. Hirata, A., and F. Hirata. "DNA Chain Unwinding and Annealing Reactions of Lipocortin (Annexin) I Heterotetramer: Regulation by Ca(2+) and Mg(2+)." *Biochem Biophys Res Commun* 291, no. 2 (2002): 205-9. <https://doi.org/10.1006/bbrc.2002.6422>.
116. Lin, S., R. Hickey, and L. Malkas. "The Biochemical Status of the DNA Synthesosome Can Distinguish between Permanent and Temporary Cell Growth Arrest." *Cell Growth Differ* 8, no. 12 (1997): 1359-69.
117. Rescher, U., N. Zobiack, and V. Gerke. "Intact Ca(2+)-Binding Sites Are Required for Targeting of Annexin 1 to Endosomal Membranes in Living Hela Cells." *J Cell Sci* 113 (Pt 22) (2000): 3931-8. <https://doi.org/10.1242/jcs.113.22.3931>.
118. Sudlow, A. W., F. Carey, R. Forder, and N. J. Rothwell. "The Role of Lipocortin-1 in Dexamethasone-Induced Suppression of Pge2 and Tnf Alpha Release from Human Peripheral Blood Mononuclear Cells." *Br J Pharmacol* 117, no. 7 (1996): 1449-56. <https://doi.org/10.1111/j.1476-5381.1996.tb15305.x>.
119. Sanches, J. M., L. M. Branco, G. H. B. Duarte, S. M. Oliani, K. R. Bortoluci, V. Moreira, and C. D. Gil. "Annexin A1 Regulates Nlrp3 Inflammasome Activation and Modifies Lipid Release Profile in Isolated Peritoneal Macrophages." *Cells* 9, no. 4 (2020). <https://doi.org/10.3390/cells9040926>.
120. Seidel, S., H. Neymeyer, T. Kahl, T. Röschel, K. Mutig, R. Flower, J. Schnermann, S. Bachmann, and A. Paliege. "Annexin A1 Modulates Macula Densa Function by Inhibiting Cyclooxygenase 2." *Am J Physiol Renal Physiol* 303, no. 6 (2012): F845-54. <https://doi.org/10.1152/ajprenal.00704.2011>.
121. Flower, R. J., and N. J. Rothwell. "Lipocortin-1: Cellular Mechanisms and Clinical Relevance." *Trends Pharmacol Sci* 15, no. 3 (1994): 71-6. [https://doi.org/10.1016/0165-6147\(94\)90281-X](https://doi.org/10.1016/0165-6147(94)90281-X).
122. Flower, R. J., and G. J. Blackwell. "Anti-Inflammatory Steroids Induce Biosynthesis of a Phospholipase A2 Inhibitor Which Prevents Prostaglandin Generation." *Nature* 278, no. 5703 (1979): 456-9. <https://doi.org/10.1038/278456a0>.
123. Leoni, G., and A. Nusrat. "Annexin A1: Shifting the Balance Towards Resolution and Repair." *Biol Chem* 397, no. 10 (2016): 971-9. <https://doi.org/10.1515/hsz-2016-0180>.

124. Chatterjee, B. E., S. Yona, G. Rosignoli, R. E. Young, S. Nourshargh, R. J. Flower, and M. Perretti. "Annexin 1-Deficient Neutrophils Exhibit Enhanced Transmigration in Vivo and Increased Responsiveness in Vitro." *J Leukoc Biol* 78, no. 3 (2005): 639-46. <https://doi.org/10.1189/jlb.0405206>.
125. Kiani-Esfahani, A., S. Kazemi Sheykhshabani, M. Peymani, M. S. Hashemi, K. Ghaedi, and M. H. Nasr-Esfahani. "Overexpression of Annexin A1 Suppresses Pro-Inflammatory Factors in Pc12 Cells Induced by 1-Methyl-4-Phenylpyridinium." *Cell J* 18, no. 2 (2016): 197-204.
126. Sakaguchi, M., H. Murata, H. Sonogawa, Y. Sakaguchi, J. Futami, M. Kitazoe, H. Yamada, and N. H. Huh. "Truncation of Annexin A1 Is a Regulatory Lever for Linking Epidermal Growth Factor Signaling with Cytosolic Phospholipase A2 in Normal and Malignant Squamous Epithelial Cells." *J Biol Chem* 282, no. 49 (2007): 35679-86. <https://doi.org/10.1074/jbc.M707538200>.
127. Sakaguchi, M., and N. H. Huh. "S100a11, a Dual Growth Regulator of Epidermal Keratinocytes." *Amino Acids* 41, no. 4 (2011): 797-807. <https://doi.org/10.1007/s00726-010-0747-4>.
128. Vander Heiden, M. G., L. C. Cantley, and C. B. Thompson. "Understanding the Warburg Effect: The Metabolic Requirements of Cell Proliferation." *Science* 324, no. 5930 (2009): 1029-33. <https://doi.org/10.1126/science.1160809>.
129. Santamaria-Kisiel, L., A. C. Rintala-Dempsey, and G. S. Shaw. "Calcium-Dependent and -Independent Interactions of the S100 Protein Family." *Biochem J* 396, no. 2 (2006): 201-14. <https://doi.org/10.1042/BJ20060195>.
130. Blume, K. E., S. Soeroes, H. Keppeler, S. Stevanovic, D. Kretschmer, M. Rautenberg, S. Wesselborg, and K. Lauber. "Cleavage of Annexin A1 by Adam10 During Secondary Necrosis Generates a Monocytic "Find-Me" Signal." *J Immunol* 188, no. 1 (2012): 135-45. <https://doi.org/10.4049/jimmunol.1004073>
131. Movitz, C., C. Sjolin, and C. Dahlgren. "Cleavage of Annexin I in Human Neutrophils Is Mediated by a Membrane-Localized Metalloprotease." *Biochim Biophys Acta* 1416, no. 1-2 (1999): 101-8. [https://doi.org/10.1016/S0005-2736\(98\)00212-0](https://doi.org/10.1016/S0005-2736(98)00212-0).
132. Vong, L., F. D'Acquisto, M. Pederzoli-Ribeil, L. Lavagno, R. J. Flower, V. Witko-Sarsat, and M. Perretti. "Annexin 1 Cleavage in Activated Neutrophils: A Pivotal Role for Proteinase 3." *J Biol Chem* 282, no. 41 (2007): 29998-30004. <https://doi.org/10.1074/jbc.M702876200>.
133. Wang, W., and C. E. Creutz. "Role of the Amino-Terminal Domain in Regulating Interactions of Annexin I with Membranes: Effects of Amino-Terminal Truncation and Mutagenesis of the Phosphorylation Sites." *Biochemistry* 33, no. 1 (1994): 275-82. <https://doi.org/10.1021/bi00167a036>
134. Rescher, U., V. Goebeler, A. Wilbers, and V. Gerke. "Proteolytic Cleavage of Annexin 1 by Human Leukocyte Elastase." *Biochim Biophys Acta* 1763, no. 11 (2006): 1320-4. <https://doi.org/10.1016/j.bbamcr.2006.08.041>.
135. Shao, G., H. Zhou, Q. Zhang, Y. Jin, and C. Fu. "Advancements of Annexin A1 in Inflammation and Tumorigenesis." *Onco Targets Ther* 12 (2019): 3245-54. <https://doi.org/10.2147/OTT.S202271>.
136. Williams, S. L., I. R. Milne, C. J. Bagley, J. R. Gamble, M. A. Vadas, S. M. Pitson, and Y. Khew-Goodall. "A Proinflammatory Role for Proteolytically Cleaved Annexin A1 in Neutrophil Transendothelial Migration." *J Immunol* 185, no. 5 (2010): 3057-63. <https://doi.org/10.4049/jimmunol.1000119>.
137. Solito, E., H. C. Christian, M. Festa, A. Mulla, T. Tierney, R. J. Flower, and J. C. Buckingham. "Post-Translational Modification Plays an Essential Role in the Translocation of Annexin A1 from the Cytoplasm to the Cell Surface." *Faseb j* 20, no. 9 (2006): 1498-500. <https://doi.org/10.1096/fj.05-5319fje>.
138. Solito, E., A. Kamal, F. Russo-Marie, J. C. Buckingham, S. Marullo, and M. Perretti. "A Novel Calcium-Dependent Proapoptotic Effect of Annexin 1 on Human Neutrophils." *Faseb j* 17, no. 11 (2003): 1544-6. <https://doi.org/10.1096/fj.02-0941fje>.

139. Frambach, Sjc, R. de Haas, J. A. M. Smeitink, G. A. Rongen, F. G. M. Russel, and T. J. J. Schirris. "Brothers in Arms: Abca1- and Abcg1-Mediated Cholesterol Efflux as Promising Targets in Cardiovascular Disease Treatment." *Pharmacol Rev* 72, no. 1 (2020): 152-90. <https://doi.org/10.1124/pr.119.017897>.
140. Hafiane, A., and J. Genest. "Atp Binding Cassette A1 (Abca1) Mediates Microparticle Formation During High-Density Lipoprotein (Hdl) Biogenesis." *Atherosclerosis* 257 (2017): 90-99. <https://doi.org/10.1016/j.atherosclerosis.2017.01.013>.
141. Nandi, S., L. Ma, M. Denis, J. Karwatsky, Z. Li, X. C. Jiang, and X. Zha. "Abca1-Mediated Cholesterol Efflux Generates Microparticles in Addition to Hdl through Processes Governed by Membrane Rigidity." *J Lipid Res* 50, no. 3 (2009): 456-66. <https://doi.org/10.1194/jlr.M800345-JLR200>.
142. Brancalone, V., E. Mitidieri, R. J. Flower, G. Cirino, and M. Perretti. "Annexin A1 Mediates Hydrogen Sulfide Properties in the Control of Inflammation." *J Pharmacol Exp Ther* 351, no. 1 (2014): 96-104. <https://doi.org/10.1124/jpet.114.217034>.
143. Perretti, M., H. Christian, S. K. Wheller, I. Aiello, K. G. Mugridge, J. F. Morris, R. J. Flower, and N. J. Goulding. "Annexin I Is Stored within Gelatinase Granules of Human Neutrophil and Mobilized on the Cell Surface Upon Adhesion but Not Phagocytosis." *Cell Biol Int* 24, no. 3 (2000): 163-74. <https://doi.org/10.1006/cbir.1999.0468>.
144. Li, Y., and D. Ye. "Molecular Biology for Formyl Peptide Receptors in Human Diseases." *J Mol Med (Berl)* 91, no. 7 (2013): 781-9. <https://doi.org/10.1007/s00109-013-1005-5>.
145. Ye, R. D., F. Boulay, J. M. Wang, C. Dahlgren, C. Gerard, M. Parmentier, C. N. Serhan, and P. M. Murphy. "International Union of Basic and Clinical Pharmacology. Lxxiii. Nomenclature for the Formyl Peptide Receptor (Fpr) Family." *Pharmacol Rev* 61, no. 2 (2009): 119-61. <https://doi.org/10.1124/pr.109.001578>.
146. Migeotte, I., D. Communi, and M. Parmentier. "Formyl Peptide Receptors: A Promiscuous Subfamily of G Protein-Coupled Receptors Controlling Immune Responses." *Cytokine Growth Factor Rev* 17, no. 6 (2006): 501-19. <https://doi.org/10.1016/j.cytogfr.2006.09.009>.
147. Perretti, M., N. Chiang, M. La, I. M. Fierro, S. Marullo, S. J. Getting, E. Solito, and C. N. Serhan. "Endogenous Lipid- and Peptide-Derived Anti-Inflammatory Pathways Generated with Glucocorticoid and Aspirin Treatment Activate the Lipoxin A4 Receptor." *Nat Med* 8, no. 11 (2002): 1296-302. <https://doi.org/10.1038/nm786>.
148. Dalpiaz, A., S. Spisani, C. Biondi, E. Fabbri, M. Nalli, and M. E. Ferretti. "Studies on Human Neutrophil Biological Functions by Means of Formyl-Peptide Receptor Agonists and Antagonists." *Curr Drug Targets Immune Endocr Metabol Disord* 3, no. 1 (2003): 33-42. <https://doi.org/10.2174/1568008033340333>.
149. Gavins, F. N., and M. J. Hickey. "Annexin A1 and the Regulation of Innate and Adaptive Immunity." *Front Immunol* 3 (2012): 354. <https://doi.org/10.3389/fimmu.2012.00354>.
150. Cooray, S. N., T. Gobbetti, T. Montero-Melendez, S. McArthur, D. Thompson, A. J. Clark, R. J. Flower, and M. Perretti. "Ligand-Specific Conformational Change of the G-Protein-Coupled Receptor Alx/Fpr2 Determines Proresolving Functional Responses." *Proc Natl Acad Sci U S A* 110, no. 45 (2013): 18232-7. <https://doi.org/10.1073/pnas.1308253110>.
151. Li, L., K. Chen, Y. Xiang, T. Yoshimura, S. Su, J. Zhu, X. W. Bian, and J. M. Wang. "New Development in Studies of Formyl-Peptide Receptors: Critical Roles in Host Defense." *J Leukoc Biol* 99, no. 3 (2016): 425-35. <https://doi.org/10.1189/jlb.2RI0815-354RR>.
152. Vacchelli, E., Y. Ma, E. E. Baracco, A. Sistigu, D. P. Enot, F. Pietrocola, H. Yang, S. Adjemian, K. Chaba, M. Semeraro, M. Signore, A. De Ninno, V. Lucarini, F. Peschiaroli, L. Businaro, A. Gerardino, G. Manic, T. Ulas,

- P. Günther, J. L. Schultze, O. Kepp, G. Stoll, C. Lefebvre, C. Mulot, F. Castoldi, S. Rusakiewicz, S. Ladoire, L. Apetoh, J. M. Bravo-San Pedro, M. Lucattelli, C. Delarasse, V. Boige, M. Ducreux, S. Delaloge, C. Borg, F. André, G. Schiavoni, I. Vitale, P. Laurent-Puig, F. Mattei, L. Zitvogel, and G. Kroemer. "Chemotherapy-Induced Antitumor Immunity Requires Formyl Peptide Receptor 1." *Science* 350, no. 6263 (2015): 972-8. <https://doi.org/10.1126/science.aad0779>.
153. Nagaya, T., K. Kawata, R. Kamekura, S. Jitsukawa, T. Kubo, M. Kamei, N. Ogasawara, K. I. Takano, T. Himi, and S. Ichimiya. "Lipid Mediators Foster the Differentiation of T Follicular Helper Cells." *Immunol Lett* 181 (2017): 51-57. <https://doi.org/10.1016/j.imlet.2016.11.006>.
154. Chen, K., Y. Xiang, J. Huang, W. Gong, T. Yoshimura, Q. Jiang, L. Tessarollo, Y. Le, and J. M. Wang. "The Formylpeptide Receptor 2 (Fpr2) and Its Endogenous Ligand Cathelin-Related Antimicrobial Peptide (Cramp) Promote Dendritic Cell Maturation." *J Biol Chem* 289, no. 25 (2014): 17553-63. <https://doi.org/10.1074/jbc.M113.535674>.
155. Kim, S. D., J. M. Kim, S. H. Jo, H. Y. Lee, S. Y. Lee, J. W. Shim, S. K. Seo, J. Yun, and Y. S. Bae. "Functional Expression of Formyl Peptide Receptor Family in Human Nk Cells." *J Immunol* 183, no. 9 (2009): 5511-7. <https://doi.org/10.4049/jimmunol.0802986>.
156. McArthur, S., T. Gobbetti, D. H. Kusters, C. P. Reutelingsperger, R. J. Flower, and M. Perretti. "Definition of a Novel Pathway Centered on Lysophosphatidic Acid to Recruit Monocytes During the Resolution Phase of Tissue Inflammation." *J Immunol* 195, no. 3 (2015): 1139-51. <https://doi.org/10.4049/jimmunol.0802986>.
157. Peshavariya, H. M., C. J. Taylor, C. Goh, G. S. Liu, F. Jiang, E. C. Chan, and G. J. Dusting. "Annexin Peptide Ac2-26 Suppresses Tnfalpha-Induced Inflammatory Responses Via Inhibition of Rac1-Dependent NADPH Oxidase in Human Endothelial Cells." *PLoS One* 8, no. 4 (2013): e60790. <https://doi.org/10.1371/journal.pone.0060790>.
158. Miki, T., S. Yano, M. Hanibuchi, and S. Sone. "Bone Metastasis Model with Multiorgan Dissemination of Human Small-Cell Lung Cancer (Sbc-5) Cells in Natural Killer Cell-Depleted Scid Mice." *Oncol Res* 12, no. 5 (2000): 209-17. <https://doi.org/10.3727/096504001108747701>.
159. Huang, S., L. Y. Chen, B. L. Zuraw, R. D. Ye, and Z. K. Pan. "Chemoattractant-Stimulated NF-κB Activation Is Dependent on the Low Molecular Weight GTPase RhoA." *J Biol Chem* 276, no. 44 (2001): 40977-81. <https://doi.org/10.1074/jbc.M105242200>.
160. Han, P. F., X. D. Che, H. Z. Li, Y. Y. Gao, X. C. Wei, and P. C. Li. "Annexin A1 Involved in the Regulation of Inflammation and Cell Signaling Pathways." *Chin J Traumatol* 23, no. 2 (2020): 96-101. <https://doi.org/10.1016/j.cjtee.2020.02.002>.
161. Biaoxue, R., C. Xiguang, and Y. Shuanying. "Annexin A1 in Malignant Tumors: Current Opinions and Controversies." *Int J Biol Markers* 29, no. 1 (2014): e8-20. <https://doi.org/10.5301/jbm.5000046>.
162. Wang, Y., L. Serfass, M. O. Roy, J. Wong, A. M. Bonneau, and E. Georges. "Annexin-I Expression Modulates Drug Resistance in Tumor Cells." *Biochem Biophys Res Commun* 314, no. 2 (2004): 565-70. <https://doi.org/10.1016/j.bbrc.2003.12.117>.
163. Liu, Y., X. Hu, C. Han, L. Wang, X. Zhang, X. He, and X. Lu. "Targeting Tumor Suppressor Genes for Cancer Therapy." *Bioessays* 37, no. 12 (2015): 1277-86. <https://doi.org/10.1002/bies.201500093>.
164. Cao, Y., Y. Li, M. Edelweiss, B. Arun, D. Rosen, E. Resetskova, Y. Wu, J. Liu, A. Sahin, and C. T. Albarracin. "Loss of Annexin A1 Expression in Breast Cancer Progression." *Appl Immunohistochem Mol Morphol* 16, no. 6 (2008): 530-4. <https://doi.org/10.1097/PAI.0b013e31817432c3>.

165. Hnisz, D., A. S. Weintraub, D. S. Day, A. L. Valton, R. O. Bak, C. H. Li, J. Goldmann, B. R. Lajoie, Z. P. Fan, A. A. Sigova, J. Reddy, D. Borges-Rivera, T. I. Lee, R. Jaenisch, M. H. Porteus, J. Dekker, and R. A. Young. "Activation of Proto-Oncogenes by Disruption of Chromosome Neighborhoods." *Science* 351, no. 6280 (2016): 1454-58. <https://doi.org/10.1126/science.aad9024>.
166. Anderson, M. W., S. H. Reynolds, M. You, and R. M. Maronpot. "Role of Proto-Oncogene Activation in Carcinogenesis." *Environ Health Perspect* 98 (1992): 13-24. <https://doi.org/10.1289/ehp.929813>.
167. Foo, S. L., G. Yap, J. Cui, and L. H. K. Lim. "Annexin-A1 - a Blessing or a Curse in Cancer?" *Trends Mol Med* 25, no. 4 (2019): 315-27. <https://doi.org/10.1016/j.molmed.2019.02.004>.
168. Sobral-Leite, M., J. Wesseling, V. T. Smit, H. Nevanlinna, M. H. van Miltenburg, J. Sanders, I. Hofland, F. M. Blows, P. Coulson, G. Patrycja, J. H. Schellens, R. Fagerholm, P. Heikkila, K. Aittomaki, C. Blomqvist, E. Provenzano, H. R. Ali, J. Figueroa, M. Sherman, J. Lissowska, A. Mannermaa, V. Kataja, V. M. Kosma, J. M. Hartikainen, K. A. Phillips, Aocs Investigators kConFab, F. J. Couch, J. E. Olson, C. Vachon, D. Visscher, H. Brenner, K. Butterbach, V. Arndt, B. Holleczeck, M. J. Hoening, A. Hollestelle, J. W. Martens, C. H. van Deurzen, B. van de Water, A. Broeks, J. Chang-Claude, G. Chenevix-Trench, D. F. Easton, P. D. Pharoah, M. Garcia-Closas, M. de Graauw, and M. K. Schmidt. "Annexin A1 Expression in a Pooled Breast Cancer Series: Association with Tumor Subtypes and Prognosis." *BMC Med* 13 (2015): 156. <https://doi.org/10.1186/s12916-015-0392-6>.
169. Vecchi, L., M. Alves Pereira Zoia, T. Goss Santos, A. de Oliveira Beserra, C. M. Colaco Ramos, B. Franca Matias Colombo, Y. C. Paiva Maia, V. Piana de Andrade, S. Teixeira Soares Mota, T. Goncalves de Araujo, F. Van Petten de Vasconcelos Azevedo, F. A. Soares, S. M. Oliani, and L. R. Goulart. "Inhibition of the Anxa1/Fpr1 Autocrine Axis Reduces Mda-Mb-231 Breast Cancer Cell Growth and Aggressiveness in Vitro and in Vivo." *Biochim Biophys Acta Mol Cell Res* 1865, no. 9 (2018): 1368-82. <https://doi.org/10.1016/j.bbamcr.2018.06.010>.
170. Boudhraa, Z., F. Rondepierre, L. Ouchchane, R. Kintossou, A. Trzeciakiewicz, F. Franck, J. Kanitakis, B. Labeille, J. Joubert-Zakeyh, B. Bouchon, J. L. Perrot, S. Mansard, J. Papon, P. Dechelotte, J. M. Chezal, E. Miot-Noirault, M. Bonnet, M. D'Incan, and F. Degoul. "Annexin A1 in Primary Tumors Promotes Melanoma Dissemination." *Clin Exp Metastasis* 31, no. 7 (2014): 749-60. <https://doi.org/10.1007/s10585-014-9665-2>.
171. Lin, Y., G. Lin, W. Fang, H. Zhu, and K. Chu. "Increased Expression of Annexin A1 Predicts Poor Prognosis in Human Hepatocellular Carcinoma and Enhances Cell Malignant Phenotype." *Med Oncol* 31, no. 12 (2014): 327. <https://doi.org/10.1007/s12032-014-0327-7>.
172. Su, N., X. Y. Xu, H. Chen, W. C. Gao, C. P. Ruan, Q. Wang, and Y. P. Sun. "Increased Expression of Annexin A1 Is Correlated with K-Ras Mutation in Colorectal Cancer." *Tohoku J Exp Med* 222, no. 4 (2010): 243-50. <https://doi.org/10.1620/tjem.222.243>.
173. Cheng, S. X., Y. Tu, and S. Zhang. "Foxm1 Promotes Glioma Cells Progression by up-Regulating Anxa1 Expression." *PLoS One* 8, no. 8 (2013): e72376. <https://doi.org/10.1371/journal.pone.0072376>.
174. Liu, Y. F., P. F. Zhang, M. Y. Li, Q. Q. Li, and Z. C. Chen. "Identification of Annexin A1 as a Proinvasive and Prognostic Factor for Lung Adenocarcinoma." *Clin Exp Metastasis* 28, no. 5 (2011): 413-25. <https://doi.org/10.1007/s10585-011-9380-1>.
175. Biaoxue, R., J. Xiling, Y. Shuanying, Z. Wei, C. Xiguang, W. Jinsui, and Z. Min. "Upregulation of Hsp90-Beta and Annexin A1 Correlates with Poor Survival and Lymphatic Metastasis in Lung Cancer Patients." *J Exp Clin Cancer Res* 31, no. 1 (2012): 70. <https://doi.org/10.1007/s10585-011-9380-1>.

176. Mota, S. T. S., L. Vecchi, D. A. Alves, A. O. Cordeiro, G. S. Guimaraes, E. Campos-Fernandez, Y. C. P. Maia, B. C. Dornelas, S. M. Bezerra, V. P. de Andrade, L. R. Goulart, and T. G. Araujo. "Annexin A1 Promotes the Nuclear Localization of the Epidermal Growth Factor Receptor in Castration-Resistant Prostate Cancer." *Int J Biochem Cell Biol* 127 (2020): 105838. <https://doi.org/10.1016/j.biocel.2020.105838>.
177. Sato, Y., K. Kumamoto, K. Saito, H. Okayama, S. Hayase, Y. Kofunato, K. Miyamoto, I. Nakamura, S. Ohki, Y. Koyama, and S. Takenoshita. "Up-Regulated Annexin A1 Expression in Gastrointestinal Cancer Is Associated with Cancer Invasion and Lymph Node Metastasis." *Exp Ther Med* 2, no. 2 (2011): 239-43. <https://doi.org/10.3892/etm.2011.210>.
178. Rodrigo, J. P., J. M. Garcia-Pedrero, M. P. Fernandez, R. O. Morgan, C. Suárez, and A. Herrero. "Annexin A1 Expression in Nasopharyngeal Carcinoma Correlates with Squamous Differentiation." *Am J Rhinol* 19, no. 5 (2005): 483-7. <https://doi.org/10.1177/194589240501900511>.
179. Petrella, A., M. Festa, S. F. Ercolino, M. Zerilli, G. Stassi, E. Solito, and L. Parente. "Annexin-1 Downregulation in Thyroid Cancer Correlates to the Degree of Tumor Differentiation." *Cancer Biol Ther* 5, no. 6 (2006): 643-7. <https://doi.org/10.4161/cbt.5.6.2700>.
180. Garcia Pedrero, J. M., M. P. Fernandez, R. O. Morgan, A. Herrero Zapatero, M. V. Gonzalez, C. Suarez Nieto, and J. P. Rodrigo. "Annexin A1 Down-Regulation in Head and Neck Cancer Is Associated with Epithelial Differentiation Status." *Am J Pathol* 164, no. 1 (2004): 73-9. [https://doi.org/10.1016/S0002-9440\(10\)63098-2](https://doi.org/10.1016/S0002-9440(10)63098-2).
181. Shen, D., F. Nooraie, Y. Elshimali, V. Lonsberry, J. He, S. Bose, D. Chia, D. Seligson, H. R. Chang, and L. Goodglick. "Decreased Expression of Annexin A1 Is Correlated with Breast Cancer Development and Progression as Determined by a Tissue Microarray Analysis." *Hum Pathol* 37, no. 12 (2006): 1583-91. <https://doi.org/10.1016/j.humpath.2006.06.001>.
182. Yom, C. K., W. Han, S. W. Kim, H. S. Kim, H. C. Shin, J. N. Chang, M. Koo, D. Y. Noh, and B. I. Moon. "Clinical Significance of Annexin A1 Expression in Breast Cancer." *J Breast Cancer* 14, no. 4 (2011): 262-8. <https://doi.org/10.4048/jbc.2011.14.4.262>.
183. de Graauw, M., M. H. van Miltenburg, M. K. Schmidt, C. Pont, R. Lalai, J. Kartopawiro, E. Pardali, S. E. Le Devedec, V. T. Smit, A. van der Wal, L. J. Van't Veer, A. M. Cleton-Jansen, P. ten Dijke, and B. van de Water. "Annexin A1 Regulates Tgf-Beta Signaling and Promotes Metastasis Formation of Basal-Like Breast Cancer Cells." *Proc Natl Acad Sci U S A* 107, no. 14 (2010): 6340-5. <https://doi.org/10.1073/pnas.0913360107>.
184. Anbalagan, D., G. Yap, Y. Yuan, V. K. Pandey, W. H. Lau, S. Arora, P. Bist, J. S. Wong, G. Sethi, P. M. Nissom, P. E. Lobie, and L. H. Lim. "Annexin-A1 Regulates Microrna-26b* and Microrna-562 to Directly Target Nf-Kappab and Angiogenesis in Breast Cancer Cells." *PLoS One* 9, no. 12 (2014): e114507. <https://doi.org/10.1371/journal.pone.0114507>.
185. Cheng, T. Y., M. S. Wu, J. T. Lin, M. T. Lin, C. T. Shun, H. Y. Huang, K. T. Hua, and M. L. Kuo. "Annexin A1 Is Associated with Gastric Cancer Survival and Promotes Gastric Cancer Cell Invasiveness through the Formyl Peptide Receptor/Extracellular Signal-Regulated Kinase/Integrin Beta-1-Binding Protein 1 Pathway." *Cancer* 118, no. 23 (2012): 5757-67. <https://doi.org/10.1002/cncr.27565>.
186. Zhu, J. F., W. Huang, H. M. Yi, T. Xiao, J. Y. Li, J. Feng, H. Yi, S. S. Lu, X. H. Li, R. H. Lu, Q. Y. He, and Z. Q. Xiao. "Annexin A1-Suppressed Autophagy Promotes Nasopharyngeal Carcinoma Cell Invasion and Metastasis by Pi3k/Akt Signaling Activation." *Cell Death Dis* 9, no. 12 (2018): 1154. <https://doi.org/10.1038/s41419-018-1204-7>.

187. Chen, P., J. Min, H. Wu, H. Zhang, C. Wang, G. Tan, and F. Zhang. "Annexin A1 Is a Potential Biomarker of Bone Metastasis in Small Cell Lung Cancer." *Oncol Lett* 21, no. 2 (2021): 141. <https://doi.org/10.3892/ol.2020.12402>.
188. Yao, X. H., Y. F. Ping, J. H. Chen, D. L. Chen, C. P. Xu, J. Zheng, J. M. Wang, and X. W. Bian. "Production of Angiogenic Factors by Human Glioblastoma Cells Following Activation of the G-Protein Coupled Formylpeptide Receptor Fpr." *J Neurooncol* 86, no. 1 (2008): 47-53. <https://doi.org/10.1007/s11060-007-9443-y>.
189. Huang, J., K. Chen, J. Chen, W. Gong, N. M. Dunlop, O. M. Howard, Y. Gao, X. W. Bian, and J. M. Wang. "The G-Protein-Coupled Formylpeptide Receptor Fpr Confers a More Invasive Phenotype on Human Glioblastoma Cells." *Br J Cancer* 102, no. 6 (2010): 1052-60. <https://doi.org/10.1038/sj.bjc.6605591>.
190. Zhou, Y., X. Bian, Y. Le, W. Gong, J. Hu, X. Zhang, L. Wang, P. Iribarren, R. Salcedo, O. M. Howard, W. Farrar, and J. M. Wang. "Formylpeptide Receptor Fpr and the Rapid Growth of Malignant Human Gliomas." *J Natl Cancer Inst* 97, no. 11 (2005): 823-35. <https://doi.org/10.1093/jnci/dji142>.
191. Chen, D. L., Y. F. Ping, S. C. Yu, J. H. Chen, X. H. Yao, X. F. Jiang, H. R. Zhang, Q. L. Wang, and X. W. Bian. "Downregulating Fpr Restrains Xenograft Tumors by Impairing the Angiogenic Potential and Invasive Capability of Malignant Glioma Cells." *Biochem Biophys Res Commun* 381, no. 3 (2009): 448-52. <https://doi.org/10.1016/j.bbrc.2009.02.065>.
192. Snapkov, I., C. O. Oqvist, Y. Figenschau, P. Kogner, J. I. Johnsen, and B. Sveinbjornsson. "The Role of Formyl Peptide Receptor 1 (Fpr1) in Neuroblastoma Tumorigenesis." *BMC Cancer* 16 (2016): 490. <https://doi.org/10.1186/s12885-016-2545-1>.
193. Alldridge, L. C., H. J. Harris, R. Plevin, R. Hannon, and C. E. Bryant. "The Annexin Protein Lipocortin 1 Regulates the Mapk/Erk Pathway." *J Biol Chem* 274, no. 53 (1999): 37620-8. <https://doi.org/10.1074/jbc.274.53.37620>.
194. Jo, E. J., H. Y. Lee, J. I. Kim, H. K. Kang, Y. N. Lee, J. Y. Kwak, and Y. S. Bae. "Activation of Formyl Peptide Receptor-Like 1 by Wkymvm Induces Serine Phosphorylation of Stat3, Which Inhibits Its Tyrosine Phosphorylation and Nuclear Translocation Induced by Hydrogen Peroxide." *Life Sci* 75, no. 18 (2004): 2217-32. <https://doi.org/10.1016/j.lfs.2004.04.023>.
195. Shi, Y., X. Lai, L. Ye, K. Chen, Z. Cao, W. Gong, L. Jin, C. Wang, M. Liu, Y. Liao, J. M. Wang, and N. Zhou. "Activated Niacin Receptor Hca2 Inhibits Chemoattractant-Mediated Macrophage Migration Via Gβγ/Pkc/Erk1/2 Pathway and Heterologous Receptor Desensitization." *Sci Rep* 7 (2017): 42279. <https://doi.org/10.1038/srep42279>.
196. Wang, X., W. Qin, M. Song, Y. Zhang, and B. Sun. "Exogenous Carbon Monoxide Inhibits Neutrophil Infiltration in Lps-Induced Sepsis by Interfering with Fpr1 Via P38 Mapk but Not Grk2." *Oncotarget* 7, no. 23 (2016): 34250-65. <https://doi.org/10.18632/oncotarget.9084>.
197. Cao, G., and Z. Zhang. "Fpr1 Mediates the Tumorigenicity of Human Cervical Cancer Cells." *Cancer Manag Res* 10 (2018): 5855-65. <https://doi.org/10.2147/CMAR.S182795>.
198. Khau, T., S. Y. Langenbach, M. Schuliga, T. Harris, C. N. Johnstone, R. L. Anderson, and A. G. Stewart. "Annexin-1 Signals Mitogen-Stimulated Breast Tumor Cell Proliferation by Activation of the Formyl Peptide Receptors (Fprs) 1 and 2." *Faseb j* 25, no. 2 (2011): 483-96. <https://doi.org/10.1096/fj.09-154096>.
199. Chen, K., Z. Bao, W. Gong, P. Tang, T. Yoshimura, and J. M. Wang. "Regulation of Inflammation by Members of the Formyl-Peptide Receptor Family." *J Autoimmun* 85 (2017): 64-77. <https://doi.org/10.1016/j.jaut.2017.06.012>.

200. Cheng, T. Y., M. S. Wu, J. T. Lin, M. T. Lin, C. T. Shun, K. T. Hua, and M. L. Kuo. "Formyl Peptide Receptor 1 Expression Is Associated with Tumor Progression and Survival in Gastric Cancer." *Anticancer Res* 34, no. 5 (2014): 2223-9.
201. Prevete, N., F. Liotti, C. Visciano, G. Marone, R. M. Melillo, and A. de Paulis. "The Formyl Peptide Receptor 1 Exerts a Tumor Suppressor Function in Human Gastric Cancer by Inhibiting Angiogenesis." *Oncogene* 34, no. 29 (2015): 3826-38. <https://doi.org/10.1038/onc.2014.309>.
202. Lin, C. Y., Y. M. Jeng, H. Y. Chou, H. C. Hsu, R. H. Yuan, C. P. Chiang, and M. Y. Kuo. "Nuclear Localization of Annexin A1 Is a Prognostic Factor in Oral Squamous Cell Carcinoma." *J Surg Oncol* 97, no. 6 (2008): 544-50. <https://doi.org/10.1002/jso.20992>.
203. Zhu, F., C. Xu, Z. Jiang, M. Jin, L. Wang, S. Zeng, L. Teng, and J. Cao. "Nuclear Localization of Annexin A1 Correlates with Advanced Disease and Peritoneal Dissemination in Patients with Gastric Carcinoma." *Anat Rec (Hoboken)* 293, no. 8 (2010): 1310-4. <https://doi.org/10.1002/ar.21176>.
204. Kunkel, T. A. "The High Cost of Living. American Association for Cancer Research Special Conference: Endogenous Sources of Mutations, Fort Myers, Florida, USA, 11-15 November 1998." *Trends Genet* 15, no. 3 (1999): 93-4. <https://doi.org/10.1002/ar.21176>.
205. Kunz, B. A., A. F. Straffon, and E. J. Vonarx. "DNA Damage-Induced Mutation: Tolerance Via Translesion Synthesis." *Mutat Res* 451, no. 1-2 (2000): 169-85. [https://doi.org/10.1016/S0027-5107\(00\)00048-8](https://doi.org/10.1016/S0027-5107(00)00048-8).
206. Makridakis, N. M., and J. K. Reichardt. "Translesion DNA Polymerases and Cancer." *Front Genet* 3 (2012): 174. <https://doi.org/10.3389/fgene.2012.00174>.
207. Dieckman, L. M., B. D. Freudenthal, and M. T. Washington. "Pcna Structure and Function: Insights from Structures of Pcna Complexes and Post-Translationally Modified Pcna." *Subcell Biochem* 62 (2012): 281-99. https://doi.org/10.1007/978-94-007-4572-8_15.
208. Choi, S., R. Srivas, K. Y. Fu, B. L. Hood, B. Dost, G. A. Gibson, S. C. Watkins, B. Van Houten, N. Bandeira, T. P. Conrads, T. Ideker, and C. J. Bakkenist. "Quantitative Proteomics Reveal Atm Kinase-Dependent Exchange in DNA Damage Response Complexes." *J Proteome Res* 11, no. 10 (2012): 4983-91. <https://doi.org/10.1021/pr3005524>.
209. Hirata, F., L. M. Thibodeau, and A. Hirata. "Ubiquitination and Sumoylation of Annexin A1 and Helicase Activity." *Biochim Biophys Acta* 1800, no. 9 (2010): 899-905. <https://doi.org/10.1016/j.bbagen.2010.03.020>.
210. Ganesan, T., A. Sinniah, Z. A. Ibrahim, Z. Chik, and M. A. Alshawsh. "Annexin A1: A Bane or a Boon in Cancer? A Systematic Review." *Molecules* 25, no. 16 (2020). <https://doi.org/10.3390/molecules25163700>.
211. Zhu, F., Y. Wang, S. Zeng, X. Fu, L. Wang, and J. Cao. "Involvement of Annexin A1 in Multidrug Resistance of K562/Adr Cells Identified by the Proteomic Study." *Omics* 13, no. 6 (2009): 467-76. <https://doi.org/10.1089/omi.2009.0046>.
212. D'Acunto, C. W., H. Gbelcova, M. Festa, and T. Ruml. "The Complex Understanding of Annexin A1 Phosphorylation." *Cell Signal* 26, no. 1 (2014): 173-8. <https://doi.org/10.1016/j.cellsig.2013.09.020>.
213. Pepinsky, R. B., and L. K. Sinclair. "Epidermal Growth Factor-Dependent Phosphorylation of Lipocortin." *Nature* 321, no. 6065 (1986): 81-4. <https://doi.org/10.1038/321081a0>.
214. Skouteris, G. G., and C. H. Schröder. "The Hepatocyte Growth Factor Receptor Kinase-Mediated Phosphorylation of Lipocortin-1 Transduces the Proliferating Signal of the Hepatocyte Growth Factor." *J Biol Chem* 271, no. 44 (1996): 27266-73. <https://doi.org/10.1074/jbc.271.44.27266>.
215. Moraes, L. A., S. Kar, S. L. Foo, T. Gu, Y. Q. Toh, P. B. Ampomah, K. Sachaphibulkij, G. Yap, O. Zharkova, H. M. Lukman, A. M. Fairhurst, A. P. Kumar, and L. H. K. Lim. "Annexin-A1 Enhances Breast Cancer Growth

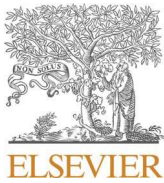
- and Migration by Promoting Alternative Macrophage Polarization in the Tumour Microenvironment." *Sci Rep* 7, no. 1 (2017): 17925. <https://doi.org/10.1038/s41598-017-17622-5>.
216. Li, Y., L. Cai, H. Wang, P. Wu, W. Gu, Y. Chen, H. Hao, K. Tang, P. Yi, M. Liu, S. Miao, and D. Ye. "Pleiotropic Regulation of Macrophage Polarization and Tumorigenesis by Formyl Peptide Receptor-2." *Oncogene* 30, no. 36 (2011): 3887-99. <https://doi.org/10.1038/onc.2011.112>.
217. Locatelli, I., S. Sutti, A. Jindal, M. Vacchiano, C. Bozzola, C. Reutelingsperger, D. Kusters, S. Bena, M. Parola, C. Paternostro, E. Bugianesi, S. McArthur, E. Albano, and M. Perretti. "Endogenous Annexin A1 Is a Novel Protective Determinant in Nonalcoholic Steatohepatitis in Mice." *Hepatology* 60, no. 2 (2014): 531-44. <https://doi.org/10.1002/hep.27141>.
218. Ferlazzo, Viviana, Pietro D'Agostino, Salvatore Milano, Rosalba Caruso, Salvatore Feo, Enrico Cillari, and Luca Parente. "Anti-Inflammatory Effects of Annexin-1: Stimulation of Il-10 Release and Inhibition of Nitric Oxide Synthesis." *International Immunopharmacology* 3, no. 10 (2003): 1363-69. [https://doi.org/10.1016/S1567-5769\(03\)00133-4](https://doi.org/10.1016/S1567-5769(03)00133-4).
219. Weyd, H., L. Abeler-Dorner, B. Linke, A. Mahr, V. Jahndel, S. Pfrang, M. Schnolzer, C. S. Falk, and P. H. Krammer. "Annexin A1 on the Surface of Early Apoptotic Cells Suppresses Cd8+ T Cell Immunity." *PLoS One* 8, no. 4 (2013): e62449. <https://doi.org/10.1371/journal.pone.0062449>.
220. Gold, R., M. Schmied, U. Tontsch, H. P. Hartung, H. Wekerle, K. V. Toyka, and H. Lassmann. "Antigen Presentation by Astrocytes Primes Rat T Lymphocytes for Apoptotic Cell Death. A Model for T-Cell Apoptosis in Vivo." *Brain* 119 (Pt 2) (1996): 651-9. <https://doi.org/10.1093/brain/119.2.651>.
221. D'Acquisto, F., A. Merghani, E. Lecona, G. Rosignoli, K. Raza, C. D. Buckley, R. J. Flower, and M. Perretti. "Annexin-1 Modulates T-Cell Activation and Differentiation." *Blood* 109, no. 3 (2007): 1095-102. <https://doi.org/10.1182/blood-2006-05-022798>.
222. Ng, F. S., K. Y. Wong, S. P. Guan, F. B. Mustafa, T. S. Kajiji, P. Bist, S. K. Biswas, W. S. Wong, and L. H. Lim. "Annexin-1-Deficient Mice Exhibit Spontaneous Airway Hyperresponsiveness and Exacerbated Allergen-Specific Antibody Responses in a Mouse Model of Asthma." *Clin Exp Allergy* 41, no. 12 (2011): 1793-803. <https://doi.org/10.1111/j.1365-2222.2011.03855.x>.
223. Kamal, A. M., R. J. Flower, and M. Perretti. "An Overview of the Effects of Annexin 1 on Cells Involved in the Inflammatory Process." *Mem Inst Oswaldo Cruz* 100 Suppl 1 (2005): 39-47. <https://doi.org/10.1590/S0074-02762005000900008>.
224. Jeppesen, D. K., A. M. Fenix, J. L. Franklin, J. N. Higginbotham, Q. Zhang, L. J. Zimmerman, D. C. Liebler, J. Ping, Q. Liu, R. Evans, W. H. Fissell, J. G. Patton, L. H. Rome, D. T. Burnette, and R. J. Coffey. "Reassessment of Exosome Composition." *Cell* 177, no. 2 (2019): 428-45 e18. <https://doi.org/10.1016/j.cell.2019.02.029>.
225. Lo Cicero, A., P. D. Stahl, and G. Raposo. "Extracellular Vesicles Shuffling Intercellular Messages: For Good or for Bad." *Curr Opin Cell Biol* 35 (2015): 69-77. <https://doi.org/10.1016/j.ceb.2015.04.013>.
226. van Niel, G., G. D'Angelo, and G. Raposo. "Shedding Light on the Cell Biology of Extracellular Vesicles." *Nat Rev Mol Cell Biol* 19, no. 4 (2018): 213-28. <https://doi.org/10.1038/nrm.2017.125>.
227. Aalberts, M., F. M. van Dissel-Emiliani, N. P. van Adrichem, M. van Wijnen, M. H. Wauben, T. A. Stout, and W. Stoorvogel. "Identification of Distinct Populations of Prostatomes That Differentially Express Prostate Stem Cell Antigen, Annexin A1, and Glipr2 in Humans." *Biol Reprod* 86, no. 3 (2012): 82. <https://doi.org/10.1095/biolreprod.111.095760>.
228. Perretti, M., D. Cooper, J. Dalli, and L. V. Norling. "Immune Resolution Mechanisms in Inflammatory Arthritis." *Nat Rev Rheumatol* 13, no. 2 (2017): 87-99. <https://doi.org/10.1038/nrrheum.2016.193>.

229. Headland, S. E., and L. V. Norling. "The Resolution of Inflammation: Principles and Challenges." *Semin Immunol* 27, no. 3 (2015): 149-60. <https://doi.org/10.1016/j.smim.2015.03.014>.
230. Gasser, O., and J. A. Schifferli. "Activated Polymorphonuclear Neutrophils Disseminate Anti-Inflammatory Microparticles by Ectocytosis." *Blood* 104, no. 8 (2004): 2543-8. <https://doi.org/10.1182/blood-2004-01-0361>.
231. Rhys, H. I., F. Dell'Accio, C. Pitzalis, A. Moore, L. V. Norling, and M. Perretti. "Neutrophil Microvesicles from Healthy Control and Rheumatoid Arthritis Patients Prevent the Inflammatory Activation of Macrophages." *EBioMedicine* 29 (2018): 60-69. <https://doi.org/10.1016/j.ebiom.2018.02.003>.
232. Yang, Y., Y. Liu, X. Yao, Y. Ping, T. Jiang, Q. Liu, S. Xu, J. Huang, H. Mou, W. Gong, K. Chen, X. Bian, and J. M. Wang. "Annexin 1 Released by Necrotic Human Glioblastoma Cells Stimulates Tumor Cell Growth through the Formyl Peptide Receptor 1." *Am J Pathol* 179, no. 3 (2011): 1504-12. <https://doi.org/10.1016/j.ajpath.2011.05.059>.
233. Link, C., F. Bujupi, P. H. Krammer, and H. Weyd. "Annexin-Coated Particles Induce Antigen-Specific Immunosuppression." *Autoimmunity* 53, no. 2 (2020): 86-94. <https://doi.org/10.1080/08916934.2019.1710134>.
234. Ampomah, Patrick B., Leonardo A. Moraes, Hakim M. Lukman, and Lina H. K. Lim. "Formyl Peptide Receptor 2 Is Regulated by Rna Mimics and Viruses through an Ifn-B-Stat3-Dependent Pathway." 32, no. 3 (2018): 1468-78. <https://doi.org/10.1096/fj.201700584RR>.
235. Oshi, M., Y. Tokumaru, S. Mukhopadhyay, L. Yan, R. Matsuyama, I. Endo, and K. Takabe. "Annexin A1 Expression Is Associated with Epithelial-Mesenchymal Transition (Emt), Cell Proliferation, Prognosis, and Drug Response in Pancreatic Cancer." *Cells* 10, no. 3 (2021). <https://doi.org/10.3390/cells10030653>.
236. Huang, S. H., Y. Li, J. Zhang, J. Rong, and S. Ye. "Epidermal Growth Factor Receptor-Containing Exosomes Induce Tumor-Specific Regulatory T Cells." *Cancer Invest* 31, no. 5 (2013): 330-5. <https://doi.org/10.3109/07357907.2013.789905>.
237. Azuma, K., K. Ota, A. Kawahara, S. Hattori, E. Iwama, T. Harada, K. Matsumoto, K. Takayama, S. Takamori, M. Kage, T. Hoshino, Y. Nakanishi, and I. Okamoto. "Association of Pd-L1 Overexpression with Activating Egfr Mutations in Surgically Resected Nonsmall-Cell Lung Cancer." *Ann Oncol* 25, no. 10 (2014): 1935-40. <https://doi.org/10.1093/annonc/mdu242>.
238. Akbay, E. A., S. Koyama, J. Carretero, A. Altabef, J. H. Tchaicha, C. L. Christensen, O. R. Mikse, A. D. Cherniack, E. M. Beauchamp, T. J. Pugh, M. D. Wilkerson, P. E. Fecci, M. Butaney, J. B. Reibel, M. Soucheray, T. J. Cohoon, P. A. Janne, M. Meyerson, D. N. Hayes, G. I. Shapiro, T. Shimamura, L. M. Sholl, S. J. Rodig, G. J. Freeman, P. S. Hammerman, G. Dranoff, and K. K. Wong. "Activation of the Pd-1 Pathway Contributes to Immune Escape in Egfr-Driven Lung Tumors." *Cancer Discov* 3, no. 12 (2013): 1355-63. <https://doi.org/10.1158/2159-8290.CD-13-0310>.
239. Chen, N., W. Fang, J. Zhan, S. Hong, Y. Tang, S. Kang, Y. Zhang, X. He, T. Zhou, T. Qin, Y. Huang, X. Yi, and L. Zhang. "Upregulation of Pd-L1 by Egfr Activation Mediates the Immune Escape in Egfr-Driven Nscl: Implication for Optional Immune Targeted Therapy for Nscl Patients with Egfr Mutation." *J Thorac Oncol* 10, no. 6 (2015): 910-23. <https://doi.org/10.1097/JTO.0000000000000500>.
240. Li, T., C. Zhang, G. Zhao, X. Zhang, M. Hao, S. Hassan, M. Zhang, H. Zheng, D. Yang, L. Liu, F. Mehraein-Ghomi, X. Bai, K. Chen, W. Zhang, and J. Yang. "Igfbp2 Regulates Pd-L1 Expression by Activating the Egfr-Stat3 Signaling Pathway in Malignant Melanoma." *Cancer Lett* 477 (2020): 19-30. <https://doi.org/10.1016/j.canlet.2020.02.036>.

241. Lo, H. W., S. C. Hsu, M. Ali-Seyed, M. Gunduz, W. Xia, Y. Wei, G. Bartholomeusz, J. Y. Shih, and M. C. Hung. "Nuclear Interaction of Egfr and Stat3 in the Activation of the Inos/No Pathway." *Cancer Cell* 7, no. 6 (2005): 575-89. <https://doi.org/10.1016/j.ccr.2005.05.007>.
242. Bronte, V., T. Kasic, G. Gri, K. Gallana, G. Borsellino, I. Marigo, L. Battistini, M. Iafrate, T. Prayer-Galetti, F. Pagano, and A. Viola. "Boosting Antitumor Responses of T Lymphocytes Infiltrating Human Prostate Cancers." *J Exp Med* 201, no. 8 (2005): 1257-68. <https://doi.org/10.1084/jem.20042028>.
243. Brand, T. M., M. Iida, N. Luthar, M. M. Starr, E. J. Huppert, and D. L. Wheeler. "Nuclear Egfr as a Molecular Target in Cancer." *Radiother Oncol* 108, no. 3 (2013): 370-7. <https://doi.org/10.1016/j.radonc.2013.06.010>.
244. Eden, E. R., E. Sanchez-Heras, A. Tsapara, A. Sobota, T. P. Levine, and C. E. Futter. "Annexin A1 Tethers Membrane Contact Sites That Mediate Er to Endosome Cholesterol Transport." *Dev Cell* 37, no. 5 (2016): 473-83. <https://doi.org/10.1016/j.devcel.2016.05.005>.
245. Zhao, X., W. Ma, X. Li, H. Li, J. Li, H. Li, and F. He. "Anxa1 Enhances Tumor Proliferation and Migration by Regulating Epithelial-Mesenchymal Transition and Il-6/Jak2/Stat3 Pathway in Papillary Thyroid Carcinoma." *J Cancer* 12, no. 5 (2021): 1295-306. <https://doi.org/10.7150/jca.52171>.
246. Stenfeldt, A.L., J. Karlsson, C. Wenneras, J. Bylund, and C. Dahlgren. "Cyclosporin H, Boc-MLF and Boc-FLFLF are antagonists that preferentially inhibit activity triggered through the formyl peptide receptor." *Inflammation* 6 (2007):224-9. <https://doi.org/10.1007/s10753-007-9040-4>.

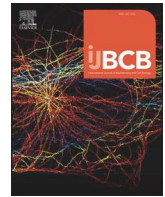
Capítulo 2

Annexin A1 promotes the nuclear localization of the epidermal growth factor receptor in castration-resistant prostate cancer



Contents lists available at ScienceDirect

International Journal of Biochemistry and Cell Biology

journal homepage: www.elsevier.com/locate/biociel

Annexin A1 promotes the nuclear localization of the epidermal growth factor receptor in castration-resistant prostate cancer

Sara Teixeira Soares Mota^{a,b,1}, Lara Vecchi^{b,*,1}, Douglas Alexsander Alves^{a,b},
 Antonielle Oliveira Cordeiro^{a,b}, Gabriela Silva Guimarães^{a,b}, Esther Campos-Fernández^b,
 Yara Cristina Paiva Maia^c, Bruno de Carvalho Dornelas^d, Stephania Martins Bezerra^e,
 Victor Piana de Andrade^e, Luiz Ricardo Goulart^{a,f}, Thaise Gonçalves Araújo^{a,b,*}

^a Laboratory of Genetics and Biotechnology, Institute of Biotechnology, Federal University of Uberlândia, Patos de Minas, MG, 387400-128, Brazil

^b Laboratory of Nanobiotechnology, Institute of Biotechnology, Federal University of Uberlândia, Uberlândia, MG, 38400-902, Brazil

^c Medical Faculty, Federal University of Uberlândia, Uberlândia, MG, 38400-902, Brazil

^d Pathology Division, Internal Medicine, University Hospital, Federal University of Uberlândia, Uberlândia, MG, 38400-902, Brazil

^e AC Camargo Cancer Hospital, São Paulo, SP, 01509-010, Brazil

^f University of California, Davis, Dept. of Medical Microbiology and Immunology, Davis, CA, 95616, USA

ARTICLE INFO

Keywords:

Epidermal growth factor receptor
 Annexin A1
 Formyl peptide receptor 1
 Cyclosporin H
 Prostate cancer
 Castration resistant

ABSTRACT

Epidermal growth factor receptor is a cancer driver whose nuclear localization has been associated with the progression of prostate cancer to the castration-resistant phenotype. Previous reports indicated a functional interaction between this receptor and the protein Annexin A1, which has also been associated with aggressive tumors. The molecular pathogenesis of castration-resistant prostate cancer remains largely unresolved, and herein we have demonstrated the correlation between the expression levels and localization of the epidermal growth factor receptor and Annexin A1 in prostate cancer samples and cell lines. Interestingly, a higher expression of both proteins was detected in castration-resistant prostate cancer cell lines and the strongest correlation was seen at the nuclear level. We verified that Annexin A1 interacts with the epidermal growth factor receptor, and by using prostate cancer cell lines knocked down for Annexin A1, we succeeded in demonstrating that Annexin A1 promotes the nuclear localization of epidermal growth factor receptor. Finally, we showed that Annexin A1 activates an autocrine signaling in castration-resistant prostate cells through the formyl peptide receptor 1. The inhibition of such signaling by Cyclosporin H inhibits the nuclear localization of epidermal growth factor receptor and its downstream signaling. The present work sheds light on the functional interaction between nuclear epidermal growth factor receptor and nuclear Annexin A1 in castration-resistant prostate cancer. Therefore, strategies to inhibit the nuclear localization of epidermal growth factor receptor through the suppression of the Annexin A1 autocrine loop could represent an important intervention strategy for castration-resistant prostate cancer.

1. Introduction

Prostate cancer (PCa) represents the second most common type of cancer in men and the second leading cause of cancer death among

males in the USA and Brazil (Rawla, 2019). In 2018, 1,276,106 new cases of PCa worldwide were reported (Bray et al., 2018). It is well-known that PCa is a highly heterogeneous disease whose subtypes are poorly understood, so far (Kaffenberger and Barbieri, 2016). Patients

* Corresponding authors at: Laboratory of Nanobiotechnology, Institute of Genetics and Biochemistry, Federal University of Uberlândia, Campus Umuarama, Bloco2E, Sala 248, 38400-902, Uberlândia, MG, Brazil.

E-mail addresses: saratsm.s@hotmail.com (S.T.S. Mota), laravecchi7@yahoo.it (L. Vecchi), douglasalexanderptu@hotmail.com (D.A. Alves), antonielle._@hotmail.com (A.O. Cordeiro), gabisg9@gmail.com (G.S. Guimarães), esther.campos.fernandez@gmail.com (E. Campos-Fernández), yara.maia@ufu.br (Y.C.P. Maia), dornelasbruno@hotmail.com (B.C. Dornelas), stephania.bezerra@accamargo.org.br (S.M. Bezerra), victor.andrade@accamargo.org.br (V.P. de Andrade), lrgoulart@ufu.br (L.R. Goulart), tgaraújo@ufu.br (T.G. Araújo).

¹ Both authors contributed equally to this work.

<https://doi.org/10.1016/j.biociel.2020.105838>

Received 15 April 2020; Received in revised form 30 July 2020; Accepted 20 August 2020

Available online 25 August 2020

1357-2725/© 2020 Elsevier Ltd. All rights reserved.

Aaltoma, S. H., Lipponen, P. K. & Kosma, V. M. (2001). Inducible nitric oxide synthase (iNOS) expression and its prognostic value in prostate cancer. *Anticancer Res*, 21, 3101-3106.

Araujo, T. G., Marangoni, K., Rocha, R. M., Maia, Y. C., Araujo, G. R., Alcantar, T. M., Alves, P. T., Calabria, L., Neves, A. F., Soares, F. A. & Goulart, L. R. (2014). Dynamic dialog between cytokeratin 18 and annexin A1 in breast cancer: a transcriptional disequilibrium. *Acta Histochem*, 116, 1178-1184.

<https://doi.org/10.1016/j.acthis.2014.06.008>

Armstrong, A. J. (2014). Biomarkers in castration-resistant prostate cancer. *Clin Adv Hematol Oncol*, 12, 115-118.

Ayuso-Sacido, A., Moliterno, J. A., Kratovac, S., Kapoor, G. S., O'Rourke, D. M., Holland, E. C., Garcia-Verdugo, J. M., Roy, N. S. & Boockvar, J. A. (2010). Activated EGFR signaling increases proliferation, survival, and migration and blocks neuronal differentiation in post-natal neural stem cells. *J Neurooncol*, 97, 323-337.

<https://doi.org/10.1007/s11060-009-0035-x>

Bai, S., Cao, S., Jin, L., Kobelski, M., Schouest, B., Wang, X., Ungerleider, N., Baddoo, M., Zhang, W., Corey, E., Vessella, R. L., Dong, X., Zhang, K., Yu, X., Flemington, E. K. & Dong, Y. (2019). A positive role of c-Myc in regulating androgen receptor and its splice variants in prostate cancer. *Oncogene*, 38, 4977-4989.

<https://doi.org/10.1038/s41388-019-0768-8>

Bai, X. F., Ni, X. G., Zhao, P., Liu, S. M., Wang, H. X., Guo, B., Zhou, L. P., Liu, F., Zhang, J. S., Wang, K., Xie, Y. Q., Shao, Y. F. & Zhao, X. H. (2004). Overexpression of annexin 1 in pancreatic cancer and its clinical significance. *World J Gastroenterol*, 10, 1466-1470.

<https://doi.org/10.3748/wjg.v10.i10.1466>

Belvedere, R., Bizzarro, V., Popolo, A., Dal Piaz, F., Vasaturo, M., Picardi, P., Parente, L. & Petrella, A. (2014). Role of intracellular and extracellular annexin A1 in migration and invasion of human pancreatic carcinoma cells. *BMC Cancer*, 14, 961.

<https://doi.org/10.1186/1471-2407-14-961>

Bernard, D., Pourtier-Manzanedo, A., Gil, J. & Beach, D. H. (2003). Myc confers androgen-independent prostate cancer cell growth. *J Clin Invest*, 112, 1724-1731.

<https://doi.org/10.1172/JCI200319035>

Biaoxue, R., Shuanying, Y., Wei, L., Zongjuan, M., Xiguang, C. & Qihong, Z. (2014a). Co-overexpression of Hsp90-beta and annexin A1 with a significantly positive correlation contributes to the diagnosis of lung cancer. *Expert Rev Mol Diagn*, 14, 1067-1079.

<https://doi.org/10.1586/14737159.2014.960517>

Biaoxue, R., Xiguang, C. & Shuanying, Y. (2014b). Annexin A1 in malignant tumors: current opinions and controversies. *Int J Biol Markers*, 29, e8-20.

<https://doi.org/10.5301/jbm.5000046>

Bizzarro, V., Belvedere, R., Migliaro, V., Romano, E., Parente, L. & Petrella, A. (2017). Hypoxia regulates ANXA1 expression to support prostate cancer cell invasion and aggressiveness. *Cell Adh Migr*, 11, 247-260.

<https://doi.org/10.1080/19336918.2016.1259056>

Bizzarro, V., Belvedere, R., Milone, M. R., Pucci, B., Lombardi, R., Bruzzese, F., Popolo, A., Parente, L., Budillon, A. & Petrella, A. (2015). Annexin A1 is involved in the acquisition and maintenance of a stem cell-like/aggressive phenotype in prostate cancer cells with acquired resistance to zoledronic acid. *Oncotarget*, 6, 25076-25092.

<https://doi.org/10.18632/oncotarget.4725>

Boegemann, M., Schrader, A. J., Krabbe, L. M. & Herrmann, E. (2015). Present, Emerging and Possible Future Biomarkers in Castration Resistant Prostate Cancer (CRPC). *Curr Cancer Drug Targets*, 15, 243-255.

<https://doi.org/10.2174/1568009615666150204145803>

Boer, J. C., Domanska, U. M., Timmer-Bosscha, H., Boer, I. G., de Haas, C. J., Joseph, J. V., Kruyt, F. A., de Vries, E. G., den Dunnen, W. F., van Strijp, J. A. & Walenkamp, A. M. (2013). Inhibition of formyl peptide receptor in high-grade astrocytoma by Chemotaxis Inhibitory Protein of *S. aureus*. *Br J Cancer*, 108, 587-596.

<https://doi.org/10.1038/bjc.2012.603>

Brand, T. M., Iida, M., Luthar, N., Starr, M. M., Huppert, E. J. & Wheeler, D. L. (2013). Nuclear EGFR as a molecular target in cancer. *Radiother Oncol*, 108, 370-377.

<https://doi.org/10.1016/j.radonc.2013.06.010>

Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R. L., Torre, L. A. & Jemal, A. (2018). Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin*, 68, 394-424.

<https://doi.org/10.3322/caac.21492>

Buckingham, J. C., John, C. D., Solito, E., Tierney, T., Flower, R. J., Christian, H. & Morris, J. (2006). Annexin 1, glucocorticoids, and the neuroendocrine-immune interface. *Ann N Y Acad Sci*, 1088, 396-409.

<https://doi.org/10.1196/annals.1366.002>

Chang, Y. S., Chen, W. Y., Yin, J. J., Sheppard-Tillman, H., Huang, J. & Liu, Y. N. (2015). EGF Receptor Promotes Prostate Cancer Bone Metastasis by Downregulating miR-1 and Activating TWIST1. *Cancer Res*, 75, 3077-3086.

<https://doi.org/10.1158/0008-5472.CAN-14-3380>

Crunkhorn, S. (2018). Cancer: Combating resistance to EGFR inhibitors. *Nat Rev Drug Discov*, 18, 18.

<https://doi.org/10.1038/nrd.2018.232>

Dalli, J., Montero-Melendez, T., McArthur, S. & Perretti, M. (2012). Annexin A1 N-terminal derived Peptide ac2-26 exerts chemokinetic effects on human neutrophils. *Front Pharmacol*, 3, 28.

<https://doi.org/10.3389/fphar.2012.00028>

Dang, C. V. (2012). MYC on the path to cancer. *Cell*, 149, 22-35.

<https://doi.org/10.1016/j.cell.2012.03.003>

Davies, A., Conteduca, V., Zoubeydi, A. & Beltran, H. (2019). Biological Evolution of Castration-resistant Prostate Cancer. *Eur Urol Focus*, 5, 147-154.

<https://doi.org/10.1016/j.euf.2019.01.016>

Delos Santos, R. C., Bautista, S., Lucarelli, S., Bone, L. N., Dayam, R. M., Abousawan, J., Botelho, R. J. & Antonescu, C. N. (2017). Selective regulation of clathrin-mediated epidermal growth factor receptor signaling and endocytosis by phospholipase C and calcium. *Mol Biol Cell*, 28, 2802-2818.

<https://doi.org/10.1091/mbc.e16-12-0871>

Dittmann, K., Mayer, C., Fehrenbacher, B., Schaller, M., Raju, U., Milas, L., Chen, D. J., Kehlbach, R. & Rodemann, H. P. (2005). Radiation-induced epidermal growth factor receptor nuclear import is linked to activation of DNA-dependent protein kinase. *J Biol Chem*, 280, 31182-31189.

<https://doi.org/10.1074/jbc.M506591200>

El Sheikh, S. S., Domin, J., Abel, P., Stamp, G. & Lalani el, N. (2004). Phosphorylation of both EGFR and ErbB2 is a reliable predictor of prostate cancer cell proliferation in response to EGF. *Neoplasia*, 6, 846-853.

<https://doi.org/10.1593/neo.04379>

Er, E. E., Mendoza, M. C., Mackey, A. M., Rameh, L. E. & Blenis, J. (2013). AKT facilitates EGFR trafficking and degradation by phosphorylating and activating PIKfyve. *Sci Signal*, 6, ra45.

<https://doi.org/10.1126/scisignal.2004015>

Erlandsson, A., Carlsson, J., Andersson, S. O., Vyas, C., Wikstrom, P., Andren, O., Davidsson, S. & Rider, J. R. (2018). High inducible nitric oxide synthase in prostate tumor epithelium is associated with lethal prostate cancer. *Scand J Urol*, 52, 129-133.

<https://doi.org/10.1080/21681805.2017.1421261>

Faria, J., de Andrade, C., Goes, A. M., Rodrigues, M. A. & Gomes, D. A. (2016). Effects of different ligands on epidermal growth factor receptor (EGFR) nuclear translocation. *Biochem Biophys Res Commun*, 478, 39-45.

<https://doi.org/10.1016/j.bbrc.2016.07.097>

Feng, Q. & He, B. (2019). Androgen Receptor Signaling in the Development of Castration-Resistant Prostate Cancer. *Front Oncol*, 9, 858.

<https://doi.org/10.3389/fonc.2019.00858>

Foj, L. & Filella, X. (2019). Identification of Potential miRNAs Biomarkers for High-Grade Prostate Cancer by Integrated Bioinformatics Analysis. *Pathol Oncol Res*, 25, 1445-1456.

<https://doi.org/10.1007/s12253-018-0508-3>

Futter, C. E. & White, I. J. (2007). Annexins and endocytosis. *Traffic*, 8, 951-958.

<https://doi.org/10.1111/j.1600-0854.2007.00590.x>

Gan, Y., Shi, C., Inge, L., Hibner, M., Balducci, J. & Huang, Y. (2010). Differential roles of ERK and Akt pathways in regulation of EGFR-mediated signaling and motility in prostate cancer cells. *Oncogene*, 29, 4947-4958.

<https://doi.org/10.1038/onc.2010.240>

Geary, L. A., Nash, K. A., Adisetiyo, H., Liang, M., Liao, C. P., Jeong, J. H., Zandi, E. & Roy-Burman, P. (2014). CAF-secreted annexin A1 induces prostate cancer cells to gain stem cell-like features. *Mol Cancer Res*, 12, 607-621.

<https://doi.org/10.1158/1541-7786.MCR-13-0469>

- Girol, A. P., Mimura, K. K., Drewes, C. C., Bolonheis, S. M., Solito, E., Farsky, S. H., Gil, C. D. & Oliani, S. M. (2013). Anti-inflammatory mechanisms of the annexin A1 protein and its mimetic peptide Ac2-26 in models of ocular inflammation in vivo and in vitro. *J Immunol*, 190, 5689-5701.
<https://doi.org/10.4049/jimmunol.1202030>
- Gravis, G. (2019). Systemic treatment for metastatic prostate cancer. *Asian J Urol*, 6, 162-168.
<https://doi.org/10.1016/j.ajur.2019.02.002>
- Guerin, O., Fischel, J. L., Ferrero, J. M., Bozec, A. & Milano, G. (2010). EGFR Targeting in Hormone-Refractory Prostate Cancer: Current Appraisal and Prospects for Treatment. *Pharmaceuticals (Basel)*, 3, 2238-2247.
<https://doi.org/10.3390/ph3072238>
- Guzel, E., Karatas, O. F., Duz, M. B., Solak, M., Ittmann, M. & Ozen, M. (2014). Differential expression of stem cell markers and ABCG2 in recurrent prostate cancer. *Prostate*, 74, 1498-1505.
<https://doi.org/10.1002/pros.22867>
- Han, E. K., Lim, J. T., Arber, N., Rubin, M. A., Xing, W. Q. & Weinstein, I. B. (1998). Cyclin D1 expression in human prostate carcinoma cell lines and primary tumors. *Prostate*, 35, 95-101.
[https://doi.org/10.1002/\(SICI\)1097-0045\(19980501\)35:2<95::AID-PROS2>3.0.CO;2-F](https://doi.org/10.1002/(SICI)1097-0045(19980501)35:2<95::AID-PROS2>3.0.CO;2-F)
- Han, G., Tian, Y., Duan, B., Sheng, H., Gao, H. & Huang, J. (2014). Association of nuclear annexin A1 with prognosis of patients with esophageal squamous cell carcinoma. *Int J Clin Exp Pathol*, 7, 751-759.
- Han, W., Carpenter, R. L., Cao, X. & Lo, H. W. (2013). STAT1 gene expression is enhanced by nuclear EGFR and HER2 via cooperation with STAT3. *Mol Carcinog*, 52, 959-969.
<https://doi.org/10.1002/mc.21936>
- Hanada, N., Lo, H. W., Day, C. P., Pan, Y., Nakajima, Y. & Hung, M. C. (2006). Co-regulation of B-Myb expression by E2F1 and EGF receptor. *Mol Carcinog*, 45, 10-17.
<https://doi.org/10.1002/mc.20147>
- Hasegawa, N., Mizutani, K., Suzuki, T., Deguchi, T. & Nozawa, Y. (2006). A comparative study of protein profiling by proteomic analysis in camptothecin-resistant PC3 and camptothecin-sensitive LNCaP human prostate cancer cells. *Urol Int*, 77, 347-354.
<https://doi.org/10.1159/000096340>
- Herbst, R. S. (2004). Review of epidermal growth factor receptor biology. *Int J Radiat Oncol Biol Phys*, 59, 21-26.
<https://doi.org/10.1016/j.ijrobp.2003.11.041>
- Herrera, F. G. & Berthold, D. R. (2016). Radiation Therapy after Radical Prostatectomy: Implications for Clinicians. *Front Oncol*, 6, 117.
<https://doi.org/10.3389/fonc.2016.00117>
- Hong, J., Wuest, T. R., Min, Y. & Lin, P. C. (2019). Oxygen tension regulates lysosomal activation and receptor tyrosine kinase degradation. *bioRxiv*, 727495.
<https://doi.org/10.3390/cancers11111653>

Hsieh, A. L., Walton, Z. E., Altman, B. J., Stine, Z. E. & Dang, C. V. (2015). MYC and metabolism on the path to cancer. *Semin Cell Dev Biol*, 43, 11-21.

<https://doi.org/10.1016/j.semcdb.2015.08.003>

Hsu, S. C., Miller, S. A., Wang, Y. & Hung, M. C. (2009). Nuclear EGFR is required for cisplatin resistance and DNA repair. *Am J Transl Res*, 1, 249-258.

Huang, W. C., Chen, Y. J., Li, L. Y., Wei, Y. L., Hsu, S. C., Tsai, S. L., Chiu, P. C., Huang, W. P., Wang, Y. N., Chen, C. H., Chang, W. C., Chang, W. C., Chen, A. J., Tsai, C. H. & Hung, M. C. (2011). Nuclear translocation of epidermal growth factor receptor by Akt-dependent phosphorylation enhances breast cancer-resistant protein expression in gefitinib-resistant cells. *J Biol Chem*, 286, 20558-20568.

<https://doi.org/10.1074/jbc.M111.240796>

Hung, L. Y., Tseng, J. T., Lee, Y. C., Xia, W., Wang, Y. N., Wu, M. L., Chuang, Y. H., Lai, C. H. & Chang, W. C. (2008). Nuclear epidermal growth factor receptor (EGFR) interacts with signal transducer and activator of transcription 5 (STAT5) in activating Aurora-A gene expression. *Nucleic Acids Res*, 36, 4337-4351.

<https://doi.org/10.1093/nar/gkn417>

Inokuchi, J., Lau, A., Tyson, D. R. & Ornstein, D. K. (2009). Loss of annexin A1 disrupts normal prostate glandular structure by inducing autocrine IL-6 signaling. *Carcinogenesis*, 30, 1082-1088.

<https://doi.org/10.1093/carcin/bgp078>

Isbarn, H., Boccon-Gibod, L., Carroll, P. R., Montorsi, F., Schulman, C., Smith, M. R., Sternberg, C. N. & Studer, U. E. (2009). Androgen deprivation therapy for the treatment of prostate cancer: consider both benefits and risks. *Eur Urol*, 55, 62-75.

<https://doi.org/10.1016/j.eururo.2008.10.008>

Ishikawa, T., Saito, H., Hirano, H., Inoue, Y. & Ikegami, Y. (2012). Human ABC transporter ABCG2 in cancer chemotherapy: drug molecular design to circumvent multidrug resistance. *Methods Mol Biol*, 910, 267-278.

https://doi.org/10.1007/978-1-61779-965-5_11

Jaganathan, S., Yue, P., Paladino, D. C., Bogdanovic, J., Huo, Q. & Turkson, J. (2011). A functional nuclear epidermal growth factor receptor, SRC and Stat3 heteromeric complex in pancreatic cancer cells. *PLoS One*, 6, e19605.

<https://doi.org/10.1371/journal.pone.0019605>

Jernberg, E., Bergh, A. & Wikstrom, P. (2017). Clinical relevance of androgen receptor alterations in prostate cancer. *Endocr Connect*, 6, R146-R161.

<https://doi.org/10.1530/EC-17-0118>

John, R. R., Malathi, N., Ravindran, C. & Anandan, S. (2017). Mini review: Multifaceted role played by cyclin D1 in tumor behavior. *Indian J Dent Res*, 28, 187-192.

https://doi.org/10.4103/ijdr.IJDR_697_16

Jones, D., Noble, M., Wedge, S. R., Robson, C. N. & Gaughan, L. (2017). Aurora A regulates expression of AR-V7 in models of castrate resistant prostate cancer. *Sci Rep*, 7, 40957.

<https://doi.org/10.1038/srep40957>

Kaffenberger, S. D. & Barbieri, C. E. (2016). Molecular subtyping of prostate cancer. *Curr Opin Urol*, 26, 213-218.

<https://doi.org/10.1097/MOU.0000000000000285>

Kim, J. H., Xu, C., Keum, Y. S., Reddy, B., Conney, A. & Kong, A. N. (2006). Inhibition of EGFR signaling in human prostate cancer PC-3 cells by combination treatment with beta-phenylethyl isothiocyanate and curcumin. *Carcinogenesis*, 27, 475-482.

<https://doi.org/10.1093/carcin/bgi272>

Kim, J. K., Kim, P. J., Jung, K. H., Noh, J. H., Eun, J. W., Bae, H. J., Xie, H. J., Shan, J. M., Ping, W. Y., Park, W. S., Lee, J. Y. & Nam, S. W. (2010). Decreased expression of annexin A10 in gastric cancer and its overexpression in tumor cell growth suppression. *Oncol Rep*, 24, 607-612.

<https://doi.org/10.3892/or.00000898>

Ku, S. Y., Gleave, M. E. & Beltran, H. (2019). Towards precision oncology in advanced prostate cancer. *Nat Rev Urol*.

<https://doi.org/10.1038/s41585-019-0237-8>

Li, C. F., Fang, F. M., Wang, J. M., Tzeng, C. C., Tai, H. C., Wei, Y. C., Li, S. H., Lee, Y. T., Wang, Y. H., Yu, S. C., Shiue, Y. L., Chu, P. Y., Wang, W. L., Chen, L. T. & Huang, H. Y. (2012). EGFR nuclear import in gallbladder carcinoma: nuclear phosphorylated EGFR upregulates iNOS expression and confers independent prognostic impact. *Ann Surg Oncol*, 19, 443-454.

<https://doi.org/10.1245/s10434-011-1942-6>

Li, X., Huang, Y., Jiang, J. & Frank, S. J. (2008). ERK-dependent threonine phosphorylation of EGF receptor modulates receptor downregulation and signaling. *Cell Signal*, 20, 2145-2155.

<https://doi.org/10.1016/j.cellsig.2008.08.006>

Li, X., Zhao, Y., Xia, Q., Zheng, L., Liu, L., Zhao, B. & Shi, J. (2016). Nuclear translocation of annexin 1 following oxygen-glucose deprivation-reperfusion induces apoptosis by regulating Bid expression via p53 binding. *Cell Death Dis*, 7, e2356.

<https://doi.org/10.1038/cddis.2016.259>

Liao, W. I., Wu, S. Y., Wu, G. C., Pao, H. P., Tang, S. E., Huang, K. L. & Chu, S. J. (2017). Ac2-26, an Annexin A1 Peptide, Attenuates Ischemia-Reperfusion-Induced Acute Lung Injury. *Int J Mol Sci*, 18.

<https://doi.org/10.3390/ijms18081771>

Liao, Y., Guo, Z., Xia, X., Liu, Y., Huang, C., Jiang, L., Wang, X., Liu, J. & Huang, H. (2019). Inhibition of EGFR signaling with Spautin-1 represents a novel therapeutics for prostate cancer. *J Exp Clin Cancer Res*, 38, 157.

<https://doi.org/10.1186/s13046-019-1165-4>

Lin, S. Y., Makino, K., Xia, W., Matin, A., Wen, Y., Kwong, K. Y., Bourguignon, L. & Hung, M. C. (2001). Nuclear localization of EGF receptor and its potential new role as a transcription factor. *Nat Cell Biol*, 3, 802-808.

<https://doi.org/10.1038/ncb0901-802>

Lo, H. W., Cao, X., Zhu, H. & Ali-Osman, F. (2010). Cyclooxygenase-2 is a novel transcriptional target of the nuclear EGFR-STAT3 and EGFRvIII-STAT3 signaling axes. *Mol Cancer Res*, 8, 232-245.

<https://doi.org/10.1158/1541-7786.MCR-09-0391>

Lo, H. W., Hsu, S. C., Ali-Seyed, M., Gunduz, M., Xia, W., Wei, Y., Bartholomeusz, G., Shih, J. Y. & Hung, M. C. (2005). Nuclear interaction of EGFR and STAT3 in the activation of the iNOS/NO pathway. *Cancer Cell*, 7, 575-589.

<https://doi.org/10.1016/j.ccr.2005.05.007>

Maione, P., Sacco, P. C., Casaluce, F., Sgambato, A., Santabarbara, G., Rossi, A. & Gridelli, C. (2016). Overcoming Resistance to EGFR Inhibitors in NSCLC. *Rev Recent Clin Trials*, 11, 99-105.

<https://doi.org/10.2174/1574887111666160330120431>

Mitchell, R. A., Luwor, R. B. & Burgess, A. W. (2018). Epidermal growth factor receptor: Structure-function informing the design of anticancer therapeutics. *Exp Cell Res*, 371, 1-19.

<https://doi.org/10.1016/j.yexcr.2018.08.009>

Montgomery, R. B., Mostaghel, E. A., Vessella, R., Hess, D. L., Kalthorn, T. F., Higano, C. S., True, L. D. & Nelson, P. S. (2008). Maintenance of intratumoral androgens in metastatic prostate cancer: a mechanism for castration-resistant tumor growth. *Cancer Res*, 68, 4447-4454.

<https://doi.org/10.1158/0008-5472.CAN-08-0249>

Mota, S. T. S., Vecchi, L., Zoia, M. A. P., Oliveira, F. M., Alves, D. A., Dornelas, B. C., Bezerra, S. M., Andrade, V. P., Maia, Y. C. P., Neves, A. F., Goulart, L. R. & Araujo, T. G. (2019). New Insights into the Role of Polybromo-1 in Prostate Cancer. *Int J Mol Sci*, 20.

<https://doi.org/10.3390/ijms20122852>

Mottet, N., Bellmunt, J., Bolla, M., Briers, E., Cumberbatch, M. G., De Santis, M., Fossati, N., Gross, T., Henry, A. M., Joniau, S., Lam, T. B., Mason, M. D., Matveev, V. B., Moldovan, P. C., van den Bergh, R. C. N., Van den Broeck, T., van der Poel, H. G., van der Kwast, T. H., Rouviere, O., Schoots, I. G., Wiegel, T. & Cornford, P. (2017). EAU-ESTRO-SIOG Guidelines on Prostate Cancer. Part 1: Screening, Diagnosis, and Local Treatment with Curative Intent. *Eur Urol*, 71, 618-629.

<https://doi.org/10.1016/j.eururo.2016.08.003>

Natarajan, K., Xie, Y., Baer, M. R. & Ross, D. D. (2012). Role of breast cancer resistance protein (BCRP/ABCG2) in cancer drug resistance. *Biochem Pharmacol*, 83, 1084-1103.

<https://doi.org/10.1016/j.bcp.2012.01.002>

Nesterov, A., Kurten, R. C. & Gill, G. N. (1995). Association of epidermal growth factor receptors with coated pit adaptins via a tyrosine phosphorylation-regulated mechanism. *J Biol Chem*, 270, 6320-6327.

<https://doi.org/10.1074/jbc.270.11.6320>

Normanno, N., De Luca, A., Bianco, C., Strizzi, L., Mancino, M., Maiello, M. R., Carotenuto, A., De Feo, G., Caponigro, F. & Salomon, D. S. (2006). Epidermal growth factor receptor (EGFR) signaling in cancer. *Gene*, 366, 2-16.

<https://doi.org/10.1016/j.gene.2005.10.018>

Otto, T. & Sicinski, P. (2017). Cell cycle proteins as promising targets in cancer therapy. *Nat Rev Cancer*, 17, 93-115.

<https://doi.org/10.1038/nrc.2016.138>

Pascal, L. E., Oudes, A. J., Petersen, T. W., Goo, Y. A., Walashek, L. S., True, L. D. & Liu, A. Y. (2007). Molecular and cellular characterization of ABCG2 in the prostate. *BMC Urol*, 7, 6.

<https://doi.org/10.1186/1471-2490-7-6>

Patton, K. T., Chen, H. M., Joseph, L. & Yang, X. J. (2005). Decreased annexin I expression in prostatic adenocarcinoma and in high-grade prostatic intraepithelial neoplasia. *Histopathology*, 47, 597-601.

<https://doi.org/10.1111/j.1365-2559.2005.02300.x>

Pereira, R. A., Ravinal, R. C., Costa, R. S., Lima, M. S., Tucci, S., Muglia, V. F., Reis, R. B. & Silva, G. E. (2014). Cyclin D1 expression in prostate carcinoma. *Braz J Med Biol Res*, 47, 515-521.

<https://doi.org/10.1590/1414-431X20143240>

Perlmutter, M. A. & Lepor, H. (2007). Androgen deprivation therapy in the treatment of advanced prostate cancer. *Rev Urol*, 9 Suppl 1, S3-8.

Perretti, M. & D'Acquisto, F. (2009). Annexin A1 and glucocorticoids as effectors of the resolution of inflammation. *Nat Rev Immunol*, 9, 62-70.

<https://doi.org/10.1038/nri2470>

Petrylak, D. P. & Crawford, E. D. (2017). Biomarkers for the Management of Castration-Resistant Prostate Cancer: We Are Not There Yet. *Target Oncol*, 12, 401-412.

<https://doi.org/10.1007/s11523-017-0500-y>

Psyrri, A., Kassar, M., Yu, Z., Bamias, A., Weinberger, P. M., Markakis, S., Kowalski, D., Camp, R. L., Rimm, D. L. & Dimopoulos, M. A. (2005a). Effect of epidermal growth factor receptor expression level on survival in patients with epithelial ovarian cancer. *Clin Cancer Res*, 11, 8637-8643.

<https://doi.org/10.1158/1078-0432.CCR-05-1436>

Psyrri, A., Yu, Z., Weinberger, P. M., Sasaki, C., Haffty, B., Camp, R., Rimm, D. & Burtneess, B. A. (2005b). Quantitative determination of nuclear and cytoplasmic epidermal growth factor receptor expression in oropharyngeal squamous cell cancer by using automated quantitative analysis. *Clin Cancer Res*, 11, 5856-5862.

<https://doi.org/10.1158/1078-0432.CCR-05-0420>

Raulf, N., Lucarelli, P., Thavaraj, S., Brown, S., Vicencio, J. M., Sauter, T. & Tavassoli, M. (2018). Annexin A1 regulates EGFR activity and alters EGFR-containing tumour-derived exosomes in head and neck cancers. *Eur J Cancer*, 102, 52-68.

<https://doi.org/10.1016/j.ejca.2018.07.123>

Rawla, P. (2019). Epidemiology of Prostate Cancer. *World J Oncol*, 10, 63-89.

<https://doi.org/10.14740/wjon1191>

Rescher, U., Goebeler, V., Wilbers, A. & Gerke, V. (2006). Proteolytic cleavage of annexin 1 by human leukocyte elastase. *Biochim Biophys Acta*, 1763, 1320-1324.

<https://doi.org/10.1016/j.bbamcr.2006.08.041>

Roach, M., 3rd (2014). Current trends for the use of androgen deprivation therapy in conjunction with radiotherapy for patients with unfavorable intermediate-risk, high-risk, localized, and locally advanced prostate cancer. *Cancer*, 120, 1620-1629.

<https://doi.org/10.1002/cncr.28594>

Rodrigues-Lisoni, F. C., Mehet, D. K., Peitl, P., Jr., John, C. D., da Silva Junior, W. A., Tajara, E., Buckingham, J. C. & Solito, E. (2006). In vitro and in vivo studies on CCR10 regulation by Annexin A1. *FEBS Lett*, 580, 1431-1438.

<https://doi.org/10.1016/j.febslet.2006.01.072>

Rong, X., Liang, Y., Han, Q., Zhao, Y., Jiang, G., Zhang, X., Lin, X., Liu, Y., Zhang, Y., Han, X., Zhang, M., Luo, Y., Li, P., Wei, L., Yan, T. & Wang, E. (2019). Molecular Mechanisms of Tyrosine Kinase Inhibitor Resistance Induced by Membranous/Cytoplasmic/Nuclear Translocation of Epidermal Growth Factor Receptor. *J Thorac Oncol*, 14, 1766-1783.

<https://doi.org/10.1016/j.jtho.2019.06.014>

Sakaguchi, M., Murata, H., Sonegawa, H., Sakaguchi, Y., Futami, J., Kitazoe, M., Yamada, H. & Huh, N. H. (2007). Truncation of annexin A1 is a regulatory lever for linking epidermal growth factor signaling with cytosolic phospholipase A2 in normal and malignant squamous epithelial cells. *J Biol Chem*, 282, 35679-35686.

<https://doi.org/10.1074/jbc.M707538200>

Sarkar, S., Brautigan, D. L. & Lerner, J. M. (2017). Aurora Kinase A Promotes AR Degradation via the E3 Ligase CHIP. *Mol Cancer Res*, 15, 1063-1072.

<https://doi.org/10.1158/1541-7786.MCR-17-0062>

Schittenhelm, J., Trautmann, K., Tabatabai, G., Hermann, C., Meyermann, R. & Beschoner, R. (2009). Comparative analysis of annexin-1 in neuroepithelial tumors shows altered expression with the grade of malignancy but is not associated with survival. *Mod Pathol*, 22, 1600-1611.

<https://doi.org/10.1038/modpathol.2009.132>

Shi, Y., Tao, Y., Jiang, Y., Xu, Y., Yan, B., Chen, X., Xiao, L. & Cao, Y. (2012). Nuclear epidermal growth factor receptor interacts with transcriptional intermediary factor 2 to activate cyclin D1 gene expression triggered by the oncoprotein latent membrane protein 1. *Carcinogenesis*, 33, 1468-1478.

<https://doi.org/10.1093/carcin/bgs171>

Sigismund, S., Avanzato, D. & Lanzetti, L. (2018). Emerging functions of the EGFR in cancer. *Mol Oncol*, 12, 3-20.

<https://doi.org/10.1002/1878-0261.12155>

Snapkov, I., Oqvist, C. O., Figenschau, Y., Kogner, P., Johnsen, J. I. & Sveinbjornsson, B. (2016). The role of formyl peptide receptor 1 (FPR1) in neuroblastoma tumorigenesis. *BMC Cancer*, 16, 490.

<https://doi.org/10.1186/s12885-016-2545-1>

Sobek, K. M., Cummings, J. L., Bacich, D. J. & O'Keefe, D. S. (2017). Contrasting roles of the ABCG2 Q141K variant in prostate cancer. *Exp Cell Res*, 354, 40-47.

<https://doi.org/10.1016/j.yexcr.2017.03.020>

Sorkin, A., Helin, K., Waters, C. M., Carpenter, G. & Beguinot, L. (1992). Multiple autophosphorylation sites of the epidermal growth factor receptor are essential for receptor kinase activity and internalization. Contrasting significance of tyrosine 992 in the native and truncated receptors. *J Biol Chem*, 267, 8672-8678.

[https://doi.org/10.1016/S0021-9258\(18\)42495-7](https://doi.org/10.1016/S0021-9258(18)42495-7)

Sorkina, T., Huang, F., Beguinot, L. & Sorkin, A. (2002). Effect of tyrosine kinase inhibitors on clathrin-coated pit recruitment and internalization of epidermal growth factor receptor. *J Biol Chem*, 277, 27433-27441.

<https://doi.org/10.1074/jbc.M201595200>

Sridhar, S. S., Freedland, S. J., Gleave, M. E., Higano, C., Mulders, P., Parker, C., Sartor, O. & Saad, F. (2014). Castration-resistant prostate cancer: from new pathophysiology to new treatment. *Eur Urol*, 65, 289-299.

<https://doi.org/10.1016/j.eururo.2013.08.008>

Stenfeldt, A. L., Karlsson, J., Wenneras, C., Bylund, J., Fu, H. & Dahlgren, C. (2007). Cyclosporin H, Boc-MLF and Boc-FLFLF are antagonists that preferentially inhibit activity triggered through the formyl peptide receptor. *Inflammation*, 30, 224-229.

<https://doi.org/10.1007/s10753-007-9040-4>

Sukowati, C. H., Rosso, N., Pascut, D., Anfuso, B., Torre, G., Francalanci, P., Croce, L. S. & Tiribelli, C. (2012). Gene and functional up-regulation of the BCRP/ABCG2 transporter in hepatocellular carcinoma. *BMC Gastroenterol*, 12, 160.

<https://doi.org/10.1186/1471-230X-12-160>

Tomasello, C., Baldessari, C., Napolitano, M., Orsi, G., Grizzi, G., Bertolini, F., Barbieri, F. & Cascinu, S. (2018). Resistance to EGFR inhibitors in non-small cell lung cancer: Clinical management and future perspectives. *Crit Rev Oncol Hematol*, 123, 149-161.

<https://doi.org/10.1016/j.critrevonc.2018.01.013>

Tran, K. & McCormack, S. (2019). Androgen Receptor Targeted Agents for Castration Resistant Prostate Cancer: A Review of Clinical Effectiveness and Cost-Effectiveness. Ottawa (ON).

Tsaur, I., Heidegger, I., Kretschmer, A., Borgmann, H., Gandaglia, G., Briganti, A., de Visschere, P., Mathieu, R., Valerio, M., van den Bergh, R., Ost, P., Mirvald, C., Tilki, D., Ploussard, G., Surcel, C. & Party, E.-Y. P. C. W. (2019). Aggressive variants of prostate cancer - Are we ready to apply specific treatment right now? *Cancer Treat Rev*, 75, 20-26.

<https://doi.org/10.1016/j.ctrv.2019.03.001>

Vecchi, L., Alves Pereira Zoia, M., Goss Santos, T., de Oliveira Beserra, A., Colaco Ramos, C. M., Franca Matias Colombo, B., Paiva Maia, Y. C., Piana de Andrade, V., Teixeira Soares Mota, S., Goncalves de Araujo, T., Van Petten de Vasconcelos Azevedo, F., Soares, F. A., Oliani, S. M. & Goulart, L. R. (2018). Inhibition of the AnxA1/FPR1 autocrine axis reduces MDA-MB-231 breast cancer cell growth and aggressiveness in vitro and in vivo. *Biochim Biophys Acta Mol Cell Res*, 1865, 1368-1382.

<https://doi.org/10.1016/j.bbamcr.2018.06.010>

Vong, L., D'Acquisto, F., Pederzoli-Ribeil, M., Lavagno, L., Flower, R. J., Witko-Sarsat, V. & Perretti, M. (2007). Annexin 1 cleavage in activated neutrophils: a pivotal role for proteinase 3. *J Biol Chem*, 282, 29998-30004.

<https://doi.org/10.1074/jbc.M702876200>

- Wallace, T. J., Torre, T., Grob, M., Yu, J., Avital, I., Brucher, B., Stojadinovic, A. & Man, Y. G. (2014). Current approaches, challenges and future directions for monitoring treatment response in prostate cancer. *J Cancer*, 5, 3-24.
<https://doi.org/10.7150/jca.7709>
- Walther, A., Riehemann, K. & Gerke, V. (2000). A novel ligand of the formyl peptide receptor: annexin I regulates neutrophil extravasation by interacting with the FPR. *Mol Cell*, 5, 831-840.
[https://doi.org/10.1016/S1097-2765\(00\)80323-8](https://doi.org/10.1016/S1097-2765(00)80323-8)
- Wang, S. C., Nakajima, Y., Yu, Y. L., Xia, W., Chen, C. T., Yang, C. C., McIntush, E. W., Li, L. Y., Hawke, D. H., Kobayashi, R. & Hung, M. C. (2006). Tyrosine phosphorylation controls PCNA function through protein stability. *Nat Cell Biol*, 8, 1359-1368.
<https://doi.org/10.1038/ncb1501>
- Wang, Y. N. & Hung, M. C. (2012). Nuclear functions and subcellular trafficking mechanisms of the epidermal growth factor receptor family. *Cell Biosci*, 2, 13.
<https://doi.org/10.1186/2045-3701-2-13>
- Wang, Y. N., Wang, H., Yamaguchi, H., Lee, H. J., Lee, H. H. & Hung, M. C. (2010a). COPI-mediated retrograde trafficking from the Golgi to the ER regulates EGFR nuclear transport. *Biochem Biophys Res Commun*, 399, 498-504.
<https://doi.org/10.1016/j.bbrc.2010.07.096>
- Wang, Y. N., Yamaguchi, H., Hsu, J. M. & Hung, M. C. (2010b). Nuclear trafficking of the epidermal growth factor receptor family membrane proteins. *Oncogene*, 29, 3997-4006.
<https://doi.org/10.1038/onc.2010.157>
- Wee, P. & Wang, Z. (2017). Epidermal Growth Factor Receptor Cell Proliferation Signaling Pathways. *Cancers (Basel)*, 9.
<https://doi.org/10.3390/cancers9050052>
- White, I. J., Bailey, L. M., Aghakhani, M. R., Moss, S. E. & Futter, C. E. (2006). EGF stimulates annexin 1-dependent inward vesiculation in a multivesicular endosome subpopulation. *EMBO J*, 25, 1-12.
<https://doi.org/10.1038/sj.emboj.7600759>
- Williams, G. H. & Stoeber, K. (2012). The cell cycle and cancer. *J Pathol*, 226, 352-364.
<https://doi.org/10.1002/path.3022>
- Yang, Y., Liu, Y., Yao, X., Ping, Y., Jiang, T., Liu, Q., Xu, S., Huang, J., Mou, H., Gong, W., Chen, K., Bian, X. & Wang, J. M. (2011). Annexin 1 released by necrotic human glioblastoma cells stimulates tumor cell growth through the formyl peptide receptor 1. *Am J Pathol*, 179, 1504-1512.
<https://doi.org/10.1016/j.ajpath.2011.05.059>
- Yi, M. & Schnitzer, J. E. (2009). Impaired tumor growth, metastasis, angiogenesis and wound healing in annexin A1-null mice. *Proc Natl Acad Sci U S A*, 106, 17886-17891.
<https://doi.org/10.1073/pnas.0901324106>

Zhang, S., Li, J., Zhou, G., Mu, D., Yan, J., Xing, J., Yao, Z., Sheng, H., Li, D., Lv, C., Sun, B., Hong, Q. & Guo, H. (2017). Aurora-A regulates autophagy through the Akt pathway in human prostate cancer. *Cancer Biomark*, 19, 27-34.

<https://doi.org/10.3233/CBM-160238>

Zhang, W., Meng, Y., Liu, N., Wen, X. F. & Yang, T. (2015). Insights into Chemoresistance of Prostate Cancer. *Int J Biol Sci*, 11, 1160-1170.

<https://doi.org/10.7150/ijbs.11439>

Capítulo 3

Interleukin-6 signaling in triple negative breast cancer cells elicits the Annexin A1/Formyl peptide receptor 1 axis and affects the tumor microenvironment

Interleukin-6 signaling in triple negative breast cancer cells elicits the Annexin A1/Formyl peptide receptor 1 axis and affects the tumor microenvironment

Lara Vecchi^{1*}, Sara Teixeira Soares Mota^{1,2#}, Mariana Alves Pereira Zóia¹, Tiago Goss dos Santos³, Adriano de Oliveira Beserra³, Victor Piana de Andrade⁴, Luiz Ricardo Goulart^{1*}, Thaise Gonçalves Araújo^{1,2*}

¹Laboratory of Nanobiotechnology, Institute of Genetics and Biochemistry, Federal University of Uberlândia, Uberlândia, MG, Brazil.

²Laboratory of Genetics and Biotechnology, Federal University of Uberlândia, Patos de Minas, MG, Brazil

³International Research Center, AC Camargo Cancer Center, São Paulo, SP, Brazil.

⁴Department of Investigative Pathology, AC Camargo Cancer Center, São Paulo, SP, Brazil.

These authors contributed equally to this work

*Corresponding authors: Lara Vecchi, Luiz R. Goulart and Thaise Gonçalves de Araujo, Laboratory of Nanobiotechnology, Institute of Genetics and Biochemistry, Federal University of Uberlândia, Campus Umuarama, Bloco 2E, Sala 248, 38400-902, Uberlândia, MG, Brazil. Phone: +55 34 3225 8440. laravecchi7@yahoo.it; lrgoulart@ufu.br/lrgoulart@ucdavis.edu; tgaraujo@ufu.br.

Abstract: Annexin A1 (AnxA1) is a pleiotropic protein that exerts essential roles in breast cancer (BC) growth and aggressiveness. In our previous work, we described the autocrine signaling of AnxA1 through Formyl Peptide Receptor 1 (FPR1) in the triple-negative (TN) BC cell line, MDA-MB-231. Here we aimed to describe the interaction between the AnxA1/FPR1 and the Interleukin-6 (IL-6) signaling pathways and their role in the tumor microenvironment (TME). First, we demonstrated that AnxA1 and IL-6 expression levels are correlated in BC tissue samples. In three TNBC cell lines, overexpression of both AnxA1 and IL-6 was also identified. Next, we inhibited FPR1, the IL-6 receptor and STAT3 in both MDA-MB-231 and MDA-MB-157 cells. The FPR1 inhibition led to increased levels of IL-6 and secreted AnxA1 in both cell lines. On the other side, inhibition of the IL-6 receptor or STAT3 led to the impairment of AnxA1 secretion, suggesting the essential role of the IL-6 signaling cascade in the activation of the AnxA1/FPR1 autocrine axis. Finally, we described the interaction between IL-6 and the AnxA1/FPR1 pathways and their role on the TME by analyzing the effect of supernatants derived from MDA-MB-231 and MDA-MB-157 cells under inhibition of FPR1 or IL-6 signaling on fibroblast cell motility.

Keywords: Annexin A1, autocrine signaling, Breast cancer, Formyl Peptide Receptor, IL-6, STAT3;

Citation: Lastname, F.; Lastname, F.; Lastname, F. Title. *Cells* **2021**, *10*, x. <https://doi.org/10.3390/xxxxx>

Academic Editor: Firstname Last-name

Received: date

Accepted: date

Published: date

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Breast cancer (BC) is the most common cancer worldwide being a major cause of morbidity and mortality in women [1, 2]. The expression of Estrogen Receptor (ER) and Progesterone Receptor (PR) - that are collectively named hormone receptors (HR) -, Human Epidermal Growth Factor Receptor 2 (HER2) and of the proliferation marker, Ki67, defines BC subtypes which are categorized into: Lumina A (HR+, HER2-), Luminal B (HR+, HER2+ or HR+, HER2-; Ki67 high), HER2-enriched (HR-, HER2+), and Triple-Negative (TN: HR-, HER2-). The majority of TNBCs belongs to the basal-like subtype and displays a highly aggressive phenotype with poorer overall survival and high relapse

References

1. Harborg S, Zachariae R, Olsen J, Johannsen M, Cronin-Fenton D, Boggild H, et al. Overweight and prognosis in triple-negative breast cancer patients: a systematic review and meta-analysis. *NPJ Breast Cancer*. 2021;7(1):119. Epub 2021/09/12. doi: 10.1038/s41523-021-00325-6. PubMed PMID: 34508075. <https://doi.org/10.1038/s41523-021-00325-6>.
2. Huang J, Chan PS, Lok V, Chen X, Ding H, Jin Y, et al. Global incidence and mortality of breast cancer: a trend analysis. *Aging (Albany NY)*. 2021;13(4):5748-803. Epub 2021/02/17. doi: 10.18632/aging.202502. PubMed PMID: 33592581; PubMed Central PMCID: PMCPCMC7950292. <https://doi.org/10.18632/aging.202502>.
3. Burguin A, Diorio C, Durocher F. Breast Cancer Treatments: Updates and New Challenges. *J Pers Med*. 2021;11(8). Epub 2021/08/28. doi: 10.3390/jpm11080808. PubMed PMID: 34442452; PubMed Central PMCID: PMCPCMC8399130. <https://doi.org/10.3390/jpm11080808>.
4. Rossing M, Pedersen CB, Tvedskov T, Vejborg I, Talman ML, Olsen LR, et al. Clinical implications of intrinsic molecular subtypes of breast cancer for sentinel node status. *Scientific reports*. 2021;11(1):2259. Epub 2021/01/28. doi: 10.1038/s41598-021-81538-4. PubMed PMID: 33500440; PubMed Central PMCID: PMCPCMC7838175. <https://doi.org/10.1038/s41598-021-81538-4>.
5. Kanwal B. Untangling Triple-Negative Breast Cancer Molecular Peculiarity and Chemo-Resistance: Trailing Towards Marker-Based Targeted Therapies. *Cureus*. 2021;13(7):e16636. Epub 2021/08/31. doi: 10.7759/cureus.16636. PubMed PMID: 34458041; PubMed Central PMCID: PMCPCMC8384383. <https://doi.org/10.7759/cureus.16636>.
6. Berger ER, Park T, Saridakis A, Golshan M, Greenup RA, Ahuja N. Immunotherapy Treatment for Triple Negative Breast Cancer. *Pharmaceuticals (Basel)*. 2021;14(8). Epub 2021/08/29. doi: 10.3390/ph14080763. PubMed PMID: 34451860; PubMed Central PMCID: PMCPCMC8401402. <https://doi.org/10.3390/ph14080763>.
7. Hussain SP, Harris CC. Inflammation and cancer: an ancient link with novel potentials. *International journal of cancer Journal international du cancer*. 2007;121(11):2373-80. doi: 10.1002/ijc.23173. PubMed PMID: 17893866. <https://doi.org/10.1002/ijc.23173>
8. Schetter AJ, Heegaard NH, Harris CC. Inflammation and cancer: interweaving microRNA, free radical, cytokine and p53 pathways. *Carcinogenesis*. 2010;31(1):37-49. doi: 10.1093/carcin/bgp272. PubMed PMID: 19955394; PubMed Central PMCID: PMC2802675. <https://doi.org/10.1093/carcin/bgp272>.
9. Gerke V, Moss SE. Annexins: From structure to function. *Physiol Rev*. 2002;82(2):331-71. doi: DOI 10.1152/physrev.00030.2001. PubMed PMID: WOS:000174854700002. <https://doi.org/10.1093/carcin/bgp272>.
10. Petrella A, Festa M, Ercolino SF, Zerilli M, Stassi G, Solito E, et al. Induction of annexin-1 during TRAIL-induced apoptosis in thyroid carcinoma cells. *Cell death and differentiation*. 2005;12(10):1358-60. doi: 10.1038/sj.cdd.4401645. PubMed PMID: 15846370. <https://doi.org/10.1038/sj.cdd.4401645>.
11. Scannell M, Maderna P. Lipoxins and annexin-1: resolution of inflammation and regulation of phagocytosis of apoptotic cells. *TheScientificWorldJournal*. 2006;6:1555-73. doi: 10.1100/tsw.2006.259. PubMed PMID: 17160341. <https://doi.org/10.1100/tsw.2006.259>.
12. Rescher U, Gerke V. Annexins--unique membrane binding proteins with diverse functions. *Journal of cell science*. 2004;117(Pt 13):2631-9. doi: 10.1242/jcs.01245. PubMed PMID: 15169834. <https://doi.org/10.1242/jcs.01245>.
13. Sakaguchi M, Murata H, Sonogawa H, Sakaguchi Y, Futami J, Kitazoe M, et al. Truncation of annexin A1 is a regulatory lever for linking epidermal growth factor signaling with cytosolic phospholipase A2 in normal and malignant squamous epithelial cells. *The Journal of biological chemistry*. 2007;282(49):35679-86. Epub 2007/10/13. doi: 10.1074/jbc.M707538200. PubMed PMID: 17932043. <https://doi.org/10.1074/jbc.M707538200>.
14. Parente L, Solito E. Annexin 1: more than an anti-phospholipase protein. *Inflammation research : official journal of the European Histamine Research Society* [et al]. 2004;53(4):125-32. Epub 2004/04/03. doi: 10.1007/s00011-003-1235-z. PubMed PMID: 15060718. <https://doi.org/10.1007/s00011-003-1235-z>.
15. Gavins FN, Yona S, Kamal AM, Flower RJ, Perretti M. Leukocyte antiadhesive actions of annexin 1: ALXR- and FPR-related anti-inflammatory mechanisms. *Blood*. 2003;101(10):4140-7. doi: 10.1182/blood-2002-11-3411. PubMed PMID: 12560218. <https://doi.org/10.1182/blood-2002-11-3411>.
16. Araújo TG, Mota STS, Ferreira HSV, Ribeiro MA, Goulart LR, Vecchi L. Annexin A1 as a Regulator of Immune Response in Cancer. 2021;10(9):2245. PubMed PMID: doi:10.3390/cells10092245. <https://doi.org/10.3390/cells10092245>.
17. Sheikh MH, Solito E. Annexin A1: Uncovering the Many Talents of an Old Protein. *Int J Mol Sci*. 2018;19(4). Epub 2018/04/05. doi: 10.3390/ijms19041045. PubMed PMID: 29614751; PubMed Central PMCID: PMCPCMC5979524. <https://doi.org/10.3390/ijms19041045>.
18. Alvarez-Teijeiro S, Menendez ST, Villarronga MA, Pena-Alonso E, Rodrigo JP, Morgan RO, et al. Annexin A1 down-regulation in head and neck squamous cell carcinoma is mediated via transcriptional control with direct involvement of miR-196a/b. *Scientific reports*. 2017;7(1):6790. Epub 2017/07/30. doi: 10.1038/s41598-017-07169-w. PubMed PMID: 28754915; PubMed Central PMCID: PMCPCMC5533727. <https://doi.org/10.1038/s41598-017-07169-w>.
19. Mota STS, Vecchi L, Alves DA, Cordeiro AO, Guimaraes GS, Campos-Fernandez E, et al. Annexin A1 promotes the nuclear localization of the epidermal growth factor receptor in castration-resistant prostate cancer. *Int J Biochem Cell Biol*. 99

- 2020;127:105838. Epub 2020/08/29. doi: 10.1016/j.biocel.2020.105838. PubMed PMID: 32858191. <https://doi.org/10.1016/j.biocel.2020.105838>.
20. Liu YF, Zhang PF, Li MY, Li QQ, Chen ZC. Identification of annexin A1 as a proinvasive and prognostic factor for lung adenocarcinoma. *Clinical & experimental metastasis*. 2011;28(5):413-25. Epub 2011/03/03. doi: 10.1007/s10585-011-9380-1. PubMed PMID: 21365324. <https://doi.org/10.1007/s10585-011-9380-1>.
21. Lin CY, Jeng YM, Chou HY, Hsu HC, Yuan RH, Chiang CP, et al. Nuclear localization of annexin A1 is a prognostic factor in oral squamous cell carcinoma. *Journal of surgical oncology*. 2008;97(6):544-50. Epub 2008/02/26. doi: 10.1002/jso.20992. PubMed PMID: 18297688. <https://doi.org/10.1002/jso.20992>.
22. Sobral-Leite M, Wesseling J, Smit VT, Nevanlinna H, van Miltenburg MH, Sanders J, et al. Annexin A1 expression in a pooled breast cancer series: association with tumor subtypes and prognosis. *BMC Med*. 2015;13:156. Epub 2015/07/04. doi: 10.1186/s12916-015-0392-6. PubMed PMID: 26137966; PubMed Central PMCID: PMC4489114. <https://doi.org/10.1186/s12916-015-0392-6>.
23. Vecchi L, Alves Pereira Zoia M, Goss Santos T, de Oliveira Beserra A, Colaco Ramos CM, Franca Matias Colombo B, et al. Inhibition of the AnxA1/FPR1 autocrine axis reduces MDA-MB-231 breast cancer cell growth and aggressiveness in vitro and in vivo. *Biochim Biophys Acta Mol Cell Res*. 2018;1865(9):1368-82. Epub 2018/06/23. doi: 10.1016/j.bbamcr.2018.06.010. PubMed PMID: 29932988.
24. de Graauw M, van Miltenburg MH, Schmidt MK, Pont C, Lalai R, Kartopawiro J, et al. Annexin A1 regulates TGF-beta signaling and promotes metastasis formation of basal-like breast cancer cells. *Proceedings of the National Academy of Sciences of the United States of America*. 2010;107(14):6340-5. Epub 2010/03/24. doi: 10.1073/pnas.0913360107. PubMed PMID: 20308542; PubMed Central PMCID: PMC2852023. <https://doi.org/10.1073/pnas.0913360107>.
25. Williams SL, Milne IR, Bagley CJ, Gamble JR, Vadas MA, Pitson SM, et al. A proinflammatory role for proteolytically cleaved annexin A1 in neutrophil transendothelial migration. *J Immunol*. 2010;185(5):3057-63. Epub 2010/08/04. doi: 10.4049/jimmunol.1000119. PubMed PMID: 20679535. <https://doi.org/10.4049/jimmunol.1000119>.
26. Solito E, de Coupade C, Parente L, Flower RJ, Russo-Marie F. IL-6 stimulates annexin 1 expression and translocation and suggests a new biological role as class II acute phase protein. *Cytokine*. 1998;10(7):514-21. Epub 1998/08/14. doi: 10.1006/cyto.1997.0325. PubMed PMID: 9702415. <https://doi.org/10.1006/cyto.1997.0325>.
27. Ni C, Gao S, Zheng Y, Liu P, Zhai Y, Huang W, et al. Annexin A1 Attenuates Neutrophil Migration and IL-6 Expression through Fpr2 in a Mouse Model of Streptococcus suis-Induced Meningitis. *Infect Immun*. 2021;89(3). Epub 2020/12/16. doi: 10.1128/IAI.00680-20. PubMed PMID: 33318141; PubMed Central PMCID: PMC78097268. <https://doi.org/10.1128/IAI.00680-20>.
28. Tanaka T, Narazaki M, Kishimoto T. IL-6 in inflammation, immunity, and disease. *Cold Spring Harb Perspect Biol*. 2014;6(10):a016295. Epub 2014/09/06. doi: 10.1101/cshperspect.a016295. PubMed PMID: 25190079; PubMed Central PMCID: PMC4176007. <https://doi.org/10.1101/cshperspect.a016295>.
29. Lee SO, Yang X, Duan S, Tsai Y, Strojny LR, Keng P, et al. IL-6 promotes growth and epithelial-mesenchymal transition of CD133+ cells of non-small cell lung cancer. *Oncotarget*. 2016;7(6):6626-38. Epub 2015/12/18. doi: 10.18632/oncotarget.6570. PubMed PMID: 26675547; PubMed Central PMCID: PMC4872738. <https://doi.org/10.18632/oncotarget.6570>.
30. Scheller J, Ohnesorge N, Rose-John S. Interleukin-6 trans-signalling in chronic inflammation and cancer. *Scand J Immunol*. 2006;63(5):321-9. Epub 2006/04/28. doi: 10.1111/j.1365-3083.2006.01750.x. PubMed PMID: 16640655. <https://doi.org/10.1111/j.1365-3083.2006.01750.x>.
31. Ma JH, Qin L, Li X. Role of STAT3 signaling pathway in breast cancer. *Cell Commun Signal*. 2020;18(1):33. Epub 2020/03/01. doi: 10.1186/s12964-020-0527-z. PubMed PMID: 32111215; PubMed Central PMCID: PMC7048131. <https://doi.org/10.1186/s12964-020-0527-z>.
32. Adachi Y, Yoshio-Hoshino N, Nishimoto N. The blockade of IL-6 signaling in rational drug design. *Curr Pharm Des*. 2008;14(12):1217-24. Epub 2008/05/14. doi: 10.2174/138161208784246072. PubMed PMID: 18473869. <https://doi.org/10.2174/138161208784246072>.
33. Culig Z, Puhr M. Interleukin-6: a multifunctional targetable cytokine in human prostate cancer. *Mol Cell Endocrinol*. 2012;360(1-2):52-8. Epub 2011/06/15. doi: 10.1016/j.mce.2011.05.033. PubMed PMID: 21664423; PubMed Central PMCID: PMC3409376. <https://doi.org/10.1016/j.mce.2011.05.033>.
34. Masjedi A, Hashemi V, Hojjat-Farsangi M, Ghalamfarsa G, Azizi G, Yousefi M, et al. The significant role of interleukin-6 and its signaling pathway in the immunopathogenesis and treatment of breast cancer. *Biomed Pharmacother*. 2018;108:1415-24. Epub 2018/10/31. doi: 10.1016/j.biopha.2018.09.177. PubMed PMID: 30372844. <https://doi.org/10.1016/j.biopha.2018.09.177>.
35. Garcia-Tunon I, Ricote M, Ruiz A, Fraile B, Paniagua R, Royuela M. IL-6, its receptors and its relationship with bcl-2 and bax proteins in infiltrating and in situ human breast carcinoma. *Histopathology*. 2005;47(1):82-9. Epub 2005/06/29. doi: 10.1111/j.1365-2559.2005.02178.x. PubMed PMID: 15982327. <https://doi.org/10.1111/j.1365-2559.2005.02178.x>.
36. Sanguinetti A, Santini D, Bonafe M, Taffurelli M, Avenia N. Interleukin-6 and pro inflammatory status in the breast tumor microenvironment. *World J Surg Oncol*. 2015;13:129. Epub 2015/04/17. doi: 10.1186/s12957-015-0529-2. PubMed PMID: 25881039; PubMed Central PMCID: PMC4397867. <https://doi.org/10.1186/s12957-015-0529-2>.

37. Tawara K, Oxford JT, Jorczyk CL. Clinical significance of interleukin (IL)-6 in cancer metastasis to bone: potential of anti-IL-6 therapies. *Cancer management and research*. 2011;3:177-89. Epub 2011/06/01. doi: 10.2147/CMR.S18101. PubMed PMID: 21625400; PubMed Central PMCID: PMC3101113. <https://doi.org/10.2147/CMR.S18101>.
38. Pu YS, Hour TC, Chuang SE, Cheng AL, Lai MK, Kuo ML. Interleukin-6 is responsible for drug resistance and anti-apoptotic effects in prostatic cancer cells. *Prostate*. 2004;60(2):120-9. Epub 2004/05/27. doi: 10.1002/pros.20057. PubMed PMID: 15162378. <https://doi.org/10.1002/pros.20057>.
39. Wang Y, Niu XL, Qu Y, Wu J, Zhu YQ, Sun WJ, et al. Autocrine production of interleukin-6 confers cisplatin and paclitaxel resistance in ovarian cancer cells. *Cancer Lett*. 2010;295(1):110-23. Epub 2010/03/20. doi: 10.1016/j.canlet.2010.02.019. PubMed PMID: 20236757. <https://doi.org/10.1016/j.canlet.2010.02.019>.
40. Wu P, Wu D, Zhao L, Shen G, Huang J, et al. Prognostic role of STAT3 in solid tumors: a systematic review and meta-analysis. *Oncotarget*. 2016;7(15):19863-83. Epub 2016/03/10. doi: 10.18632/oncotarget.7887. PubMed PMID: 26959884; PubMed Central PMCID: PMC4991424. <https://doi.org/10.18632/oncotarget.7887>.
41. Spitzner M, Ebner R, Wolff HA, Ghadimi BM, Wienands J, Grade M. STAT3: A Novel Molecular Mediator of Resistance to Chemoradiotherapy. *Cancers (Basel)*. 2014;6(4):1986-2011. Epub 2014/10/01. doi: 10.3390/cancers6041986. PubMed PMID: 25268165; PubMed Central PMCID: PMC4276953. <https://doi.org/10.18632/oncotarget.7887>.
42. Banerjee K, Resat H. Constitutive activation of STAT3 in breast cancer cells: A review. *Int J Cancer*. 2016;138(11):2570-8. Epub 2015/11/13. doi: 10.1002/ijc.29923. PubMed PMID: 26559373; PubMed Central PMCID: PMC4801660. <https://doi.org/10.1002/ijc.29923>.
43. Yeo SK, Guan JL. Breast Cancer: Multiple Subtypes within a Tumor? *Trends Cancer*. 2017;3(11):753-60. Epub 2017/11/10. doi: 10.1016/j.trecan.2017.09.001. PubMed PMID: 29120751; PubMed Central PMCID: PMC5802368. <https://doi.org/10.1016/j.trecan.2017.09.001>.
44. Al-Thoubaity FK. Molecular classification of breast cancer: A retrospective cohort study. *Ann Med Surg (Lond)*. 2020;49:44-8. Epub 2020/01/01. doi: 10.1016/j.amsu.2019.11.021. PubMed PMID: 31890196; PubMed Central PMCID: PMC6926136. <https://doi.org/10.1016/j.amsu.2019.11.021>.
45. Al-Mahmood S, Sapiezynski J, Garbuzenko OB, Minko T. Metastatic and triple-negative breast cancer: challenges and treatment options. *Drug Deliv Transl Res*. 2018;8(5):1483-507. Epub 2018/07/07. doi: 10.1007/s13346-018-0551-3. PubMed PMID: 29978332; PubMed Central PMCID: PMC6133085. <https://doi.org/10.1007/s13346-018-0551-3>.
46. Yin L, Duan JJ, Bian XW, Yu SC. Triple-negative breast cancer molecular subtyping and treatment progress. *Breast Cancer Res*. 2020;22(1):61. Epub 2020/06/11. doi: 10.1186/s13058-020-01296-5. PubMed PMID: 32517735; PubMed Central PMCID: PMC7285581. <https://doi.org/10.1186/s13058-020-01296-5>.
47. Okano M, Oshi M, Butash AL, Katsuta E, Tachibana K, Saito K, et al. Triple-Negative Breast Cancer with High Levels of Annexin A1 Expression Is Associated with Mast Cell Infiltration, Inflammation, and Angiogenesis. *Int J Mol Sci*. 2019;20(17). Epub 2019/08/30. doi: 10.3390/ijms20174197. PubMed PMID: 31461932; PubMed Central PMCID: PMC6747082. <https://doi.org/10.3390/ijms20174197>.
48. Yang YH, Aeberli D, Dacumos A, Xue JR, Morand EF. Annexin-1 regulates macrophage IL-6 and TNF via glucocorticoid-induced leucine zipper. *J Immunol*. 2009;183(2):1435-45. Epub 2009/06/26. doi: 10.4049/jimmunol.0804000. PubMed PMID: 19553536. <https://doi.org/10.4049/jimmunol.0804000>.
49. Moraes LA, Ampomah PB, Lim LHK. Annexin A1 in inflammation and breast cancer: a new axis in the tumor microenvironment. *Cell adhesion & migration*. 2018;12(5):417-23. Epub 2018/08/21. doi: 10.1080/19336918.2018.1486143. PubMed PMID: 30122097; PubMed Central PMCID: PMC6363057. <https://doi.org/10.1080/19336918.2018.1486143>.
50. Mauer J, Denson JL, Bruning JC. Versatile functions for IL-6 in metabolism and cancer. *Trends Immunol*. 2015;36(2):92-101. Epub 2015/01/27. doi: 10.1016/j.it.2014.12.008. PubMed PMID: 25616716. <https://doi.org/10.1016/j.it.2014.12.008>.
51. Johnson DE, O'Keefe RA, Grandis JR. Targeting the IL-6/JAK/STAT3 signalling axis in cancer. *Nat Rev Clin Oncol*. 2018;15(4):234-48. Epub 2018/02/07. doi: 10.1038/nrclinonc.2018.8. PubMed PMID: 29405201; PubMed Central PMCID: PMC5858971. <https://doi.org/10.1016/j.it.2014.12.008>.
52. Weng YS, Tseng HY, Chen YA, Shen PC, Al Haq AT, Chen LM, et al. MCT-1/miR-34a/IL-6/IL-6R signaling axis promotes EMT progression, cancer stemness and M2 macrophage polarization in triple-negative breast cancer. *Mol Cancer*. 2019;18(1):42. Epub 2019/03/20. doi: 10.1186/s12943-019-0988-0. PubMed PMID: 30885232; PubMed Central PMCID: PMC6421700. <https://doi.org/10.1186/s12943-019-0988-0>.
53. Qin JJ, Yan L, Zhang J, Zhang WD. STAT3 as a potential therapeutic target in triple negative breast cancer: a systematic review. *Journal of experimental & clinical cancer research : CR*. 2019;38(1):195. Epub 2019/05/16. doi: 10.1186/s13046-019-1206-z. PubMed PMID: 31088482; PubMed Central PMCID: PMC6518732. <https://doi.org/10.1186/s13046-019-1206-z>.
54. Jin K, Pandey NB, Popel AS. Simultaneous blockade of IL-6 and CCL5 signaling for synergistic inhibition of triple-negative breast cancer growth and metastasis. *Breast Cancer Res*. 2018;20(1):54. Epub 2018/06/15. doi: 10.1186/s13058-018-0981-3. PubMed PMID: 29898755; PubMed Central PMCID: PMC6000947. <https://doi.org/10.1186/s13058-018-0981-3>.
55. Fasoulakis Z, Kolios G, Papamanolis V, Kontomanolis EN. Interleukins Associated with Breast Cancer. *Cureus*. 2018;10(11):e3549. Epub 2019/01/17. doi: 10.7759/cureus.3549. PubMed PMID: 30648081; PubMed Central PMCID: PMC6324869. <https://doi.org/10.7759/cureus.3549>.
56. Studebaker AW, Storci G, Werbeck JL, Sansone P, Sasser AK, Tavoroli S, et al. Fibroblasts isolated from common sites of breast cancer metastasis enhance cancer cell growth rates and invasiveness in an interleukin-6-dependent manner. *Cancer*

- Res. 2008;68(21):9087-95. Epub 2008/11/01. doi: 10.1158/0008-5472.CAN-08-0400. PubMed PMID: 18974155. <https://doi.org/10.1158/0008-5472.CAN-08-0400>.
57. Costa A, Kieffer Y, Scholer-Dahirel A, Pelon F, Bourachot B, Cardon M, et al. Fibroblast Heterogeneity and Immunosuppressive Environment in Human Breast Cancer. *Cancer Cell*. 2018;33(3):463-79 e10. Epub 2018/02/20. doi: 10.1016/j.ccell.2018.01.011. PubMed PMID: 29455927. <https://doi.org/10.1016/j.ccell.2018.01.011>.
58. Karakasheva TA, Lin EW, Tang Q, Qiao E, Waldron TJ, Soni M, et al. IL-6 Mediates Cross-Talk between Tumor Cells and Activated Fibroblasts in the Tumor Microenvironment. *Cancer Res*. 2018;78(17):4957-70. Epub 2018/07/07. doi: 10.1158/0008-5472.CAN-17-2268. PubMed PMID: 29976575; PubMed Central PMCID: PMC6125177. <https://doi.org/10.1158/0008-5472.CAN-17-2268>.
59. Toyoshima Y, Kitamura H, Xiang H, Ohno Y, Homma S, Kawamura H, et al. IL6 Modulates the Immune Status of the Tumor Microenvironment to Facilitate Metastatic Colonization of Colorectal Cancer Cells. *Cancer Immunol Res*. 2019;7(12):1944-57. Epub 2019/09/27. doi: 10.1158/2326-6066.CIR-18-0766. PubMed PMID: 31554639. <https://doi.org/10.1158/2326-6066.CIR-18-0766>.
60. Jones SA, Jenkins BJ. Recent insights into targeting the IL-6 cytokine family in inflammatory diseases and cancer. *Nat Rev Immunol*. 2018;18(12):773-89. Epub 2018/09/27. doi: 10.1038/s41577-018-0066-7. PubMed PMID: 30254251. <https://doi.org/10.1038/s41577-018-0066-7>.
61. Hirano T. IL-6 in inflammation, autoimmunity and cancer. *Int Immunol*. 2021;33(3):127-48. Epub 2020/12/19. doi: 10.1093/intimm/dxaa078. PubMed PMID: 33337480; PubMed Central PMCID: PMC7799025. <https://doi.org/10.1093/intimm/dxaa078>.
62. Alraouji NN, Aboussekhra A. Tocilizumab inhibits IL-8 and the proangiogenic potential of triple negative breast cancer cells. *Mol Carcinog*. 2021;60(1):51-9. Epub 2020/12/03. doi: 10.1002/mc.23270. PubMed PMID: 33264466. <https://doi.org/10.1002/mc.23270>.