



**UNIVERSIDADE FEDERAL DE UBERLÂNDIA**  
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**AVALIAÇÃO DA MICROTOMOGRAFIA COMPUTADORIZADA DO REPARO  
ÓSSEO ALVEOLAR: QUAL REGIÃO VOCÊ DEVE ANALISAR?**

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ÓSSEO ALVEOLAR: QUAL REGIÃO VOCÊ DEVE ANALISAR?**

Trabalho de conclusão de curso apresentado a Faculdade de Odontologia da Universidade Federal de Uberlândia, como requisito parcial para obtenção do título de Cirurgião-dentista.

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## ABSTRACT

**Objective:** Evaluate the influence of the extraction socket (distal or lingual root) and the type of region of interest (ROI) definition (manual or predefined) on the assessment of alveolar repair after tooth extraction using micro-computed tomography (micro-CT).

**Materials and Methods:** The sample consisted of micro-CT volumes of the mandibles submitted to extraction of the mandibular right first molar (M1) of seven Wistar rats. The reconstructed images were analyzed using the extraction sockets: D- distal and IL- intermediate lingual root; and the ROI: MA- manual, CR- central round and PR- peripheral round. The BV/TV values obtained were analyzed by ANOVA two-way with Tukey post hoc test ( $\alpha = 5\%$ ).

**Results:** In relation to the analyzed extraction socket, the D socket resulted in significantly lower BV/TV values than the IL socket, for the groups MA ( $P = 0.001$ ), CR ( $P < 0.001$ ), PR ( $P < 0.001$ ). For the region of interest (ROI), BV/TV was significantly higher ( $P < 0.001$ ) for the MA group ( $75.11 \pm 6.69$ ) compared to CR ( $65.31 \pm 5.16$ ) and PR groups, with lower BV/TV for CR ( $55.96 \pm 7.35$ ) in the D socket. However, no significant difference was observed for the groups MA ( $91.38 \pm 4.32$ ,  $P = 0.855$ ), CR ( $92.12 \pm 5.49$ ,  $P = 0.769$ ) and PR ( $93.08 \pm 4.16$ ,  $P = 0.453$ ) in the IL extraction socket.

**Conclusion:** Different ROIs definitions and the extraction socket being analyzed affect the morphometric results in micro-CT. The predefined method with standardized ROI in the central region of the bone defect in the distal extraction socket resulted in a more effective assessment of bone volume demonstrating the most critical region of the bone neoformation process.

**keyword:** micro-computed tomography, imaging, animal models, alveolar bone, tooth extraction

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## 1 INTRODUCTION

The alveolar bone is characterized by a continuous and rapid remodeling in response to stimuli <sup>1,2,3</sup>. Several clinical procedures, such as rehabilitative therapies, depend on the socket-healing process after tooth extractions and your understanding in preclinical studies is crucial. Therefore, the characterization of the dynamic process of bone to replace an extracted tooth and improvement of alveolar bone regenerative strategies is a topic of special interest in Dentistry <sup>4,5,6</sup>.

The Micro-Computed Tomography (Micro-CT) quickly become a standard tool in the assessment of bone microarchitecture in animal models, as the complimentary alternative the traditional histomorphometry <sup>7,8,9,10,11,12</sup>. The nondestructive method, short turnaround time, volumetric analyses are appealing in the analysis of the microarchitecture of native and newly formed bone by means of morphometric parameters (2D and 3D) and mineral density <sup>7,9,13,14</sup>. In addition, the same sample can later be prepared for histologic analysis <sup>7</sup>. As Micro-CT use has expanded, analyses have grown to encompass complex structures as in the dentoalveolar complex, and has been widely used to study alveolar bone remodeling <sup>12,15,16,17</sup>. However, some variables and selected parameters in Micro-CT may affect the morphological outcomes<sup>18</sup>.

One analysis parameter that influences on the quality of the results is the use the ROIs, which should be delimited based on experimental questions to be answered and it should be used with caution as it can bias results and provide imprecise values <sup>12,19,20</sup>. This process could be done manually <sup>15,21</sup> or in an automatic/semi-automatic fashion <sup>11,22,23</sup>. Different formats of ROIs (rectangular, polygonal, round, cubic, ring)

can be used as a representative sample in bone neoformation studies <sup>12,24</sup>. Another factor taken into account in alveolar repair analysis is the location of the ROI, i.e., the extraction socket used for analysis. Studies have evaluated the distal extraction socket <sup>21,24</sup>, the mesial extraction socket <sup>23,25</sup>, nonetheless, densities of the dentoalveolar tissues and the amount of neoformed bone vary by location and the size of the socket, which makes it difficult to direct compare with other models and studies <sup>12,24</sup>.

Despite current advances, there is no standardization of parameters for analyzing Micro-CT images in preclinical studies. Related to protocols for rodent jawbones, the literature is particularly scarce and the methodology for a reproducible quantitative analysis is not informative or detailed enough, what makes it difficult to understand how analyses were performed and if findings can be broadly compared to other models and studies <sup>12,24</sup>, which may compromise the scientific impact of the studies <sup>20</sup>. We began this study with the following question: Micro-CT evaluation of alveolar bone repair: what region should you analyse? Thus, the aim of this study was to evaluate the influence of the extraction socket (distal or lingual root) and the type of region of interest (ROI) definition (manual or predefined) on the assessment of alveolar repair after tooth extraction using micro-CT, and understand the effect of variation in these parameters and which method can result in more effective assessment of bone volume. The null hypothesis was that the type of ROI and the analysis region did not influence the outcome from Micro-CT analysis.

## 2 MATERIALS AND METHODS

This study was carried out in compliance which conform to ARRIVE guidelines for preclinical studies and the normative guidelines of the National Council for Animal Control and Experimentation (CONCEA). The sample consisted of micro-CT images of the mandible of 7 male Wistar rats that were acquired for previously approved research by the Research Ethics Committee of the institution (CEUA protocol 013/19).

Mandibular right first molar (M1) extractions were performed using luxators holleback Sculptor No.3 (Golgran, São Paulo, São Paulo, Brasil) and micro-forceps (n°5, Golgran, São Paulo, São Paulo, Brasil) to avoid fracture of the roots following the established protocols<sup>5</sup>. The animals were euthanized 14 days after tooth extraction by intraperitoneal overdose of thiopental (150 mg/kg). The hemimandibles were fixed in 4% paraformaldehyde solution and 10% 0.1 M phosphate buffer (pH 7.4) during 48h and scanned using X-ray microcomputed tomography Skyscan 1176 (Bruker, Kontich, Belgium) using the following settings: isotropic voxel size of 9  $\mu\text{m}$  (65 kV; 385  $\mu\text{A}$ ); aluminum filter of 1 mm; rotation step of 0.5°; 3 frames per rotation degree; and 180° rotation, scan time 55 minutes. Each mandible was fixed with wax on the scanning platform to ensure a standardized position with the long axis vertical to the horizontal plane wrapped in wet paper to maintain moisture during the scanning procedure. After scanning, the images were imported into NRecon Reconstruction software (version 1.6.6.0, Bruker, Kontich, Belgium) for reconstruction in grayscale, presenting x-ray attenuation coefficients with values related to bone structure. The images were reconstructed using the following settings: 40% for beam hardening correction, 0 for



smoothing and 12 to reduce ring artifacts. The reconstructions included the mandibular first and second molar region.

Data Viewer software (SkyScan) was used to adjust the images of the hemi-mandibles to standard positioning in order to get better positioning of the distal and lingual tooth socket of the extracted mandibular first molar. The images in the axial/transverse plane with standard orientation were exported to CTAn software (version 1.14.4.1, SkyScan, Bruker, Belgium) and only the sections including the distal and lingual extraction socket of the extracted mandibular first molar was selected to delineate the ROI showing alveolar socket healing, being a total of 80 slices along the cervical, middle and apical third of the socket for all samples. The reconstructed images were analyzed using the extraction sockets: D- distal and IL- intermediate lingual root; and the ROI: MA- manual, CR- central round and PR- peripheral round. The segmentation of region of interest in the D and IL extraction socket was performed individually for each ROI format. In the MA group, a personalized ROI was acquired manually using the computer mouse to delimit the extraction socket, respecting its contours. In the CR group, a predefined ROI of round shape of 0.5 mm in diameter was selected and centrally positioned in the extraction socket of the mandibular right first molar (M1). In the same manner, for the analysis of PR group, a standardized ROI of round format of 0.5 mm in diameter was positioned in the region close to the buccal surface in the extraction socket of the M1 (Fig.1). The entire region was defined by interpolation of the ROIs every 10 slices. After ROI delimitation, a global threshold ranging from 71-255 was established. The threshold was defined as the mean of the automatic threshold values (Otsu method) calculated from 10 samples from a previous study. A single operator carried out all the analysis. Then, three-dimensional analysis

of the region of interest was performed to examine the features of the bone microarchitecture, according to Bouxsein *et al.* 2010 <sup>7</sup>: percent bone volume (BV/TV,%).

## 2.1 Statistical analysis

Statistical analysis was performed using SigmaPlot® (SigmaPlot v13.1; Systat Software Inc.). The influence of extraction socket (distal or lingual root) and the type of region of interest (ROI) definition (manual or predefined) on BV/TV values was assessed using two-way analysis of variance (ANOVA) with Tukey's post hoc test, considering a significance level of  $\alpha=0.05$ .

## 3 RESULTS

The BV/TV means and standard deviations are described in Table 1. Two-way ANOVA showed that BV/TV values were significantly influenced by type of ROI ( $P < 0.001$ ), analysis region ( $P < 0.001$ ), and the interaction between these factors ( $P < 0.001$ ).

In relation to the analyzed extraction socket, the D socket resulted in significantly lower BV/TV values than the IL socket, for the groups MA ( $P = 0.001$ ), CR ( $P < 0.001$ ), PR ( $P < 0.001$ ). For the region of interest (ROI), BV/TV was significantly higher ( $P < 0.001$ ) for the MA group ( $75.11 \pm 6.69$ ) compared to CR ( $65.31 \pm 5.16$ ) and PR groups, with lower BV/TV for CR ( $55.96 \pm 7.35$ ) in the D socket. However, no significant difference was observed for the groups MA ( $91.38 \pm 4.32$ ,  $P = 0.855$ ), CR ( $92.12 \pm 5.49$ ,  $P = 0.769$ ) and PR ( $93.08 \pm 4.16$ ,  $P = 0.453$ ) in the IL extraction socket.

## 4 DISCUSSION

The parameters used in the analysis in Micro-CT may have a direct influence on the final characteristics of the image <sup>26</sup> and, consequently, in the morphometric outcome <sup>20</sup>. This study evaluated the influence of analysis parameters, including the region of interest (ROI) and the extraction socket of Micro-CT images on the evaluation of alveolar bone repair after tooth extraction the M1 in rats. The amount of neoformed bone (BV/TV) were significantly influenced by the ROI shape, the evaluated extraction socket, and the interaction between these factors. Thus, the null hypothesis that these variables would not influence the outcome from Micro-CT analysis was rejected.

Several preclinical models have been used to evaluate bone repair process in surgically created defects, such as femur/tibia <sup>27,28,29,30</sup>, and extraction socket in mandibles <sup>21,23, 25,31,32</sup>. The literature is conflicting regarding the portion of extraction sockets selected for the ROI. Alveolar repair has been analyzed in different extraction sockets (e.g., extraction socket of the distal <sup>21,24</sup> and mesial <sup>23, 25</sup>. Moreover, different ROIs formats are found in the literature, including those manually delineated <sup>15,21</sup> and predefined shapes <sup>11,22,33</sup>. However, the method used for ROI delimitation can affect the morphometric results in Micro-CT <sup>20,34</sup>. Besides, it has been demonstrated that by changing the position of the ROI, a variation of 12%–37% in the BV value was observed <sup>35</sup>. Thus, the direct comparison of results among the studies is not recommended.

In our study, a rounded standardized shape for determination of the ROI was evaluated and changes in its position resulted in significant differences for BV/TV values. Less bone neoformation for the central round ROI was detected in comparison to the peripheral ROI. In addition, higher BV/TV values were observed for manual ROI

for distal root M1. The bone healing occurs as a centripetal process, as a result of bone formation originating on the socket walls and subsequently confluent until filling all the socket extension <sup>33,36,37</sup>. This process explains the results found in the present study, demonstrating a critical area of bone repair in the central region of the socket, with the presence of a hypomineralized bone (osteoid) <sup>38</sup>. Additionally, ROIs delineated manually to quantify the bone neoformation process may include regions that are not part of the region of interest (e.g., cortical bone or cancellous bone outside the extraction socket) since the healing area might not be well defined. Thus, inaccurate BV/TV values may have been obtained with this method. On the other hand, standardized ROIs used as a representative sample can avoid biased results in the evaluation of bone repair <sup>6,12,24</sup>, taking into account that the ROI must be chosen stringently and consistently across samples to minimize selection bias <sup>12</sup>. Thus, the use of standardized ROIs compatible with the critical region of the socket throughout analyses is strongly recommended.

Although the bone neoformation process in extraction socket is a well-known process <sup>39,40</sup>, large bone defects (critical defect) represent major problems on the bone-healing process <sup>41</sup>, being bone grafts necessary for reconstruction. In the present study, the BV/TV values were not affected by the ROI format for the IL socket. Moreover, a significant higher amount of new bone formation was observed in the IL socket compared to the D socket, regardless of the ROI shape, demonstrating more bone formation with almost complete socket healing. This result was expected because of the difference in the root diameter. IL sockets are of smaller size and consequently leads to earlier socket healing <sup>41</sup>.

It is important to previously determine all the crucial parameters to answer the purpose of the study <sup>9</sup>. Another important aspect of the micro-CT analysis that must be taken into consideration is the size of the socket (i.e., the root to be analyzed). Studies that aim the evaluation of therapies to increase bone quality or quantity <sup>42</sup>, or in the evaluation of systemic conditions such as radiotherapy <sup>30,43</sup>, diabetes <sup>29</sup> and osteoporosis <sup>17</sup>, the evaluation of larger diameter sockets is recommended for being more critical, so the effect of such therapies or conditions can be properly evaluated.

The majority of the experimental reports lack details of the parameters applied for micro-CT image acquisition and analysis <sup>24,44</sup>. Regarding the ROI definition, few studies have clearly reported how this parameter was set. The method of the study should provide descriptions of the size and location of the ROI since the results can be affected by those parameters. Furthermore, it may be useful to provide a figure of a representative sample to illustrate the selected ROI and demonstrate the region being analyzed <sup>7,12</sup>.

Regarding the experimental period, extraction socket repair is completed in 28 days in healthy rats <sup>6,45,46</sup>. Our study is in accordance with the period of the normal course of alveolar bone repair in rats after tooth extraction, with the bone formation phase observed at 14 days.

## **5 CONCLUSION**

Within the limitations of this preclinical study, the following conclusions can be drawn:

1. Different ROIs shapes and positions within the extraction socket affect the morphometric results in micro-CT.
2. Bone neoformation outcome (BV/TV) for alveolar bone repair after tooth extraction were significantly influenced by the ROI (manual drawn or predefined shape) and the extraction socket (distal or lingual root).
3. The predefined method with standardized ROI in the central region of the bone defect in the distal extraction socket resulted in a more effective assessment of bone volume demonstrating the most critical region of the bone neoformation process.

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## **Conflict of Interest**

The authors declare that they have no conflict of interest.

## 6 REFERENCES

1. Javed A, Chen H, Ghori FY. Genetic and transcriptional control of bone formation. *Oral Maxillofac Surg Clin North Am* 2010; 22(3): 283-93. doi: 10.1016/j.coms.2010.05.001.
2. Pagni G, Pellegrini G, Giannobile WV, Rasperini G. Postextraction alveolar ridge preservation: biological basis and treatments. *Int J Dent* 2012; 2012: 151030. doi: 10.1155/2012/151030.
3. Haworth S, Shungin D, Kwak SY, Kim HY, West NX, Thomas SJ, et al. Tooth loss is a complex measure of oral disease: Determinants and methodological considerations. *Community Dent Oral Epidemiol* 2018; 46(6): 555-62. doi: 10.1111/cdoe.12391.
4. Calixto RF, Teófilo JM, Brentegani LG, Lamano-Carvalho TL. Grafting of tooth extraction socket with inorganic bovine bone or bioactive glass particles: comparative histometric study in rats. *Implant Dent* 2007; 16(3): 260-9. doi: 10.1097/ID.0b013e3180500b95.
5. Lin Z, Rios HF, Volk SL, Sugai JV, Jin Q, Giannobile WV. Gene expression dynamics during bone healing and osseointegration. *J Periodontol* 2011; 82(7): 1007-17. doi: 10.1902/jop.2010.100577.
6. Hassumi JS, Mulinari-Santos G, Fabris ALDS, Jacob RGM, Gonçalves A, Rossi AC, et al. Alveolar bone healing in rats: micro-CT, immunohistochemical and molecular analysis. *J Appl Oral Sci* 2018; 18; 26:e20170326. doi: 10.1590/1678-7757-2017-0326.

7. Bouxsein ML, Boyd SK, Christiansen BA, Guldberg RE, Jepsen KJ, Müller R. Guidelines for assessment of bone microstructure in rodents using micro-computed tomography. *J Bone Miner Res* 2010; 25: 1468-86.
8. Kruse C, Spin-Neto R, Reibel J, Wenzel A, Kirkevang LL. Diagnostic validity of periapical radiography and CBCT for assessing periapical lesions that persist after endodontic surgery. *Dentomaxillofac Radiol* 2017; 46(7): 20170210. doi: 10.1259/dmfr.20170210.
9. Irie MS, Rabelo GD, Spin-Neto R, Dechichi P, Borges JS, Soares PBF. Use of Micro-Computed Tomography for Bone Evaluation in Dentistry. *Braz Dent J* 2018; 29(3): 227-238. doi: 10.1590/0103-6440201801979.
10. Van 't Hof RJ, Dall'Ara E. Analysis of Bone Architecture in Rodents Using Micro-Computed Tomography. *Methods Mol Biol* 2019; 1914: 507-531. doi: 10.1007/978-1-4939-8997-3\_28.
11. Gaêta-Araujo H, Nascimento EHL, Brasil DM, Madlum DV, Haiter-Neto F, Oliveira-Santos C. Influence of reconstruction parameters of micro-computed tomography on the analysis of bone mineral density. *Imaging Sci Dent* 2020; 50(2): 153-159. doi: 10.5624/isd.2020.50.2.153.
12. Chavez MB, Chu EY, Kram V, de Castro LF, Somerman MJ, Foster BL. Guidelines for Micro-Computed Tomography Analysis of Rodent Dentoalveolar Tissues. *JBMR Plus* 2021;5(3): e10474. doi: 10.1002/jbm4.10474.
13. Mashiatulla M, Ross RD, Sumner DR. Validation of cortical bone mineral density distribution using micro-computed tomography. *Bone* 2017; 99: 53-61. doi: 10.1016/j.bone.2017.03.049.



14. Suttapreyasri S, Suapear P, Leepong N. The Accuracy of Cone-Beam Computed Tomography for Evaluating Bone Density and Cortical Bone Thickness at the Implant Site: Micro-Computed Tomography and Histologic Analysis. *J Craniofac Surg* 2018; 29(8): 2026-2031. doi: 10.1097/SCS.0000000000004672.
15. Chen CH, Wang L, Serdar Tulu U, Arioka M, Moghim MM, Salmon B, et al. An osteopenic/osteoporotic phenotype delays alveolar bone repair. *Bone* 2018; 112: 212-219. doi: 10.1016/j.bone.2018.04.019.
16. de Oliveira Puttini I, Gomes-Ferreira PHDS, de Oliveira D, Hassumi JS, Gonçalves PZ, Okamoto R. Teriparatide improves alveolar bone modelling after tooth extraction in orchietomized rats. *Arch Oral Biol* 2019; 102: 147-154. doi: 10.1016/j.archoralbio.2019.04.007.
17. Só BB, Silveira FM, Llantada GS, Jardim LC, Calcagnotto T, Martins MAT, et al. Effects of osteoporosis on alveolar bone repair after tooth extraction: A systematic review of preclinical studies. *Arch Oral Biol* 2021; 125: 105054. doi: 10.1016/j.archoralbio.2021.105054.
18. Christiansen BA. Effect of micro-computed tomography voxel size and segmentation method on trabecular bone microstructure measures in mice. *Bone Rep* 2016; 5: 136-40. doi: 10.1016/j.bonr.2016.05.006.
19. Djomehri SI, Candell S, Case T, Browning A, Marshall GW, Yun W, et al. Mineral density volume gradients in normal and diseased human tissues. *PLoS One* 2015; 10(4): e0121611. doi: 10.1371/journal.pone.0121611..
20. Kalatzis-Sousa NG, Spin-Neto R, Wenzel A, Tanomaru-Filho M, Faria G. Use of micro-computed tomography for the assessment of periapical lesions in small

- rodents: a systematic review. *Int Endod J* 2017; 50(4): 352-366. doi: 10.1111/iej.12633.
21. Yamasaki MC, Nejaim Y, Roque-Torres GD, Freitas DQ. Meloxicam as a Radiation-Protective Agent on Mandibles of Irradiated Rats. *Braz Dent J* 2017; 28(2): 249-255. doi: 10.1590/0103-6440201701271.
22. Yang S, Zhu L, Xiao L, Shen Y, Wang L, Peng B, et al. Imbalance of interleukin-17+ T-cell and Foxp3+ regulatory T-cell dynamics in rat periapical lesions. *J Endod* 2014; 40(1): 56-62. doi: 10.1016/j.joen.2013.09.033.
23. Chen Y, Guo Y, Li J, Chen YY, Liu Q, Tan L, et al. Endoplasmic reticulum stress remodels alveolar bone formation after tooth extraction. *J Cell Mol Med* 2020; 24(21): 12411-20. doi: 10.1111/jcmm.15753.
24. Chatterjee M, Faot F, Correa C, Duyck J, Naert I, Vandamme K. A robust methodology for the quantitative assessment of the rat jawbone microstructure. *Int J Oral Sci* 2017; 9(2): 87-94. doi: 10.1038/ijos.2017.11.
25. Wang JY, Huo L, Yu RQ, Rao NJ, Lu WW, Zheng LW. Skeletal Site-Specific Response of Jawbones and Long Bones to Surgical Interventions in Rats Treated with Zoledronic Acid. *Biomed Res Int* 2019; 2019: 5138175. doi: 10.1155/2019/5138175.
26. van Vlijmen OJ, Rangel FA, Bergé SJ, Bronkhorst EM, Becking AG, Kuijpers-Jagtman AM. Measurements on 3D models of human skulls derived from two different cone beam CT scanners. *Clin Oral Investig* 2011; 15(5): 721-7. doi: 10.1007/s00784-010-0440-8.
27. Ribeiro LL, Bosco AF, Nagata MJ, de Melo LG. Influence of bioactive glass and/or acellular dermal matrix on bone healing of surgically created defects in

- rat tibiae: a histological and histometric study. *Int J Oral Maxillofac Implants* 2008; 23(5): 811-7.
28. Truedsson A, Wang JS, Lindberg P, Gordh M, Sunzel B, Warfvinge G. Bone substitute as an on-lay graft on rat tibia. *Clin Oral Implants Res* 2010; 21(4): 424-9. doi: 10.1111/j.1600-0501.2009.01875.x.
29. Dias PC, Limirio PHJO, Linhares CRB, Bergamini ML, Rocha FS, Morais RB, et al. Hyperbaric Oxygen therapy effects on bone regeneration in Type 1 diabetes mellitus in rats. *Connect Tissue Res* 2018; 59(6): 574-580. doi: 10.1080/03008207.2018.1434166.
30. Mendes EM, Irie MS, Rabelo GD, Borges JS, Dechichi P, Diniz RS, et al. Effects of ionizing radiation on woven bone: influence on the osteocyte lacunar network, collagen maturation, and microarchitecture. *Clin Oral Investig* 2020; 24(8): 2763-771. doi: 10.1007/s00784-019-03138-x.
31. Arioka M, Zhang X, Li Z, Tulu US, Liu Y, Wang L, et al. Osteoporotic Changes in the Periodontium Impair Alveolar Bone Healing. *J Dent Res* 2019; 98(4): 450-58. doi: 10.1177/0022034518818456.
32. Miranda TS, Napimoga MH, De Franco L, Marins LM, Malta FS, Pontes LA, et al. Strontium ranelate improves alveolar bone healing in estrogen-deficient rats. *J Periodontol* 2020; 91(11): 1465-74. doi: 10.1002/JPER.19-0561.
33. Vieira AE, Repeke CE, Ferreira Junior Sde B, Colavite PM, Bigueti CC, Oliveira RC, et al. Intramembranous bone healing process subsequent to tooth extraction in mice: micro-computed tomography, histomorphometric and molecular characterization. *PLoS One* 2015; 10(5): e0128021. doi: 10.1371/journal.pone.0128021.

34. Lazenby RA, Skinner MM, Kivell TL, Hublin JJ. Scaling VOI size in 3D  $\mu$ CT studies of trabecular bone: a test of the over-sampling hypothesis. *Am J Phys Anthropol* 2011; 144(2): 196-203. doi: 10.1002/ajpa.21385.
35. Whitehouse WJ, Dyson ED. Scanning electron microscope studies of trabecular bone in the proximal end of the human femur. *J Anat* 1974; 118(Pt 3): 417-44.
36. Scala A, Lang NP, Schweikert MT, de Oliveira JA, Rangel-Garcia I Jr, Botticelli D. Sequential healing of open extraction sockets. An experimental study in monkeys. *Clin Oral Implants Res* 2014; 25(3): 288-295. doi: 10.1111/clr.12148.
37. Einhorn TA, Gerstenfeld LC. Fracture healing: mechanisms and interventions. *Nat Rev Rheumatol* 2015; 11(1): 45-54. doi: 10.1038/nrrheum.2014.164.
38. Zhang H, Chavez MB, Kolli TN, Tan MH, Fong H, Chu EY, Li Y, Ren X, Watanabe K, Kim DG, Foster BL. Dentoalveolar Defects in the Hyp Mouse Model of X-linked Hypophosphatemia. *J Dent Res*. 2020 Apr;99(4):419-428. doi: 10.1177/0022034520901719.
39. Marmary Y, Brayer L, Tzukert A, Feller L. Alveolar bone repair following extraction of impacted mandibular third molars. *Oral Surg Oral Med Oral Pathol* 1986; 61(4): 324-6. doi: 10.1016/0030-4220(86)90409-3.
40. Van der Weijden F, Dell'Acqua F, Slot DE. Alveolar bone dimensional changes of post-extraction sockets in humans: a systematic review. *J Clin Periodontol* 2009; 36(12): 1048-58. doi: 10.1111/j.1600-051X.2009.01482.x.
41. Bernabé PF, Melo LG, Cintra LT, Gomes-Filho JE, Dezan E Jr, Nagata MJ. Bone healing in critical-size defects treated with either bone graft, membrane, or a combination of both materials: a histological and histometric study in rat

- tibiae. *Clin Oral Implants Res* 2012; 23(3): 384-8. doi: 10.1111/j.1600-0501.2011.02166.x.
42. de Freitas Silva L, de Carvalho Reis ENR, Barbara TA, Bonardi JP, Garcia IR Junior, de Carvalho PSP, et al. Assessment of bone repair in critical-size defect in the calvarium of rats after the implantation of tricalcium phosphate beta ( $\beta$ -TCP). *Acta Histochem* 2017; 119(6): 624-631. doi: 10.1016/j.acthis.2017.07.003.
43. Soares PBF, Soares CJ, Limirio PHJO, Lara VC, Moura CCG, Zanetta-Barbosa D. Biomechanical and morphological changes produced by ionizing radiation on bone tissue surrounding dental implant. *J Appl Oral Sci* 2020; 28: e20200191. doi: 10.1590/1678-7757-2020-0191.
44. Faot F, Chatterjee M, de Camargos GV, Duyck J, Vandamme K. Micro-CT analysis of the rodent jaw bone micro-architecture: A systematic review. *Bone Rep* 2015; 2: 14-24. doi: 10.1016/j.bonr.2014.10.005.
45. Luvizuto ER, Dias SM, Queiroz TP, Okamoto T, Garcia IR Jr, Okamoto R, et al. Osteocalcin immunolabeling during the alveolar healing process in ovariectomized rats treated with estrogen or raloxifene. *Bone* 2010; 46(4): 1021-9. doi: 10.1016/j.bone.2009.12.016.
46. Luvizuto ER, Queiroz TP, Dias SM, Okamoto T, Dornelles RC, Garcia IR Jr, et al. Histomorphometric analysis and immunolocalization of RANKL and OPG during the alveolar healing process in female ovariectomized rats treated with oestrogen or raloxifene. *Arch Oral Biol* 2010; 55(1): 52-9. doi: 10.1016/j.archoralbio.2009.11.001.

## 7 TABLES

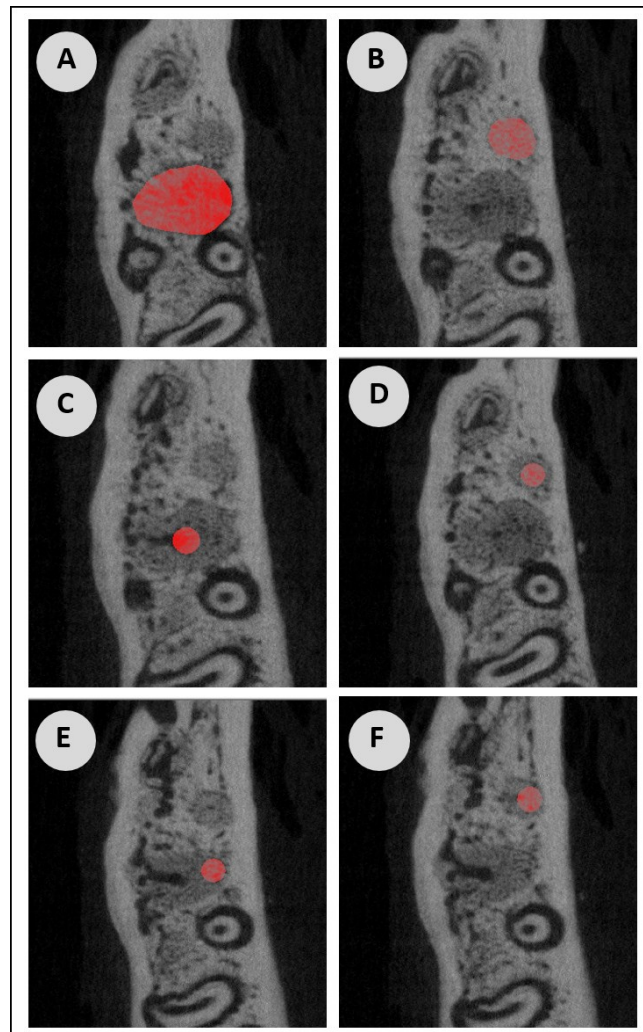
**Table 1.** Mean and standard deviation of BV/TV (%) values of the bone morphometric parameters analyzed in the alveolar repair area by micro-CT.

Extraction socket \ Region of interest	Manual ROI	Central round ROI	peripheral round ROI
<b>Distal M1</b>	75.11 ± 6.69 <sup>Ab</sup>	55.96 ± 7.35 <sup>Cb</sup>	65.31 ± 5.16 <sup>Bb</sup>
<b>Intermediate lingual M1</b>	91.38 ± 4.32 <sup>Aa</sup>	92.12 ± 5.49 <sup>Aa</sup>	93.08 ± 4.16 <sup>Aa</sup>

Different letters indicate statistically significant differences verified by Tukey test ( $p < 0.05$ ). Uppercase letters are used for comparing region of interest (manual, central round, peripheral round), lowercase letters are used for comparing extraction socket (distal and intermediate lingual root of the lower first molar (M1)).

**Figure caption**

**Fig. 1.** Demonstration of the region of interest (ROI) delimitation in the analyzed extraction socket. A) manual ROI and distal extraction socket of the lower first molar (M1). B) manual ROI and intermediate lingual extraction socket of the M1. C) central round and distal extraction socket of the M1. D) central round and intermediate lingual extraction socket of the M1. E) peripheral round and distal extraction socket of the M1. F) peripheral round and intermediate lingual extraction socket of the M1.

**Figure 1**

# Imaging Science in Dentistry

## Instructions to Authors

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Manuscripts should be prepared according to the research and publication ethics guidelines recommended by the International Committee of Medical Journal Editors (ICMJE, <http://www.icmje.org/>), Council of Science Editors (<http://www.councilscienceeditors.org/>), World Association of Medical Editors (WAME, <http://www.wame.org/>), and the Korean Association Medical Journal Editors ([https://www.kamje.or.kr/en/main\\_en](https://www.kamje.or.kr/en/main_en)).

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Studies on human subjects must have been approved by the Institutional Review Board (IRB). Also, informed consent must be obtained from the patients who participated in the study. The manuscript must include a statement of the informed consent and ethical approval including IRB information in Materials and Methods. These documents can be requested from the editor, reviewer, or publisher. In case of animal study, authors should indicate whether institutional and national guides for the care and use of laboratory animals were followed.

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## GUIDELINES FOR EACH TYPE OF MANUSCRIPT

### Original Articles

#### 1)Abstract

- This should not exceed 250 words and should be provided on a separate page.

- The abstract should be constructed under the following subheadings: Purposes, Materials and Methods, Results, and Conclusion.

- Describe each item separately in the following order.

**Purpose:** In one or two sentences, indicate the specific purpose of the article, and indicate why it is worthy of attention. The purpose stated here must be identical to the one given in the title of the paper and the introduction.

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- Do not use abbreviations or reference citations

- At the bottom of the abstract, select up to 4 key words from the current Medical Subject Headings (MeSH) in Index Medicus. Refer the website "<http://www.nlm.nih.gov/mesh/MBrowser.html>".

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Briefly describe the purpose of the investigation, including relevant background information.

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Describe the research plan, the materials (or subjects), and the methods used, in that order. When experimental methodology is the main issue of the paper, describe the process in detail so as to recreate the experiment as closely as possible. The statements for IRB and informed consent should be described in Material and Methods.

## 4)Results

Present these in a clear, logical sequence. Since biometrics involves variations in exact measurements, follow the rule of using statistics when experimentation is described. If tables are used, do not duplicate tabular data in the text, but do describe important trends and points.

## 5)Discussion

Observations pertaining to the results of research and other related materials should be interpreted for your readers. Emphasize new and important observations; do not merely repeat the contents of the results. Explain the meaning of the observed opinion along with its limits, and within the limits of the research results connect the conclusion to the purpose of the research. In a concluding paragraph, summarize the result and its meaning.

## 6)References

- Start on a separate page, numbering the references consecutively in the order in which they appear in the text.

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- Journal titles should be abbreviated according to the Index Medicus.
- All authors are to be listed when six or fewer; when there are seven or more, the first six should be given, followed by 'et al'.
- After writing the authors' last names first, the initial of their first and middle names should be capitalized.
- For all references, the starting page and the last page numbers are to be given.
- The formats are illustrated in the following examples.

**Journal article:**

Hayakawa Y, Eraso FE, Scarfe WC, Farman AG, Nishidawa K, Kuroyanagi K, et al. Modulation transfer function analysis of a newly revised rotational panoramic machine. *Dentomaxillofac Radiol* 1996; 25: 32-6.

**Complete book:**

Goaz PW, White SC. *Oral radiology; principles and interpretation*. 3rd ed. St. Louis: Mosby-Year Book Inc; 1994.

**Chapter in the book:**

Phillips SJ, Whisnant JP. Hypertension and stroke. In: Laragh JH, Brenner BM. *Hypertension: pathophysiology, diagnosis, and management*. 2nd ed. New York: Raven Press; 1995. p. 465-78.

**Journal article in press:**

Figueiredo PT, Leite AF, Freitas AC, Nascimento LA, Cavalcanti MG, Melo NS, et al. Comparison between computed tomography and clinical evaluation in tumour/node stage and follow-up of oral cavity and oropharyngeal cancer. *Dentomaxillofac Radiol* (in press).

**Abstracts:**

Mileman PA, Espelid I. Radiographic treatment decisions - a comparison between Dutch and Norwegian practitioners. *J Dent Res* 1986; 65: 609 (Abstr 32).

**Letter to the editor:**

Farman AG. Panoramic radiographic images and the prediction of asymmetry. *Dentomaxillofac Radiol* 2006; 35: 129 (letter).

**Others:**

Follow the form indicated in Uniform Requirements (1997).

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- Describe the description using one complete sentence rather than a phrase or clause.
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- Representations of microscopic images should include the magnifying power.

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- Tables are to be numbered in the order in which they are cited in the text.
- Abbreviations should be defined in an explanatory note below each table.
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Universidade Federal de Uberlândia  
 – Comissão de Ética na Utilização de Animais –



## CERTIFICADO

Certificamos que o projeto intitulado "Efeito da radioterapia na progressão do reparo ósseo e de sítios pós-extração dentária", protocolo nº 013/19, sob a responsabilidade de **Priscilla Barbosa Ferreira Soares** – que envolve a produção, manutenção e/ou utilização de animais pertencentes ao filo Chordata, subfilo Vertebrata, para fins de pesquisa científica – encontra-se de acordo com os preceitos da Lei nº 11.794, de 8 de outubro de 2008, do Decreto nº 6.899, de 15 de julho de 2009, e com as normas editadas pelo Conselho Nacional de Controle da Experimentação Animal (CONCEA), e foi **APROVADA** pela COMISSÃO DE ÉTICA NA UTILIZAÇÃO DE ANIMAIS (CEUA) da UNIVERSIDADE FEDERAL DE UBERLÂNDIA, em reunião **10 de maio de 2019**.

(We certify that the project entitled "Efeito da radioterapia na progressão do reparo ósseo e de sítios pós-extração dentária", protocol 013/19, under the responsibility of Priscilla Barbosa Ferreira Soares - involving the production, maintenance and/or use of animals belonging to the phylum Chordata, subphylum Vertebrata, for purposes of scientific research - is in accordance with the provisions of Law nº 11.794, of October 8th, 2008, of Decree nº 6.899 of July 15th, 2009, and the rules issued by the National Council for Control of Animal Experimentation (CONCEA) and it was approved for ETHICS COMMISSION ON ANIMAL USE (CEUA) from FEDERAL UNIVERSITY OF UBERLÂNDIA, in meeting of May 10th, 2019).

Vigência do Projeto	Início: 01/07/2019 Término: 01/11/2022
Espécie / Linhagem / Grupos Taxonômicos	Rato heterogêneo Wistar
Número de animais	112
Peso / Idade	300g/ 90 dias
Sexo	Macho
Origem / Local	Rede de Biotérios de Roedores da UFU – REBIR-UFU
Local onde serão mantidos os animais:	Rede de Biotérios de Roedores da UFU – REBIR-UFU

Uberlândia, 04 de Junho de 2019.

Prof. Dr. Lúcio Vilela Carneiro Girão  
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 Portaria Nº 542 DE 10 DE MAIO DE 2019