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**IDENTIFICAÇÃO E ANÁLISE DA DIVERSIDADE DE GLICOSIL HIDROLASES
DA FAMÍLIA GH18 EM ALGAS, BRIÓFITAS E PTERIDÓFITAS**

**PATOS DE MINAS - MG
MARÇO DE 2021**

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Dissertação de Mestrado apresentada ao Programa de Pós-graduação em Biotecnologia como requisito parcial para obtenção do título de mestre em Biotecnologia.

Orientador: Prof. Dr. Aulus Estevão Anjos de Deus Barbosa

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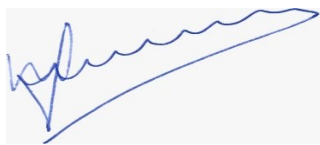
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RESUMO

O Reino Plantae é um grupo monofilético composto por organismos eucariontes e fotossintetizantes. Em termos evolutivos, as algas verdes são consideradas ancestrais das plantas terrestres, pois este grupo compartilha características importantes, como a capacidade de realizar fotossíntese e os mesmos tipos de clorofilas. Ao longo da evolução, ao se adaptarem ao ambiente terrestre, as plantas também desenvolveram importantes mecanismos fisiológicos de defesa ao ataque de predadores. Neste sentido, as quitinases vegetais são importantes proteínas que produzem resposta de defesa ao ataque de patógenos e artrópodes que contêm quitina, atuando também no desenvolvimento das plantas. As quitinases pertencem à família GH18 e GH19 das glicosil hidrolases e até o momento foram amplamente descritas em gimnospermas e angiospermas, porém pouco descritas em espécies vegetais mais primitivas. Assim, este trabalho tem por objetivo, através de um conjunto de ferramentas de bioinformática, identificar e realizar anotação das quitinases da família GH18 em algas, briófitas e pteridófitas. Inicialmente foi realizada uma busca por sequências referências de quitinases vegetais no site CAZy e a criação de um banco de dados de proteínas traduzidas a partir dos transcriptomas de 388 espécies de algas, briófitas e pteridófitas, fornecidas pela plataforma OneKP. A busca por sequências de possíveis quitinases da família GH18 neste banco de dados foi feita por meio de alinhamentos no programa BLASTp. Então, foi realizada uma análise de domínios conservados no programa Blast2GO das possíveis quitinases encontradas. Foram realizados alinhamentos múltiplos no programa MUSCLE para selecionar apenas as sequências com o domínio catalítico conservado. Para classificar as quitinases identificadas, foi realizada a construção de árvores filogenéticas utilizando o método Neighbor-Joining. No total foram encontradas 648 prováveis quitinases da família GH18, sendo 162 em algas verdes e 23 em outros filos de algas, 238 em briófitas e 225 em pteridófitas. Para comparar as relações estruturais, foi realizada a modelagem de estruturas tridimensionais no servidor Swiss-Model. Os resultados desta pesquisa são importantes para uma melhor compreensão das origens das quitinases em grupos vegetais primitivos, mostrando que estas enzimas podem ser encontradas desde as algas mais antigas e possibilitar a comparação com quitinases de plantas vasculares superiores.

Palavras-chave: Quitinases. Evolução. Transcriptômica. Defesa vegetal.

ABSTRACT

Kingdom Plantae is a monophyletic group composed of eukaryotic and photosynthetic organisms. Green algae are considered ancestors of terrestrial plants, as these groups share important characteristics, such as the ability to perform photosynthesis and the same chlorophylls types. Throughout evolution, when adapting to the terrestrial environment, plants have also developed important physiological mechanisms to defend against predators. In this sense, plant chitinases are important proteins that produce a defense response against pathogens and arthropods that contain chitin, also acting on plant development. Chitinases belong to the GH18 and GH19 family of glycosyl hydrolases and have so far been widely described in gymnosperms and angiosperms, but little described in more primitive plant species. Thus, this work aims, through a set of bioinformatics tools, to identify and annotate chitinases of the GH18 family in algae, bryophytes, and pteridophytes. Initially, a search for plant chitinases reference sequences was carried out on the CAZy website, and the creation of protein database translated from transcriptomes of 388 algae, bryophytes, and pteridophytes species, provided by the OneKP platform. Search for sequences of putative chitinases of the GH18 family in this database was done through alignments in the BLASTp program. Then, was performed a conserved domains analysis, using Blast2GO, of the putative chitinases found. Multiple alignments were performed with MUSCLE method to select only the sequences with the conserved catalytic domain. To classify identified chitinases, phylogenetic trees were constructed using Neighbor-Joining method. In total, 648 putative chitinases from the GH18 family were found, 162 in green algae and 23 in other algae phyla, 238 in bryophytes, and 225 in pteridophytes. To compare structural relationships, three-dimensional structures were modeled on the Swiss-Model server. The results of this research are important for a better understanding of chitinases origins in primitive plant groups, showing that these enzymes can be found from the oldest algae and enable the comparison with chitinases of superior vascular plants.

Keywords: Chitinases. Evolution. Transcriptomic. Plant defense.

LISTA DE ABREVIATURAS E SIGLAS

GH – Glicosil hidrolases

GH18 – Glicosil hidrolases da família 18

GH19 – Glicosil hidrolases da família 19

BLASTp – Basic Local Alignment Search Tool protein

MEGA – Molecular Evolutionary Genetics Analysis

MUSCLE – Multiple Sequence Comparison by Log-Expectation

NCBI – National Center for Biotechnology

PDB – Protein Data Bank

PR – Pathogenesis Related

JTT – Jones Taylor Thornton

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CAPÍTULO I – REFERENCIAL TEÓRICO

1 INTRODUÇÃO

1.1 Problema de pesquisa

O Reino Vegetal é formado por organismos fotossintetizantes organizados em diferentes filos de algas, briófitas e plantas vasculares, incluindo pteridófitas, gimnospermas e angiospermas. Ao longo da evolução, as plantas foram adquirindo tecidos especializados, o que favoreceu a sua adaptação e ocupação do ambiente terrestre (INGROUILLE; EDDIE, 2006). Evidências apontam que as plantas vasculares evoluíram a partir de algas verdes, devido às semelhanças compartilhadas entre esses grupos, como por exemplo, a presença de pigmentos fotossintetizantes armazenados em plastídios e parede celular constituída por celulose (CASANOVA; NAIRN, 2016).

Ao longo do processo evolutivo, desde as algas, foram selecionados uma série de mecanismos fisiológicos de defesa contra o ataque de predadores. Sabe-se que em plantas vasculares as proteínas quitinases estão relacionadas a mecanismos de defesa contra o ataque de patógenos e artrópodes que contém quitina, pois essas proteínas agem promovendo a hidrólise do polímero de quitina (KESARI et al., 2015). Além dessa importante função, tem sido relatado que as quitinases também desempenham papel no desenvolvimento e crescimento das plantas terrestres (EDREVA, 2005; HAMID et al., 2013). Uma grande diversidade de quitinases são atualmente estudadas em plantas gimnospermas e angiospermas, mas até o momento, pouco relatadas em algas, briófitas e pteridófitas.

Diversos estudos sugerem que as quitinases passaram por um rápido processo evolutivo, uma vez que muitas quitinases, até então estudadas, demonstraram grandes diferenças em suas sequências de aminoácidos, conformação estrutural e até mesmo mecanismos enzimáticos diferentes (FUKAMIZO, 2000). Há uma grande diversidade de quitinases relatadas em plantas vasculares superiores, mas será que os grupos mais ancestrais das plantas vasculares mais recentes apresentam também essa grande variedade de quitinases? As quitinases em algas, briófitas e pteridófitas divergem muito em similaridade de sequências e estruturas tridimensionais quando comparadas com as quitinases de plantas vasculares superiores?

1.1 Hipótese

Até o momento, estudos têm relatado a identificação de quitinases em diversas plantas vasculares superiores (gimnospermas e angiospermas), mas os estudos de quitinases em algas, briófitas e pteridófitas ainda são limitados. Pressupõe-se que nesses grupos ancestrais há também uma grande diversidade de quitinases, que possam ter diferenças significativas com as quitinases de plantas vasculares superiores, uma vez que é sugerido que as quitinases vem sofrendo rápidas adaptações evolutivas.

1.2 Objetivos

1.2.1 Objetivo geral

Realizar investigação e anotação de proteínas quitinases nos grupos ancestrais das plantas terrestres, desde algas, briófitas e pteridófitas, por meio de um conjunto de ferramentas de bioinformática.

1.2.2 Objetivos específicos

- Identificar e analisar as sequências de aminoácidos de prováveis proteínas de espécies de algas, briófitas e pteridófitas para verificar se há presença de proteínas quitinases;
- Analisar se as proteínas identificadas são quitinases, verificando a presença do domínio catalítico característico da família GH18;
- Classificar as quitinases identificadas da família GH18 por meio de análise filogenética;
- Modelar estruturas tridimensionais de quitinases de cada grupo analisado para comparar as relações estruturais.

1.3 Justificativa

Sabe-se que as quitinases da família GH18 são proteínas presentes em plantas e em diversos organismos. Possuem funções importantes para o desenvolvimento e crescimento das plantas, principalmente relacionadas a defesa contra o ataque de patógenos e artrópodes que contém quitina, uma vez que essas enzimas promovem a hidrólise do polímero de quitina. A

maioria dos estudos relatam análises de quitinases em gimnospermas e angiospermas, no entanto, existem poucos dados sobre quitinases de algas, briófitas e pteridófitas disponíveis. Devido à importância dessas proteínas no Reino vegetal e para compreender melhor a dinâmica dessas proteínas em algas, briófitas e pteridófitas, este estudo busca investigar e realizar anotação da diversidade de quitinases da família GH18 nesses grupos vegetais distintos e mais primitivos. A anotação de dados poderá ser útil para o fornecimento de informações para estudos futuros e auxiliar na compreensão das relações evolutivas entre quitinases de grupos vegetais ancestrais com as plantas vasculares superiores.

2 REFERENCIAL TEÓRICO

2.1 Origem e evolução das plantas

O Reino Plantae é um grupo monofilético composto por mais de 370.000 espécies de organismos eucariontes, geralmente autotróficos, e adaptados ao ambiente terrestre (CHRISTENHUSZ; BYNG, 2016). Este grande grupo abrange alguns tipos de algas unicelulares e multicelulares, briófitas, pteridófitas, gimnospermas e angiospermas (BENNICI, 2008). Estão incluídos neste grupo organismos eucariontes que possuem cloroplastos, realizam fotossíntese através da clorofila e armazenam os produtos da fotossíntese, como o amido, além de possuírem parede celular constituída por celulose (BLACKWELL, 2003; NISHIO, 2000). As plantas são seres pluricelulares com elevado nível de organização e especialização, enquanto em algas não existe diferenciação de tecidos, mas sim um conjunto de células, algumas podendo apresentar certa especialização (NISHIO, 2000). Atualmente as plantas terrestres são agrupadas no clado Embriófitas, que incluem briófitas, pteridófitas, gimnospermas e angiospermas (EVERT; EICHHORN, 2014).

Evidências evolutivas indicam que as algas verdes são ancestrais das plantas terrestres, com registros fósseis relatados de 900 milhões de anos (INGROUILLE; EDDIE, 2006). Enquanto, que a expansão das plantas no ambiente terrestre ocorreu por volta de 470 milhões de anos atrás (ISHIZAKI, 2017). Estudos indicam que a origem das plantas é baseada a partir de uma cianobactéria de vida livre (ancestral procariótico comum) que são eubactérias gram-negativas referidas como "algas azuis-verdes", das quais derivaram os plastídios; organelas que acumulam pigmentos fotossintetizantes e possibilitam assim, a realização da fotossíntese, e essa teoria é apoiada pela semelhança do material genético que os constituem (MARTIN; GARG; ZIMORSKI, 2015; MCFADDEN, 2014). Algumas cianobactérias apresentam clorofilas *a* e *b*,

pigmentos carotenoides e membranas tilacoides, assim como plastídios; outras apresentam ficobilina, que são pigmentos encontrados em algas vermelhas, e que reforça ainda mais essa teoria (BHATTACHARYA; YOON; HACKETT, 2004).

A teoria do surgimento de plastídios em algas verdes e plantas envolve um processo de endossimbiose primária de uma cianobactéria por protistas não fotossintéticos, enquanto a diferença de complexidade entre os outros tipos de algas é descrito por um evento de endossimbiose secundária que envolve o englobamento de um plastídio primário (como é o caso de algumas algas euglenoides que se tornaram fotossintéticas por englobar o plastídio de uma alga verde fotossintética), e até mesmo por endossimbioses terciárias por englobamento de plastídios secundários (observado em algas que apresentam de três a quatro membranas envolvendo as organelas fotossintéticas) (BHATTACHARYA; YOON; HACKETT, 2004; MARTIN; GARG; ZIMORSKI, 2015). Ao longo do processo evolutivo das plantas, diversas alterações significantes ocorreram no ambiente, o que favoreceu o desenvolvimento de caracteres adaptativos as novas condições (ANDERSON; WILLIS; MITCHELL-OLDS, 2011). Assim, as plantas terrestres desenvolveram estruturas adaptativas mais complexas, como vasos condutores de nutrientes e água, presença de embrião, além de apresentarem órgãos reprodutivos complexos, como o órgão reprodutor feminino, denominado de arquegônio, e o órgão reprodutor masculino, conhecido como anterídio (GRAHAM; COOK; BUSSE, 2000; ISHIZAKI, 2017).

2.2 Algas

Assim como as plantas terrestres, as algas realizam papel importante no habitat em que vivem, podendo ser encontradas em regiões rochosas, em água doce e águas marinhas, e até mesmo em regiões desérticas (PANAWALA, 2017a). Uma característica marcante em algas, assim como em plantas terrestres, é a capacidade de realizarem fotossíntese, atuando, portanto, no ciclo do carbono com a transformação de dióxido de carbono (CO_2) em carboidratos e em carbonato de cálcio (RAVEN et al., 2012). Além da capacidade de consumir dióxido de carbono, que é um dos principais gases do efeito estufa, e produzir oxigênio, as algas são a base essencial na cadeia alimentar de muitos organismos marinhos, alguns de água doce e em menor parte cadeias alimentares terrestres (PANAWALA, 2017a).

Existem vários filos de algas conhecidos, além do importante grupo de algas verdes, abrangendo um número de aproximadamente 70.000 espécies vivas e extintas, com diferenças significativas, que foram classificadas de acordo com sua fisiologia (INGROUILLE; EDDIE,

2006). Entre elas, encontram-se as algas euglenoides (filo Euglenophyta), que são consideradas uma das linhagens mais antigas; as criptófitas (filo Cryptophyta); haptófitas (filo Haptophyta); dinoflagelados (filo Dinophyta); diatomáceas (filo Bacillariophyta); algas douradas (filo Chrysophyta); algas verde-amarelas ou xantofíceas (filo Heterokontophyta); algas pardas (filo Ochrophyta); algas vermelhas (filo Rhodophyta) e algas verdes (filo Chlorophyta) (RAVEN; GIORDANO, 2014). A diferença evolutiva nos filios de algas está relacionada principalmente ao tipo de pigmento que constituem seus plastídios. Algumas algas apresentam clorofilas *a* e *b* ou *c* em suas estruturas, podem armazenar pigmentos carotenoides, enquanto outras podem armazenar o pigmento ficobilina (CAVALIER-SMITH, 1982; LEWIS; MCCOURT, 2004).

2.2.1 Algas verdes

As algas verdes constituem um grande grupo diversificado de aproximadamente 8,000 espécies, que variam de organismos simples unicelulares até organismos multicelulares complexos. Podem ser encontradas, em grande parte, em regiões aquáticas, principalmente água doce, com exceções de alguns grupos que podem se desenvolver em regiões extremas, como a superfície de neve, desertos, tronco de árvores, solo e também em associações simbióticas com líquens, fungos, protozoários de água doce, esponjas e celenterados (GUIRY, 2012; LEWIS; MCCOURT, 2004).

As algas verdes apresentam semelhanças com briófitas e plantas vasculares, uma vez em que ambas armazenam amido nos plastos, possuem parede celular constituída por celulose e sintetizam clorofilas *a* e *b* (LEWIS; MCCOURT, 2004). O grupo de algas verdes (filo Chlorophyta) é dividido em três principais classes: Chlorophyceae, Charophyceae e Ulvophyceae (COOPER, 2014). A classe Chlorophyceae (conhecidas por clorófitas) abrange algas verdes com características de morfologia e ecologia diferentes das demais algas deste grupo, podendo haver clorófitas unicelulares e multicelulares, de complexidade diferente (FANG et al., 2017). A classe Charophyceae (carófitas) é composta principalmente por algas verdes de água doce, com algumas espécies habitando regiões de águas salinas e de vida livre em terra, enquanto Ulvophyceae (ulvófitas) é uma classe de menor dimensão, composta principalmente por algas marinhas, filamentosas ou lâminas achatadas (RAVEN; GIORDANO, 2014).

No filo de algas verdes, é evidenciado que algas da classe Charophyceae são as que possuem maior grau de parentesco com briófitas e plantas vasculares, devido a similaridades

morfológicas, bioquímicas e genéticas observadas entre esses grupos (BENNICI, 2008; ISHIZAKI, 2017). Carófitas e briófitas armazenam clorofilas *a* e *b* e betacaroteno como pigmentos fotossintetizantes; armazenam alimento na forma de amido e possuem parede celular constituída por celulose (BLACKWELL, 2003). Devido a preservação dessas características no decorrer da evolução, as algas verdes são consideradas ancestrais das plantas terrestres (denominadas embriófitas), e por isso são muito estudadas para compreender a origem e evolução das plantas (CASANOVA; NAIRN, 2016).

2.3 Briófitas

O grupo das briófitas é constituído por aproximadamente 25,000 espécies de plantas que possuem crescimento regulado por células apicais, tecidos pouco diferenciados, além de não possuírem vasos condutores especializados em transporte de água e nutrientes (FRAHM et al., 2003; PITTERMANN, 2010). As briófitas são plantas de pequeno porte, justamente por não possuírem tecido vascular, e habitam geralmente regiões úmidas e sombreadas, junto ao solo, sobre rochas ou sobre outras plantas; algumas espécies podem ainda se desenvolver em regiões desérticas, sob condições extremas de calor (PANAWALA; 2017a).

As briófitas apresentam tecidos de maior especialização e muitas características que não estão presentes em algas verdes, mas que são compartilhadas com pteridófitas, como os gametângios masculinos e femininos, denominados anterídios e arquegônios, e tecidos produzidos por um meristema apical (EVERT; EICHHORN, 2014). Assim, devido às semelhanças compartilhadas entre esses grupos, estudos evolutivos apontam as briófitas como as primeiras plantas terrestres simples, derivadas diretamente de algas verdes, sendo consideradas, portanto, os ancestrais das pteridófitas, auxiliando na compreensão do surgimento e evolução das plantas na Terra (BENNICI, 2008; PANAWALA; 2017a).

As briófitas são divididas em três filos: Marchantiophyta (hepáticas), Bryophyta (musgos) e Anthoceroophyta (antóceros) (CASANOVA; NAIRN, 2016; PUTTICK et al., 2018). As briófitas hepáticas, são geralmente talosas ou folhosas; os musgos são as briófitas mais conhecidas e espalhadas pelo mundo; enquanto que o grupo dos antóceros é considerado o menos diversificado de briófitas (FRANGEDAKIS et al., 2020; HE-NYGRÉN et al., 2006).

2.4 Pteridófitas

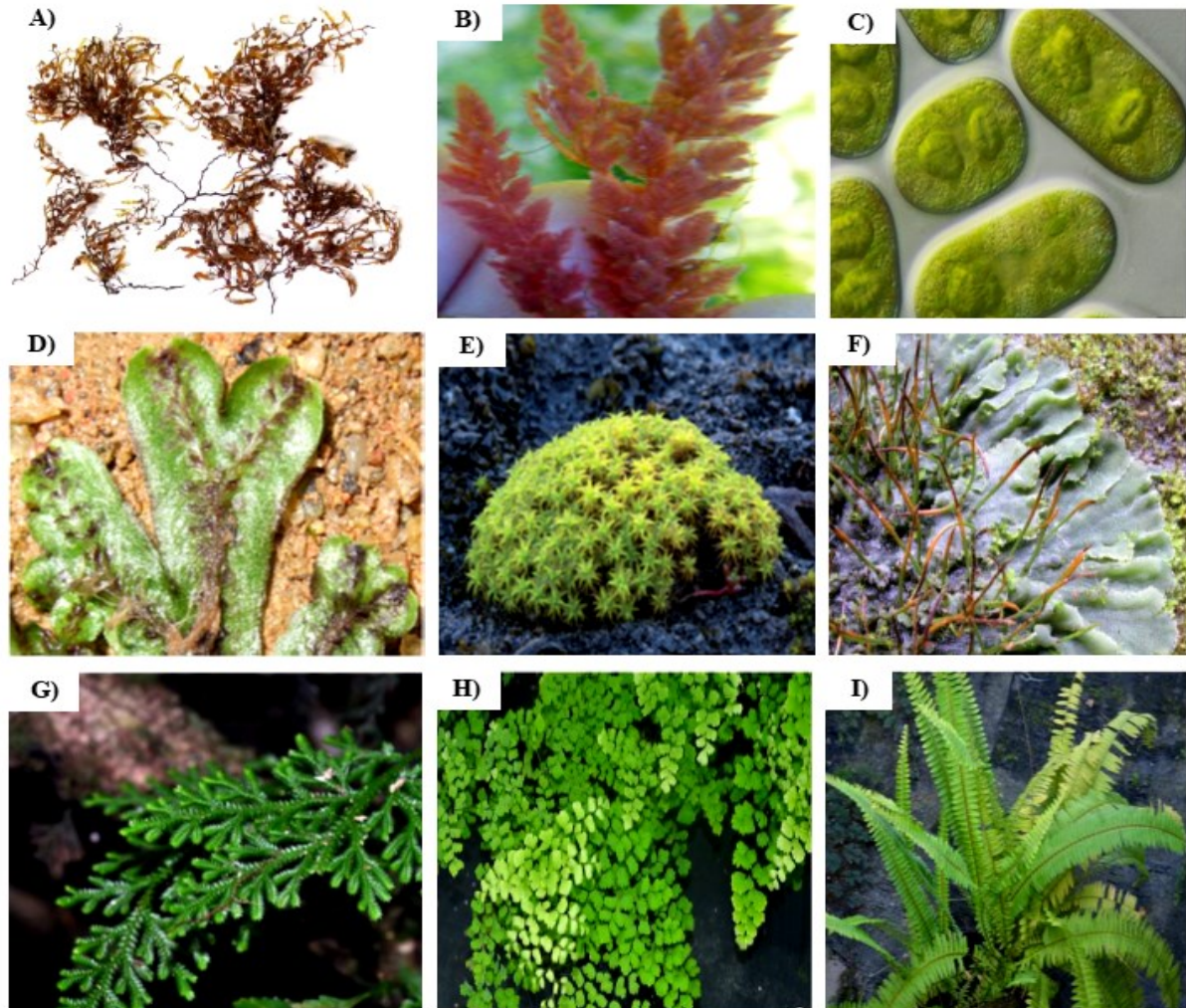
As primeiras pteridófitas surgiram a cerca de 400 milhões de anos durante a Era Paleozoica, dos períodos Siluriano ao Devoniano. As pteridófitas são plantas vasculares sem sementes, com aproximadamente 10.000 espécies, encontradas em diferentes habitats que incluem rochas, solo, ambientes escuros e úmidos, predominantemente em regiões tropicais (TRYON, R. M; TRYON, A. F, 1982).

Um importante estágio inicialmente observado em pteridófitas, no decorrer da evolução, foi o estabelecimento de um esporófito livre e independente, o que contribuiu para o processo de adaptação ao ambiente terrestre, uma vez que possibilitou maior resistência e um maior crescimento vegetal (BLACKWELL, 2003). Outra característica importante que surgiu nas pteridófitas foi a presença de vasos condutores, xilema e floema, o que contribuiu para a sobrevivência das plantas, pois facilitou o processo de transporte de água e nutrição para plantas maiores (PANAWALA, 2017b). O surgimento de heteroesporia ocorreu também primeiramente em pteridófitas, nos quais as plantas passaram a produzir esporos diferentes por meiose, sendo estes denominados micrósporos e megásporos, originando respectivamente, os gametófitos masculinos e femininos (WILLIS; MCELWAIN, 2002).

Atualmente, o grupo de plantas pteridófitas compreende as divisões Lycophyta e Monilophyta (samambaias e cavalinhas) (CHRISTENHUSZ; CHASE, 2014; PRYER et al., 2004). Este grupo também era composto pelos filos Rhyniophyta, Zosterophyllophyta e Trimerophytophyta que entraram em extinção no final do Devoniano. O estudo de pteridófitas é importante para a melhor compressão do processo evolutivo das plantas, com ênfase na evolução de gimnospermas e angiospermas.

A figura 1 contém exemplos de espécies das principais divisões dos filos de algas, briófitas e pteridófitas.

Figura 1 - Exemplos de espécies de algas, briófitas e pteridófitas. **A)** Alga marrom do filo Ochrophyta (espécie *Sargassum natans*). **B)** Alga vermelha do filo Rhodophyta (espécie *Asparagopsis armata*). **C)** Alga verde unicelular carófitas (espécie *Cylindrocystis crassa*). **D)** Briófitas do filo Marchantiophyta (espécie *Marchantia emarginata*). **E)** Musgo da espécie *Syntrichia ruralis*. **F)** Briófitas do filo Anthocerotophyta (espécie *Phaeomegaceros hirticalyx*). **G)** Pteridófitas do clado Lycophyta (espécie *Selaginella involvens*). **H)** Pteridófitas do clado Monilophyta conhecida como avenca (espécie *Adiantum capillus-veneris*). **I)** Pteridófitas do clado Monilophyta conhecida como samambaia (espécie *Nephrolepis cordifolia*).



Fonte: Baskaran et al. (2018); Cotas et al. (2020); Ho (2013); Huttunen; Bell e Hedenäs (2018); Mendez-Tejeda e Rosado (2019); Samolov et al. (2020); Villarreal et al. (2014).

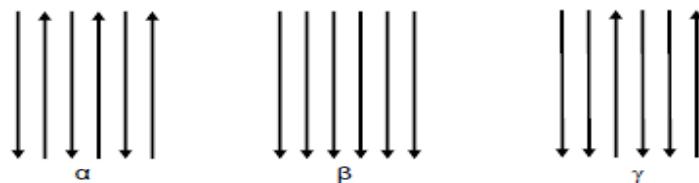
2.5 Quitina

A quitina foi isolada pela primeira vez a partir de fungos em um estudo realizado por Braconnot (1811). A quitina é considerada o segundo polissacarídeo mais abundante da natureza (MARTÍNEZ-CABALLERO et al., 2014), com a celulose se destacando em primeiro lugar (CAMPANA-FILHO et al., 2007). É um polímero linear, insolúvel, composto por uma longa cadeia de unidades repetitivas de 2-acetamido-2-desoxi-D-glucose, geralmente designado também de N-acetil-D-glucosamina (GlcNAc), que são unidos por ligação glicosídicas β -(1-4), (SEIDL, 2008).

A quitina é um biopolímero que compõe a matriz da estrutura esquelética de muitos invertebrados, estando presente no exoesqueleto de artrópodes, moluscos e celenterados, em algas diatomáceas, e também nas paredes celulares de determinados fungos, desempenhando função de revestimento e materiais de suporte aos organismos (CAMPANA-FILHO et al., 2007).

Existem na natureza três formas diferentes de quitina as quais diferem na disposição das suas cadeias. Na forma α as cadeias estão orientadas no sentido antiparalelo, na forma β as cadeias estão dispostas no sentido paralelo e em γ ocorre um misto de cadeias paralelas e antiparalelas (Figura 2) (SEIDL, 2008). Essas diferenças estruturais conferem a quitina funções diferentes nos organismos, na qual a forma α fornece maior rigidez, enquanto as formas β e γ conferem maior flexibilidade e resistência aos organismos (KAYA et al., 2017).

Figura 2 - Orientação das cadeias poliméricas de quitina. Cadeias α são posicionadas no sentido antiparalelo, na forma β as cadeias são dispostas no sentido paralelo, enquanto na forma γ ocorre um misto de cadeias α e β .



Fonte: Matsiu (2007).

A quitina é um biopolímero com alta versatilidade e é muito utilizada na indústria de alimentos como agente floculante, no tratamento de efluentes, como adsorvente na clarificação de óleos, e em grande parte na produção de quitosana (MOURA et al., 2006).

2.6 Quitinases em plantas

As plantas não contêm quitina em seus constituintes, porém, possuem genes que expressam quitinases de forma constitutiva ou induzível (OHNUMA et al., 2011a). As plantas geralmente expressam níveis relativamente baixos de quitinases e só sintetizam níveis mais elevados da enzima quando entram em contato com fitopatógenos que contem quitina. Além de estar presente em plantas, as quitinases também são amplamente encontradas em crustáceos, bactérias e fungos, as quais podem desempenhar diferentes funções. Em plantas, as quitinases podem ser expressas em caules, sementes, tubérculos e flores (KHAN et al., 2015).

As plantas produzem uma série de mecanismos de defesa, e entre eles destaca-se a expressão de um complexo de proteínas relacionadas a patogênese, denominadas proteínas PR (*Pathogenesis-Related*) (NEUHAUS et al., 1996). As quitinases estão incluídas neste grupo e são vistas como proteínas de defesa quando submetidas a estresses bióticos e abióticos. Essas proteínas apresentam a capacidade de induzir resistência local ou sistêmica ao ataque de fitopatógenos, como fungos patogênicos, por agir diretamente no polímero de quitina, que é um dos principais componentes da parede celular da maioria dos fungos (KESARI et al., 2015). Algumas quitinases relacionadas a patogênese exibem também atividades antibacteriana, nematicidas e antivirais (CHEN et al., 2015; MEDEIROS et al., 2018; KAUR et al., 2019)

As quitinases de plantas podem também apresentar um nível maior de expressão quando induzidas por fatores abióticos (CAO; TAN, 2019). Determinadas condições ambientais extremas, como elevadas concentrações salinas, longos períodos de frio e estresse hídrico podem promover a transcrição de alguns genes de quitinase, sugerindo assim, que essas proteínas possam estar envolvidas com mecanismos de resposta ao estresse em plantas (BADARIOTTI et al., 2006; CAO; TAN, 2019). Também é sugerido a atuação de quitinases no crescimento e desenvolvimento vegetal, e podem ser principalmente detectadas nos estágios iniciais de crescimento (EDREVA, 2005; HAMID et al., 2013).

2.7 Características de quitinases

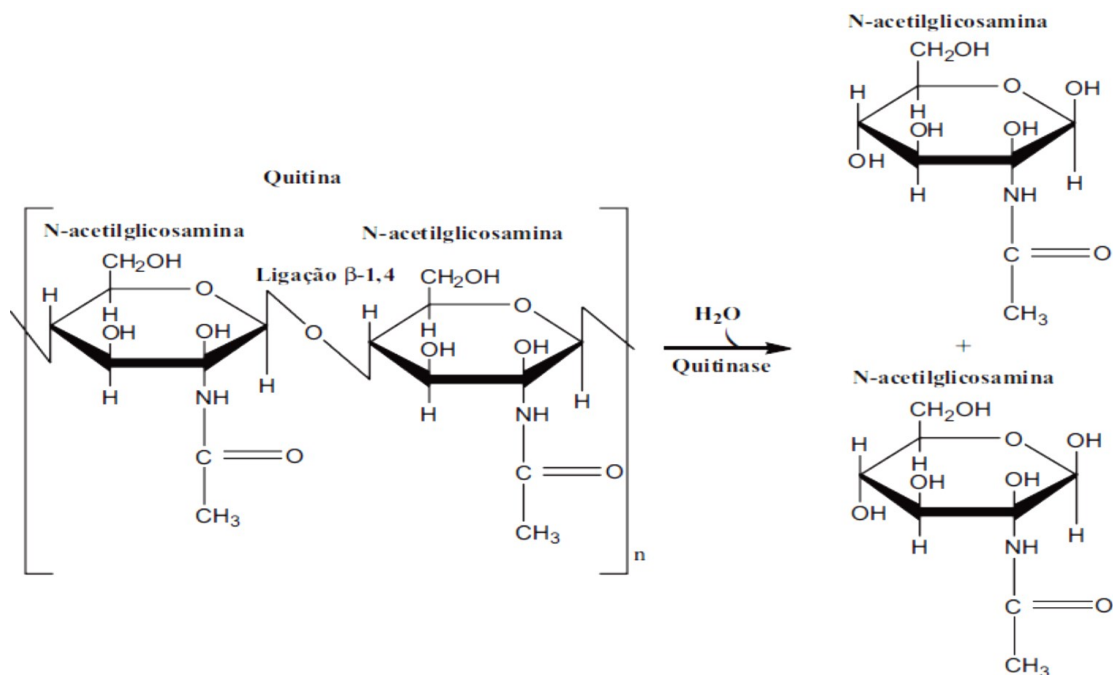
As quitinases são enzimas que catalisam a hidrólise das ligações glicosídicas β -1,4 entre os resíduos de N-acetilglucosamina (GlcNAc) que compõem o polímero de quitina (Figura 3) (MARTÍNEZ-CABALLERO et al., 2014; SEIDL et al., 2005).

Dependendo de seus padrões de clivagem, que estão relacionados ao tipo de produto formado após a hidrólise do substrato, as quitinases podem ser divididas em duas categorias: endoquitinases e exoquitinases. De acordo com o Comitê de Nomenclatura da União Internacional de Bioquímica e Biologia Molecular, as quitinases podem ser classificadas em três grupos de enzimas com três números distintos (números E.C., do inglês Enzyme Commission): (i) endoquitinases (E.C. 3.2.1.14), que clivam aleatoriamente ligações glicosídicas internas ao longo do polímero de quitina e geram múltímeros de GlcNAc com alto peso molecular, (ii) exoquitinases que clivam na extremidade não redutora da cadeia polimérica, levando a formação de monômeros de N-acetilglucosamina (E.C. 3.2.1.200) e (iii) exoquitinases que catalisam a liberação de N-acetilglucosamina a partir da extremidade redutora da quitina (E.C. 3.2.1.201) (BRZEZINSKA et al., 2014; SEIDL, 2008). Por estarem

mais amplamente distribuídas entre os organismos, as endoquitinases são mais estudadas do que as exoquitinases (HAN et al., 2016).

As quitinases vegetais ainda podem apresentar isoformas básicas e ácidas, de acordo com seus pontos isoelétricos, e são provenientes de regiões diferentes das plantas. Quitinases básicas são encontradas no vacúolo de plantas, enquanto quitinases ácidas são frequentemente secretadas para regiões extracelulares (LEGER et al., 1996).

Figura 3 - Hidrólise de quitina por proteínas quitinases.



Fonte: Fleuri e Sato (2005).

2.8 Classificação das quitinases

As quitinases pertencem a superfamília das glicosil hidrolases (GH), uma família de enzimas que hidrolisam ligações glicosídicas em carboidratos (NEUHAUS et al., 1996). Baseando-se na semelhança na sequência de aminoácidos do domínio catalítico de quitinases e no mecanismo de catálise, as quitinases vegetais, e foram divididas em famílias GH18 e GH19 das glicosil hidrolases (KESARI et al., 2015; LOMBARD et al., 2014). A família GH18 possui sequências de aminoácidos com baixa similaridade entre si, abrangendo quitinases de diversos organismos como vírus, bactérias, fungos, animais e algumas plantas. Já a família GH19 possuem sequências de alta similaridade entre si, e é composta principalmente por quitinases

vegetais, podendo incluir também algumas quitinases bacterianas (PATIL; GHORMADE,; DESHPANDE, 2000).

Baseando-se em análises evolutivas, acredita-se que as quitinases das famílias GH18 e GH19 tenham evoluído de ancestrais diferentes, pois não compartilham similaridade em suas sequências de aminoácidos, e apresentam estruturas tridimensionais e mecanismos enzimáticos diferentes (FUKAMIZO, 2000). Apesar de quitinases da família GH18 apresentarem baixa similaridade com as quitinases da família GH19, suas sequências se assemelham às de quitinases de fungos e bactérias (KESARI et al., 2015).

Inicialmente, as quitinases vegetais foram divididas em seis classes (I-VI) (NEUHAUS et al., 1996). Posteriormente, foi identificada uma nova classe de quitinases, com características diferentes das quitinases anteriormente caracterizadas, as quais foram designadas de classe VII (JI; JINYUAN, 2002). As classes III e V pertencem à família GH18, enquanto as classes I, II, IV, VI e VII são integrantes da família GH19 (CAO; TAN, 2019). A maioria das quitinases vegetais estão distribuídas entre as classes I, II, III, IV e V.

As quitinases de classe I apresentam um domínio de ligação a quitina rico em cisteína na região N-terminal, contendo aproximadamente 40 aminoácidos, e um domínio catalítico C-terminal, altamente conservado, e geralmente são localizadas no vacúolo das plantas (ARIE et al., 2000; SARMA et al., 2012). As quitinases de classe II, são semelhantes às quitinases de classe I, porém apresentam apenas o domínio catalítico e não apresentam o domínio de ligação a quitina, e são frequentemente secretadas para o espaço extracelular (apoplasto). A classe III é composta por quitinases que apresentam baixa similaridade com as classes I e II, não apresentam domínio de ligação a quitina e são geralmente secretadas para o apoplasto (ARIE et al., 2000). Na classe IV encontram-se quitinases semelhantes a classe I, com a presença de domínios catalítico e de ligação a quitina, porém são de tamanhos relativamente menores, apresentando algumas deleções nos domínios catalíticos (ARIE et al., 2000; MARTÍNEZ-CABALLERO et al., 2014). Quitinases dessa classe tem sido identificadas, principalmente, em plantas dicotiledôneas (EL-KATATNY et al., 2001). Já a classe V, compreende um grupo de quitinases que geralmente possuem dois domínios de ligação a quitina e um domínio catalítico, de pouca semelhança com as demais classes (EL-KATATNY et al., 2001). As quitinases de classe VI possuem os domínios de ligação a quitina duplicados na região N-terminal, enquanto as quitinases de classe VII não possuem domínios de ligação a quitina (SU et al., 2015).

As proteínas relacionadas a patogênese (PR) também são divididas em famílias, de acordo com o grau de identidade de sequências e com a atividade biológica. Entre as 17 famílias

atualmente descritas de proteínas PR, as quitinases estão distribuídas nas famílias PR-3, 4, 8 e 11 (SELS et al., 2008).

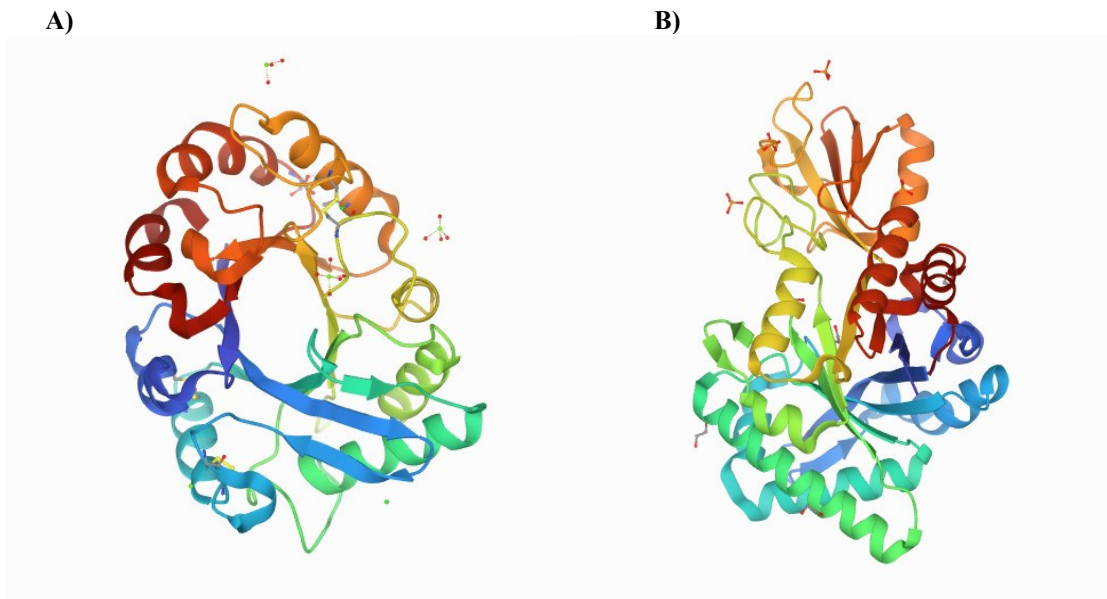
As plantas, em geral, apresentam uma grande quantidade de proteínas semelhantes à quitinases (CLPs) que também fazem parte das famílias GH18 e GH19 das glicosil hidrolases e, assim como as quitinases, também apresentam o domínio catalítico, porém, devido a modificações de resíduos de aminoácidos, ocorre a perda da função catalítica (MARTÍNEZ-CABALLERO et al., 2014). Dessa forma, as CLPs têm a capacidade de se ligarem a quitina, mas não conseguem promover a hidrólise das ligações glicosídicas, e, portanto, não realizam catálise.

2.9 Família GH18

A família GH18 é composta por endoquitinases e exoquitinases que possuem um domínio catalítico (C-terminal) caracterizado por uma estrutura de barril (α / β) constituído por 8 folhas- β e 8 α -hélices (Figura 4) (BADARIOTTI et al., 2006; CHUANG; LIN, H. Y.; LIN, F. P, 2008). Além do domínio catalítico, essas proteínas também podem apresentar domínios de ligação a quitina, situados na região N-terminal, que são regiões que não realizam catálise, mas que intensificam a afinidade da quitinase ao substrato, o que pode proporcionar um aumento da eficiência da hidrólise da quitina (YAN; FONG, 2015). Os domínios de ligação a quitina geralmente são conectados ao domínio catalítico por ligantes, o que contribui para que os domínios funcionem e evoluam de uma forma independente (KARLSSON; STENLID, 2009). Muitas quitinases GH18 possuem também um peptídeo sinal, que as direciona para serem secretadas para o espaço extracelular ou para uma localização subcelular (SEIDL et al., 2005).

As quitinases da família GH18 realizam reações de hidrólise das ligações glicosídicas pelo mecanismo de retenção da configuração anomérica, formando produtos com conformação β (BRAMELD et al., 1998; SEIDL et al., 2005).

Figura 4 - Estruturas tridimensionais de quitinases da família GH18 constituídas por α -hélices e folhas- β . **A)** Quitinase de classe III de *Punica granatum*. **B)** Quitinase de classe V de *Nicotiana Tabaccum*.



Fonte: Masuda; Zhao; Mikami (2015) e Ohnuma et al., (2011b).

A família GH18 é, portanto, constituída de quitinases de classes III e V. As enzimas da classe III apresentam semelhanças com quitinases bacterianas, pois compartilham do mesmo mecanismo de hidrólise e similaridade na estrutura tridimensional (ISELI et al., 1996). A classe III tem uma característica peculiar, e são consideradas enzimas bifuncionais, pois além de hidrolisar quitina, também possuem atividade de lisozima, podendo assim, contribuir para a proteção contra infecções bacterianas (PARK et al., 2002). A função biológica principal da família GH18 em plantas está relacionada a degradação da quitina, mas em outros organismos elas podem desempenhar outras funções. Em fungos filamentosos a família GH18 está relacionada aos processos de degradação e modificação da parede celular, germinação de esporos, ramificação de hifas, diferenciação de esporos, autólise e micoparasitismo (ADAMS, 2004). Muitas espécies de bactérias secretam quitinases para utilizar quitina como uma fonte nutritiva de carbono e nitrogênio ou para defesa contra outros microrganismos (BHATTACHARYA; YOON; HACKETT, 2004). Por exemplo, em *Crassostrea gigas* (ostra-do-pacífico) as quitinases da família GH18 estão envolvidas no desenvolvimento embrionário inicial e funcionam como efetores da imunidade (BADARIOTTI, 2006)

CAPÍTULO II – ARTIGO CIENTÍFICO

Identification and analysis of GH18 glycosyl hydrolases in algae, bryophytes, and pteridophytes

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ABSTRACT

The Plantae Kingdom is a monophyletic group composed of eukaryotic and photosynthetic organisms. Throughout evolution, to adapt to the terrestrial environment, plants have developed important physiological defense mechanisms against predators and pathogens. Plant chitinases are important proteins that produce a defense response to pathogens and arthropods that contain chitin, also acting in plant development. Chitinases belong to a large protein family called glycosyl hydrolases (GH) and until now have been widely described in gymnosperms and angiosperms, however, little is reported in more ancient plant species. Plant chitinases are part of the GH18 and GH19 glycosyl hydrolases families. In this study, it was performed searching and annotation of GH18 chitinases in algae, bryophytes, and pteridophytes species, using a set of bioinformatics tools. In total, 648 putative chitinases from the GH18 family were found. The most complete sequences (270 sequences) were selected to additional analyzes. Using multiple sequence alignments, it was possible to observe the high catalytic domain conservation in all analyzed groups. However, some sequences showed substituted amino acids in catalytic residues, which can compromise the chitinolytic function. Phylogenetic analyzes enabled identified chitinases classification in GH18 classes III and V. Structural comparisons of some identified chitinases demonstrated high conservation between three-dimensional chitinase structures in algae, bryophytes and pteridophytes. The results of this study are important to assist in understanding the chitinases relationship between the most primitive plant groups with superior vascular plants, in addition to providing data for future studies related to GH18 chitinases.

Keywords: Chitinases, GH18, plant evolution, bioinformatics, transcriptome.

1 Introduction

The Plantae Kingdom is composed of approximately 370,000 eukaryotic organisms, usually autotrophic and adapted to terrestrial life (Christenhusz and Byng, 2016). This large group includes algae, bryophytes, pteridophytes, gymnosperms and angiosperms (Bennici, 2008). Like terrestrial plants, algae have great ecological importance because they perform photosynthesis and are responsible for much of the oxygen production in the terrestrial environment (Vecchi et al., 2020).

Green algae constitute a diverse group of approximately 8,000 species, being simple single-celled organisms and complex multicellular organisms (Guiry, 2012). Green algae are similar to bryophytes and vascular plants, since both stock starch in the plastids and have chlorophylls *a* and *b* (Nishio, 2000). Green algae group (phylum Chlorophyta) is divided into three main classes: Chlorophyceae, Charophyceae (also known as Streptophyta), and Ulvophyceae (Endymion D Cooper, 2014). Many studies report that Charophyceae algae have highest degree of relationship with bryophytes and vascular plants, due to morphological, biochemical, and genetic similarities (Bennici, 2008). Bryophytes group consists of approximately 25,000 species that show growth regulated by apical cells, poorly differentiated tissues, but do not have conduct vessels specialized in transporting water and nutrients (Frahm et al., 2003; Pittermann, 2010). They are, therefore, small plants, precisely because they do not have vascular tissue. Pteridophytes are seedless vascular plants, with approximately 10,000 species found in different habitats that include rocks, soil, dark and humid environments, predominantly in tropical regions (R M Tryon and Tryon, 1982). These plants have genes that allow them to adapt to different environments, allowing survival in the face of biotic and abiotic challenges.

Chitinases (EC 3.2.1.14) are enzymes that hydrolyze β -1,4 glycosidic bonds between N-acetylglucosamine (GlcNAc) residues that make up the chitin polymer (Martínez-Caballero et al., 2014; Seidl, 2008). Chitin is a linear, insoluble polymer, composed of a long chain of repetitive units of 2-acetamido-2-deoxy-D-glucose, generally also called N-acetyl-D-glucosamine, which are linked by β - (1-4) glycosidic bonds. Depending on their cleavage patterns, which are related to product type formed after substrate hydrolysis, chitinases can be divided into two categories: endo-chitinases and exo-chitinases (M S Brzezinska et al., 2014; Seidl, 2008).

Chitinases are widely found in plants, crustaceans, bacteria, and fungi, which can perform different functions. In plants, chitinases can be located in stems, seeds, turtles, and

flowers (Khan et al., 2015). Plants do not contain chitin in their constituents, however, they have genes capable of producing chitinases in a constitutive or inducible way (Ohnuma et al., 2011). Plants generally express relatively low chitinases levels and only synthesize in higher levels when attacked by phytopathogens. Chitinases are included in a protein group related to pathogenesis called PR proteins (*Related to pathogenesis*) because their expression is induced when in contact with certain pathogens (Neuhaus et al., 1996). Chitinases can induce local or systemic resistance to phytopathogens attack, such as pathogenic fungi, by acting directly on chitin polymer, which is a major component of fungi cell wall (Kesari et al., 2015). Some pathogenesis-related chitinases may also exhibit potential antibacterial, nematicide, and antiviral activities (Chen et al., 2015; Medeiros et al., 2018; Kaur et al., 2019). Plant chitinases can also show a higher expression level when induced by abiotic factors (Cao and Tan, 2019). Certain extreme environmental conditions, such as high salt concentrations, long cold periods and water stress can promote chitinase genes transcription, thus suggesting these proteins may be involved in plants stress response mechanisms (Badariotti et al., 2006; Cao and Tan, 2019). Studies suggest chitinases also act on plant growth and development since chitinase accumulation was observed in the plants transition during flowering and senescence (Edreva, 2005).

Chitinases belong to glycosyl hydrolases (GH) superfamily that hydrolyze glycosidic bonds in carbohydrates (Neuhaus et al., 1996). Based on the similarity of chitinases catalytic domain sequence and in catalysis mechanism, plant chitinases were classified into the GH18 and GH19 glycosyl hydrolases families (Kesari et al., 2015; LOMBARD et al., 2014). Based on evolutionary analysis, it is suggested that chitinases of the GH18 and GH19 families have evolved from different ancestors, as they share low similarity between their amino acid sequences, and have different three-dimensional structures and enzymatic mechanisms (Fukamizo, 2000). Although GH18 chitinases show low similarity with GH19 chitinases, their sequences are similar to those fungi and bacteria chitinases (Kesari et al., 2015).

Plant chitinases were divided into seven classes (I-VII) (Li and Liu, 2003; Neuhaus et al., 1996). Classes III and V belong to the GH18 family, while classes I, II, IV, VI, and VII are members of the GH19 family (Cao and Tan, 2019). Class III is composed of chitinases that do not show similarity with classes I and II, do not show the chitin-binding domain and are generally located in the apoplast (Arie et al., 2000). Class V, on the other hand, comprises a group of chitinases that have two chitin-binding domains and a catalytic domain, of little resemblance to the other classes (El-Katatny et al., 2001).

GH18 chitinases perform glycosidic bonds hydrolysis by retention mechanism of the anomeric configuration, forming products with β conformation (Ken A Brameld et al., 1998; Seidl et al., 2005). They have a catalytic domain (C-terminal) characterized by a barrel-shaped structure consisting of 8 α -helices and 8 β -sheets (Chuang et al., 2008). In addition to the catalytic domain, these enzymes may present chitin-binding domains in the N-terminal region, which are regions that do not perform catalysis, but that enhances chitinase affinity to substrate, which can provide an increase in the efficiency of chitin hydrolysis (Yan and Fong, 2015b).

In this study, we investigated the presence of GH18 family chitinases in transcriptomes of several algae, bryophytes, and pteridophytes species, performed annotation and comparative analysis of these proteins diversity, and explored the phylogenetic relationships and structural characterization. As most chitinase studies are related to superior vascular plants (gymnosperms and angiosperms), the data from this study may be important to help in understanding the relationships of these proteins in the most primitive groups with the most current plants groups.

2 Materials and methods

2.1 Chitinases proteins identification in plant species

Initially, a search for plant chitinases amino acid sequences was performed. Selected sequences were used as references in subsequent analyzes. These sequences were obtained from the CAZy database - Carbohydrate Active Enzymes (<http://www.cazy.org/Glycoside-Hydrolases.html>). Six amino acid GH18 family chitinases sequences (classes III and V) from different plant species were selected. Class III sequences from *Acacia Koa* (AFY08286.1), *Bambusa oldhamii* (ABW75910.1), and *Punica granatum* (PDB: 4TOQ_1) were obtained. From class V, *Arabidopsis thaliana* (PDB: 3AQU_1), *Lotus japonicus* (ANS10044.1), and *Momordica charantia* (AAM18075.1) were obtained.

2.2 Protein database from algae, bryophytes, and pteridophytes

Protein sequences from different algae, bryophytes, and pteridophytes phyla were obtained on the OneKP platform (<https://db.cngb.org/onekp/>) and used to build a database. This platform provides predicted amino acid sequence data from transcriptome of plant species. In addition to green algae phylum, algae species from the phyla Euglenophyta, Dinophyta,

Glaucophyta, Haptophyta, Chrysophyta, Ochrophyta, and Rhodophyta were also analyzed. In this work, 196 families and 397 species provided by the OneKP platform were analyzed (Table 1).

Table 1. Number of families and species analyzed in different phyla of algae, bryophytes, and pteridophytes.

Groups analyzed	Families	Species
Euglenophyta	1	1
Dinophyta	1	1
Glaucophyta	1	4
Haptophyta	3	3
Chrysophyta	4	5
Ochrophyta	11	24
Rhodophyta	16	28
Green Algae	63	156
Bryophytes	58	77
Pteridophytes	38	98
TOTAL	196	397

To identify the putative chitinases present in that bank, an alignment by BLASTp was performed out in the BLAST program using the reference sequences of GH18 plant chitinases.

2.3 *Conserved domains analysis*

Subsequently, a conserved domains analysis was performed on putative chitinases found in algae, bryophytes, and pteridophytes, through alignments by BLASTp and InterProScan using Blast2GO 5.2 program (Conesa et al., 2005). The conserved domains analyzed at this step can be accessed in data bank Pfam (Mistry et al., 2021). For the next stage, only chitinases with the GH18 catalytic domains were analyzed.

2.4 *Chitinase multiple alignments*

Chitinase multiple alignments of each investigated group were performed against reference chitinases. Alignments were performed using MUSCLE method in MEGA 7.0.26 software (Kumar et al., 2016). In this step, conserved domains were also analyzed to verify whether chitinases selected in the previous step contained GH18 chitinase catalytic domain amino acid sequence. Proteins that presented incomplete catalytic domain were discarded from the subsequent analyzes.

2.5 *Phylogenetic Analysis*

Phylogenetic analyzes were performed to classify these proteins into classes III and V of the GH18 family. Phylogenetic trees were built using the MEGA 7.0.26 program (Kumar et al., 2016) using the Neighbor-Joining method (Saitou and Nei, 1987) and Jones-Taylor-Thornton (JTT) model, with phylogeny test by the “bootstrap” method with 500 replicates, (Jones et al., 1992). A phylogenetic tree was elaborated for each analyzed plant group (Supplementary figures). From each group we selected some chitinases sequences and a phylogenetic tree was created to represent the chitinases identified in this study.

2.6 *Modeling of three-dimensional structures*

Three-dimensional structures modeling was performed using the Swiss-Model server (<https://swissmodel.expasy.org/>). For structures prediction, two amino acid sequences were selected from each analyzed plant group, one from class III and the other from class V.

3 **Results and discussion**

Using the data provided by the OneKP platform were identified 648 putative GH18 family chitinases with catalytic domain PFAM-IPR001223 (Table 2). Many of the sequences indicated as chitinases were incomplete or even had large deletions in the conserved domain, and therefore we did not use these to perform alignments and phylogenetic trees. Thus, we selected 270 complete chitinase sequences, which were used for classification and phylogenetic analysis. Similarity and E-value data for the amino acid sequences provided by Blast2GO are available in Supplementary Table S1.

Table 2. Number of chitinases found in each plant group, including families, species, and variation in transcripts number by species.

Groups analyzed	Transcripts found	Families	Species	Variation in transcripts number by specie
Euglenophyta	0	0	0	0
Dinophyta	1	1	1	1
Glaucophyta	1	1	1	1
Haptophyta	0	0	0	0
Chrysophyta	1	1	1	1
Ochrophyta	12	2	2	1 – 8
Rhodophyta	8	4	6	1 – 2

Green Algae	162	31	80	1 – 9
Bryophytes	238	51	70	1 – 13
Pteridophytes	225	16	44	1 – 15
TOTAL	648	107	205	

In algae phyla Euglenophyta and Haptophyta, putative chitinases were not identified. In the phyla Dinophyta, Glaucophyta and Chrysophyta, only one chitinase was identified in each phylum. In Rhodophyta (phylum of red algae) 8 chitinases were found and in the phylum Ochrophyta (brown algae) 12 chitinases were found. However, there are few species of these algae groups in database (Table 1), and this may have influenced the low number of identified chitinases.

162 chitinases were found in green algae, 102 of them in charophytes. Although chitinases were identified in the Chlorophyceae and Ulvophyceae classes, as well as in other smaller classes, the Charophyceae class stood out with the highest chitinases presence. Charophyceae (charophytes) is composed mainly of green freshwater algae and is considered the group that has the highest degree of phylogenetic relationships with bryophytes and vascular plants, due to similarities in photosynthetic pigments (chlorophylls *a* and *b* and beta-carotene), in the energy storage in the form of starch and the cell wall constituted by cellulose (Will. H Blackwell, 2003; Nishio, 2000). *Desmediceae* family, from charophytes group, presented 53 chitinases, being the green algae family with the highest amount of chitinases identified. This family consists of microscopic green algae that are found inhabiting all aquatic freshwater environments (Gontcharov and Melkonian, 2008). Within this family, *Bambusina borrhari* stood out with the presence of 9 chitinases.

It is likely that green algae, as well as terrestrial plants, use chitinases as a form of defense against arthropod attack and pathogenic fungi that contain chitin. There are many fungi that make symbiotic associations with algae (interaction beneficial to both), however, some aquatic pathogenic fungi can cause disease and affect algae survival (Raghukumar, 2006; Rasconi et al., 2011). Chytridiomycete fungi are algae parasites, and have vegetative and reproductive structures constituted of chitinous walls (Rasconi et al., 2011). In addition to being widely present in freshwater regions, there are also many species of fungi inhabiting marine waters and many algae are prevalent in these regions (Amend et al., 2019; Ji and Wang, 2016), this may further reinforce the fact that the presence of chitinases in algae may be related to defense mechanisms against these microorganisms. Aquatic insects also feed on algae, and chitinases can protect against these organisms (Tuno et al., 2018).

Bryophytes are divided into three phyla: Liverworts, Bryophyta and Hornworts (Michelle T Casanova and Nairn, 2016). 238 chitinases were identified in bryophytes, the majority belong to the Liverworts phylum, reaching 125 chitinases. In Hornworts 22 chitinases were found, while in Mosses 87 chitinases were found. In the genus *Marchantia* (Liverworts) the highest chitinases amount was found, reaching 20 chitinases. However, the species *Phaeoceros carolinianus* (Hornworts) presented 15 chitinases, being, therefore, the bryophyte with the highest chitinase amount.

Currently, the pteridophytes group comprises two phyla with existing representatives: Lycophytes and Monilophytes (known as ferns) (Peterson, 1994; Pryer et al., 2001). In total, 225 chitinases were found in pteridophytes. In the Lycophytes phylum 126 chitinases were identified, while in Monilophytes 99 chitinases were identified. *Selaginella* and *Isoetes* genera (Lycophytes) were the ones that presented the highest amount of chitinases, especially *Selaginella wallacei* and *Isoetes tegetiformans* with 15 chitinases each.

In angiosperm plants, *Arabidopsis thaliana* and tomato (*Solanum Lycopersicum*), 10 and 11 GH18 chitinases were identified, respectively (Cao and Tan, 2019; Passarinho and de Vries, 2002). Wang et al. (2015) identified 12 genes from GH18 chitinases in blackberry (*Morus L.*). In *Eucalyptus grandis*, 36 GH18 chitinases genes were identified (Tobias et al., 2017). In this study, the maximum number of chitinases found in a species of green alga is relatively less than the number of chitinases identified in species of bryophytes and pteridophytes. These numbers are also close to the number of chitinases described in angiosperms in the literature.

3.1 *Chitinases conserved domains analysis*

Although the amino acid sequence similarity between class III and V chitinases is relatively low (approximately 15% identity), both have a fundamental catalytic motive for chitinolytic activity, formed by two aspartate residues and one of glutamate (DxDxE), which is conserved in chitinases of various organisms (Ohnuma et al., 2011). Substitutions or deletions of these conserved residues can interfere with chitinolytic function (Tyler et al., 2010). This catalytic motif was also evidenced in class III and V chitinases identified in this study, however, in some sequences these residues are substituted by other amino acids, assuming that some of the identified chitinases may have lost their chitinolytic function.

Enzymes catalytic domains are highly conserved regions and, therefore, these regions were used as a parameter for chitinases selection in analyzed plant species. Chitinase-binding

domains are common to class V chitinases, but it is not present in all classes, because despite being important in the interaction between the chitin polymer and chitinase, catalysis does not have to occur. Chitin-binding domains interact selectively and non-covalently with chitin and can occur in one or more copies, usually having 43 amino acid residues (Lerner and Raikhel, 1992).

Evolutionary studies indicate that chitinases have undergone a rapid evolutionary process. Chitinases acquired evolutionary adaptations, presenting different isoforms, new functions, coming from different organisms (Tyler et al., 2010). Also, several chitinases maintained the chitin bind ability while losing the hydrolysis capacity, which can affect various biological processes (Oyeleye and Normi, 2018). An example is enzymes without conserved catalytic residues, such as nodulins, which are proteins involved in interactions with symbiotic bacteria, as well as narbonins and concanavalin B (seed proteins), which are also grouped within the GH18 family (Tyler et al., 2010).

3.1.1 Class III

Class III enzymes have similarities with bacterial chitinases, as they share the same hydrolysis mechanism and similarity in the three-dimensional structure (Iseli et al., 1996). Class III has a peculiar characteristic, and they are considered bifunctional enzymes, because, in addition to hydrolyzing chitin, they also have lysozyme activity, thus being able to contribute against bacterial infections (Park et al., 2002).

Although certain chitinases have highly conserved domains, they can present different substrate specificities. This fact was observed in class III chitinases in rice, in which two chitinases with highly similar sequences had different expression patterns, where one expressed in response to pathogens attack, while the other had its expression induced in response to environmental stresses (Park et al., 2002). Therefore, small changes in the sequences can infer the GH18 chitinases function. Analysis and comparison of chitinase sequences are very important to assist in predicting these proteins function, as well as in the new chitinases groups discovery.

Although GH18 chitinases are divided into classes III and V, they show low similarity to each other (Ebrahim et al., 2011). Thus, the GH18 chitinase alignments were performed separately for each class, thus making it possible to observe differences in the catalytic domains between these classes (Supplementary figures). In this work, 124 complete class III chitinases

sequences were identified. Among them, we selected six sequences for the representative class III alignment, two of green algae, two of bryophytes, and two pteridophytes sequences (Fig. 1).

It was possible to observe a high similarity between the class III chitinases even with the *Bambusa oldhamii* chitinase, which was used as a reference in the alignment. Even green algae chitinase (considered the terrestrial plants ancestral group) has shown its sequence very similar to chitinases identified in bryophytes and pteridophytes. In class III, in green algae and mainly in bryophytes, some sequences with the catalytic motif DxDxE were observed, presenting residues replaced by other amino acids. However, in pteridophytes this important region for chitinolytic activity is highly conserved in all analyzed species.

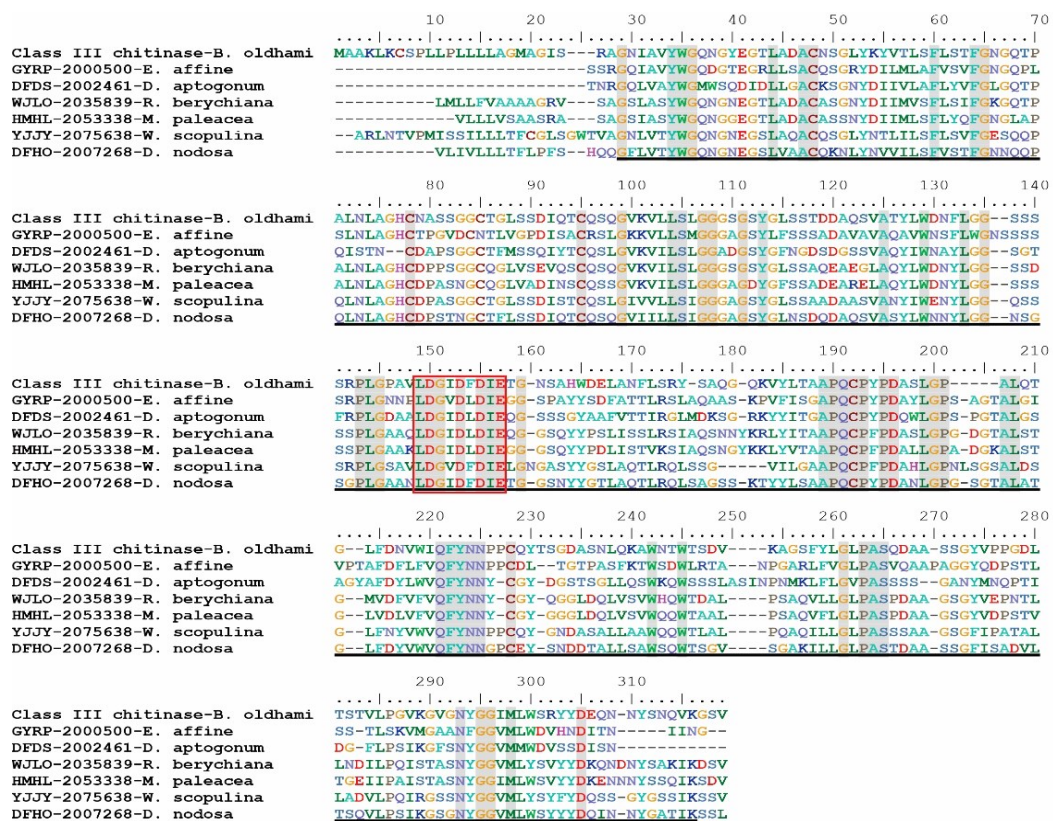


Fig. 1. Class III chitinase multiple alignments of sequences identified in green algae, bryophytes, and pteridophytes. Shaded regions indicate conserved residues in the species. Region marked by a red rectangle indicates the conserved DxDxE catalytic motif that is fundamental for chitinolytic activity. Regions highlighted in black at the bottom of the alignment indicate the conserved catalytic domains. The identification code GYRP-2000500 indicates a green algae chitinase from *Euastrum affine* and DFDS-2002461 indicates a green algae chitinase from *Desmidium aptogonum*. Codes WJLO-2035839 and HMHL-2053338 indicate chitinases of bryophytes *Riccia berychiana* and *Marchantia paleacea*, respectively. Codes YJYY-2075638 and DFHO-2007268 indicate chitinases identified in pteridophytes *Woodсия scopulina* and *Danaea nodosa*, respectively.

3.1.2 Class V

Ohnuma et al. (2011) observed in *Arabidopsis thaliana* the presence of a conserved motif in the catalytic class V domain (constituted, consecutively, by the amino acids aspartate,

leucine, aspartate, tryptophan, and glutamate - DLDWE). This region proved to be important in the process of catalyzing these enzymes and was also observed in class V chitinases of the GH18 family analyzed in this study (Fig. 2). Class V chitinases identified in this study also demonstrated high sequence similarity. In total, were found 140 complete sequences from GH18 class V chitinases. Of these, we selected six sequences to elaborate a representative alignment, two of green algae, two of bryophytes, and two sequences of pteridophytes (Fig. 2). In class V, only one green algae sequence presented the catalytic motif DxDxE with residues replaced by other amino acids. In the other analyzed algal phyla (Rhodophyta and Ochrophyta), in bryophytes and pteridophytes, this region proved to be highly conserved in all analyzed species.

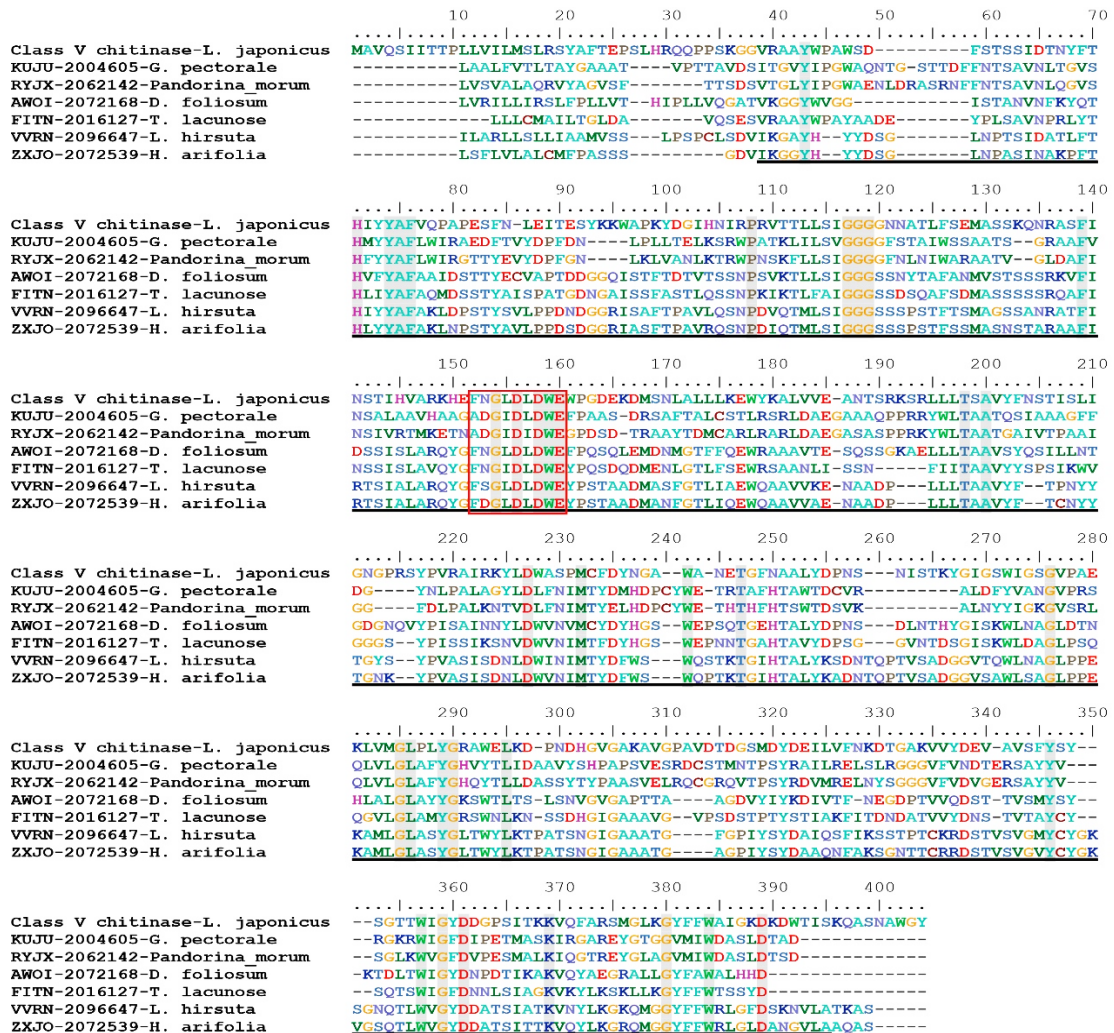


Fig. 2. Class V chitinases multiple alignments of sequences identified in green algae, bryophytes, and pteridophytes. Shaded regions indicate conserved residues. Region marked by a red rectangle indicates the conserved DxDxE catalytic motif that is fundamental for chitinolytic activity. Regions highlighted in black at the bottom of the alignment indicate the conserved catalytic domains. Codes KUJU-2004605 and RYJX-206214 represent green algae chitinases from *Gonium pectorale* and *Pandorina morum*. Codes AWOI-2072168 and FITN-2016127 indicate bryophytes chitinases from *Diphyscium foliosum* and *Treubia lacunose*, respectively. While in pteridophytes code VVRN-2096647 indicates chitinase from *Lonchitis hirsuta* and ZXJO-2072539 indicates *Hemionitis arifolia* chitinase.

3.2 *Phylogenetic relationships and classification of identified chitinases*

270 algae, bryophytes and pteridophytes chitinases sequences were evaluated by phylogenetic analysis. To summarize this result, a phylogenetic tree was made up with 32 complete chitinase sequences, together with the six reference sequences (Fig. 3). Phylogenetic trees with complete sequences allows that all the information contained in the protein sequences can contribute to the phylogenetic positioning (Tyler et al., 2010). Class III chitinases from all groups analyzed were grouped in a different clade from Class V, precisely because they share low sequence similarity. In Fig. 3, it is noted that most sequences were grouped with phylogenetic relationships of the different plant clades (algae, green algae, bryophytes, and pteridophytes). Phylogenetic trees with all the complete sequences identified in algae, green algae, bryophytes, and pteridophytes used for classification are available in Supplementary figures.

We identified three complete chitinases in algae from different phyla, two chitinases in *Sargassum thunbergii* (phylum Ochrophyta), and one chitinase in *Symphyocladia latiuscula* (phylum Rhodophyta). These chitinases were grouped in class V and none were grouped in class III. In the analyzed Chrysophyta, Dinophyta, and Glaucophyta phyla, complete chitinases were not found. This shows that chitinase genes are not present or are not expressed in most primitive algal species, this question would be answered only with complete genomes sequencing of these species.

In the green algae phylum, a much larger chitinases number from GH18 family was identified when compared with the other analyzed algae phyla, with a total of 81 complete chitinases. Through phylogenetic analysis, it was possible to verify that, from these proteins identified in green algae, 34 were grouped in the class III chitinases, while 47 are class V members.

In the phylogenetic analysis of different bryophytes species were found 99 complete chitinases of the GH18 family, greater than the chitinases number identified in green algae. It was possible to observe 42 chitinases were grouped in the class III chitinase branch and 57 were class V chitinases in phylogenetic analysis. In pteridophytes, 87 complete chitinases were identified, 48 were grouped in the class III chitinases and 39 are class V members.

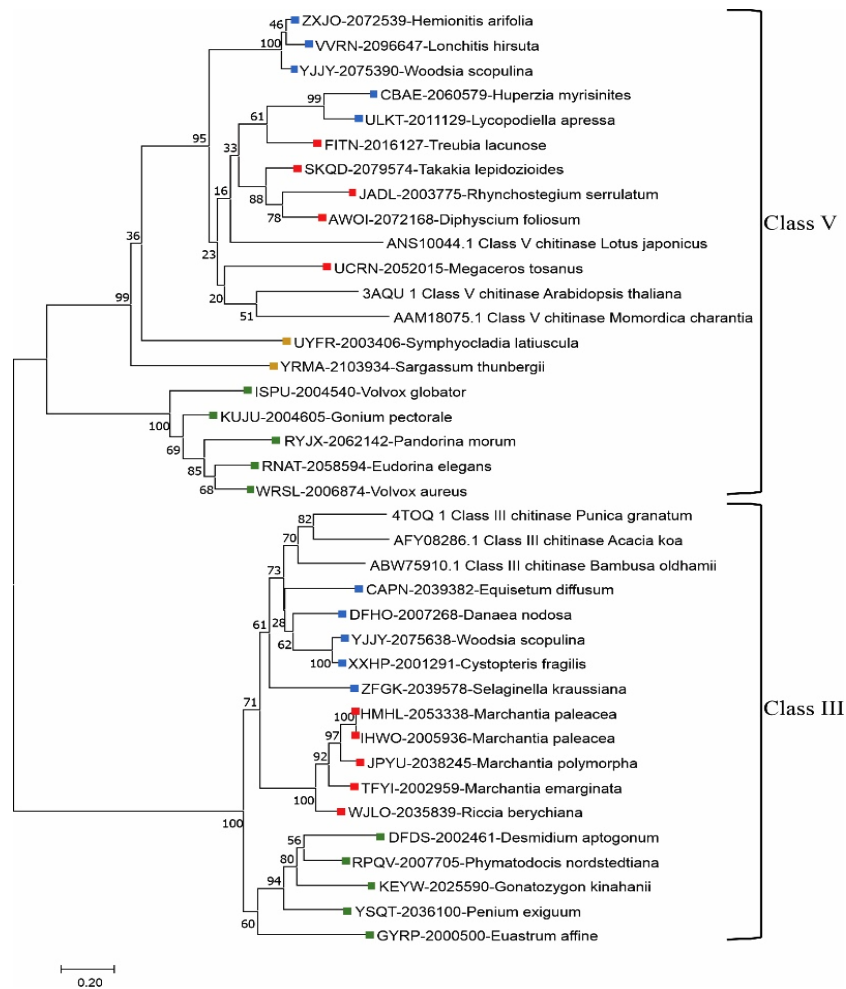


Fig. 3. Phylogenetic relationships of class III and V GH18 family chitinases identified in algae, bryophytes, and pteridophytes. 32 chitinases sequences were selected from the groups analyzed for the development of the phylogenetic tree. Blue squares represent chitinases identified in bryophytes, red squares indicate chitinases identified in pteridophytes, yellow squares indicate chitinases identified in algae while the green squares indicate chitinases in green algae. Angiosperm reference sequences are not indicated by squares.

3.3 Structural characterization

Catalytic domains showed a high structure similarity in all analyzed groups (Fig. 4 and 5). According to Fukamizo (2000), structural differences may be associated with the functional chitinases diversity, therefore related to the different physiological functions that these enzymes can perform. Thus, three-dimensional structures can provide important information for chitinases catalytic mechanisms studies, since the enzyme plays its role when the arrangement of the three-dimensional structure occurs properly.

Comparing class III with class V, it is possible to notice a low similarity in the 3D structures analyzed, precisely because classes III and V consist of catalytic domains with low similarity to each other (also, the catalytic domain comprises a large part of the protein chitinase). In both class III and class V structures, it is possible to observe the α -helices are

located in the outer region, involving the β - sheets, leading to the barrel shape. Analyzing the class III three-dimensional structures of green algae (Fig. 4A), bryophytes (Fig. 4B) and pteridophytes (Fig. 4C), we observed that these structures are high similarity to each other.

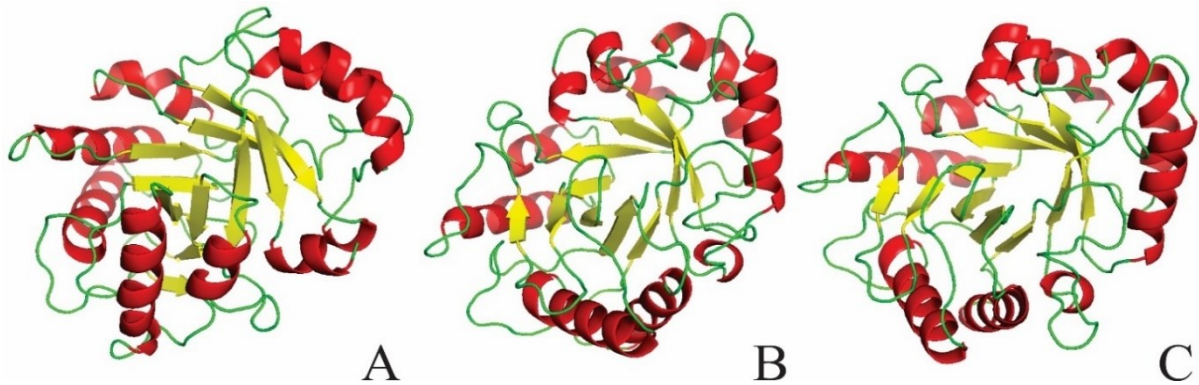


Fig. 4. Structural comparisons of class III GH18 family chitinases. (A) Three-dimensional structure of green algae chitinase *Desmidium aptogonum* DFDS-2002461. (B) Three-dimensional structure of bryophyte chitinase *Riccia berychiana* WJLO-2035839. (C) Three-dimensional structure of pteridophyte chitinase *Woodsia scopulina* (YJJY-2075638). Structures were modeled using the Swiss-Model server, highlighting the yellow β - sheets and red α - helices.

In class V (Fig. 5), three-dimensional structures of green algae (Fig. 5A) bryophytes (Fig. 5B) and pteridophytes (Fig. 5C) also are highly similar to each other. The arrangements of α -helices and β -sheets proved to be similar in bryophytes and pteridophytes class V chitinases.

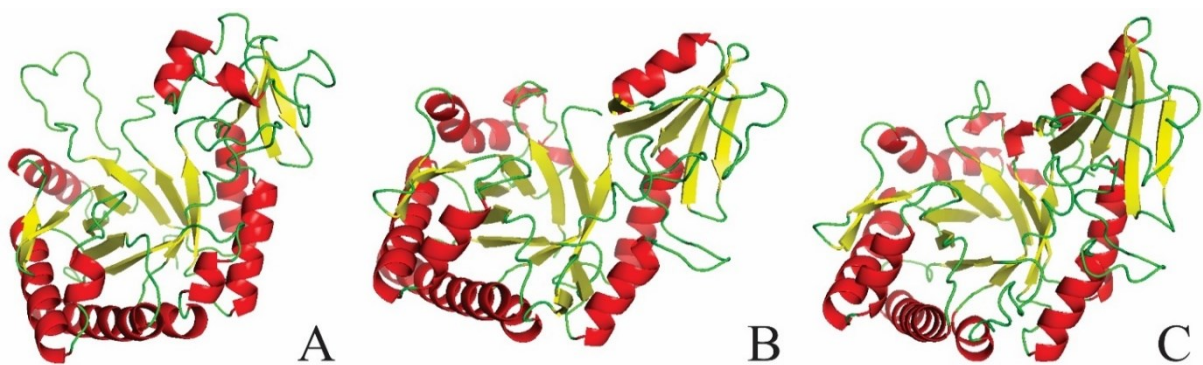


Fig. 5. Structural comparisons of class V GH18 family chitinases. (A) Three-dimensional structure of green algae chitinase *Gonium pectorale* KUJU-2004605. (B) Three-dimensional structure of bryophyte chitinase *Diphyscium foliosum* AWOI-2072168. (C) Three-dimensional structure of pteridophyte chitinase *Lonchitis hirsuta* VVRN-2096647. Structures were modeled using the Swiss-Model server, highlighting the yellow β - sheets and red α - helices.

4 Conclusions

In this study, we investigated chitinase presence in the most ancient terrestrial plants groups, which are algae, bryophytes, and pteridophytes, since most studies of chitinases are reported in higher vascular plants. A large chitinases number from the GH18 family were found in green algae, bryophytes, and pteridophytes species. However, only three chitinases were identified in two algae phyla, Rhodophyta and Ochrophyta. In the green algae, bryophytes and pteridophytes, a high number of chitinases distributed in different species was identified. Chitinases with highly conserved catalytic domains were identified in all groups analyzed, but some sequences showed low similarity and even changes in the catalytic residues essential for catalysis. Phylogenetic tree separated class III from class V because despite being part of the same family, these classes have low similarity to each other. Three-dimensional structures analyzed of green algae, bryophytes and pteridophytes showed high similarity to each other. Chitinases identification and characterization are important to understand the relationships of these proteins with current plant groups since studies suggest these proteins have undergone a rapid evolutionary process, leading to new functions emergence or even the loss of chitinolytic function. The results of this study can also serve to provide data for future studies related to GH18 family chitinases.

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References

- Amend, A., Burgaud, G., Cunliffe, M., Edgcomb, V.P., Ettinger, C.L., Gutiérrez, M.H., Heitman, J., Hom, E.F.Y., Ianiri, G., Jones, A.C., Kagami, M., Picard, K.T., Quandt, C.A., Raghukumar, S., Riquelme, M., Stajich, J., Vargas-Muñiz, J., Walker, A.K., Yarden, O., Gladfelter, A.S., 2019. Fungi in the marine environment: Open questions and unsolved problems. *MBio* 10, 1–15. <https://doi.org/10.1128/mBio.01189-18>
- Arie, M., Hikichi, K., Takahashi, K., Esaka, M., 2000. Characterization of a basic chitinase which is secreted by cultured pumpkin cells. *Physiol. Plant.* 110, 232–239. <https://doi.org/10.1034/j.1399-3054.2000.110213.x>
- Badariotti, F., Kyriotou, M., Lelong, C., Dubos, M.P., Renard, E., Galera, P., Favrel, P., 2006. The phylogenetically conserved molluscan chitinase-like protein 1 (Cg-Clp1), homologue of human HC-gp39, stimulates proliferation and regulates synthesis of extracellular matrix components of mammalian chondrocytes. *J. Biol. Chem.* 281, 29583–29596. <https://doi.org/10.1074/jbc.M605687200>
- Bennici, A., 2008. Origin and early evolution of land plants. *Commun. Integr. Biol.* 1, 212–218. <https://doi.org/10.4161/cib.1.2.6987>
- Blackwell, Will. H, 2003. Two Theories of Origin of the Land-Plant Sporophyte: Which Is Left Sanding? *Bot. Rev.* 69, 125–148. [https://doi.org/https://doi.org/10.1663/0006-8101\(2003\)069\[0125:TTOOOT\]2.0.CO;2](https://doi.org/https://doi.org/10.1663/0006-8101(2003)069[0125:TTOOOT]2.0.CO;2)
- Brameld, Ken A., Shrader, W.D., Imperiali, B., Goddard, W.A., 1998. Substrate assistance in the mechanism of family 18 chitinases: Theoretical studies of potential intermediates and inhibitors. *J. Mol. Biol.* 280, 913–923. <https://doi.org/10.1006/jmbi.1998.1890>
- Brzezinska, M. S., Jankiewicz, U., Burkowska, A., Walczak, M., 2014. Chitinolytic microorganisms and their possible application in environmental protection. *Curr. Microbiol.* 68, 71–81. <https://doi.org/10.1007/s00284-013-0440-4>
- Cao, J., Tan, X., 2019. Comprehensive analysis of the chitinase family genes in tomato (*Solanum Lycopersicum*). *Plants* 8. <https://doi.org/10.3390/plants8030052>
- Casanova, Michelle T., Nairn, L.C., 2016. Macroalgae, charophytes and bryophytes. *Veg. Aust. Riverine Landscapes*.
- Chen, L., Jiang, H., Cheng, Q., Chen, J., Wu, G., Kumar, A., Sun, M., Liu, Z., 2015. Enhanced nematicidal potential of the chitinase pachi from *Pseudomonas aeruginosa* in association with Cry21Aa. *Sci. Rep.* 5, 1–11. <https://doi.org/10.1038/srep14395>
- Christenhusz, M.J.M., Byng, J.W., 2016. *Phytotaxa*. *Phytotaxa* 261, 201–217.

- <https://doi.org/10.11646/phytotaxa.261.3.1>
- Chuang, H.H., Lin, H.Y., Lin, F.P., 2008. Biochemical characteristics of C-terminal region of recombinant chitinase from *Bacillus licheniformis*- implication of necessity for enzyme properties. *FEBS J.* 275, 2240–2254. <https://doi.org/10.1111/j.1742-4658.2008.06376.x>
- Conesa, A., Götz, S., García-Gómez, J.M., Terol, J., Talón, M., Robles, M., 2005. Blast2GO: A universal tool for annotation, visualization and analysis in functional genomics research. *Bioinformatics* 21, 3674–3676. <https://doi.org/10.1093/bioinformatics/bti610>
- Cooper, Endymion D., 2014. Overly simplistic substitution models obscure green plant phylogeny. *Trends Plant Sci.* 19, 576–582. <https://doi.org/10.1016/j.tplants.2014.06.006>
- Medeiros, S.C., Monteiro-Júnior, J.E., Passos Sales, G.W., Grangeiro, T.B., Pinto Nogueira, N.A., 2018. Chitinases as antibacterial proteins: A systematic review. *J. Young Pharm.* 10, 144–148. <https://doi.org/10.5530/jyp.2018.10.33>
- Ebrahim, S., Usha, K., Singh, B., 2011. Pathogenesis-related (PR)-proteins: Chitinase and β -1,3-glucanase in defense mechanism against malformation in mango (*Mangifera indica* L.). *Sci. Hortic. (Amsterdam)*. 130, 847–852. <https://doi.org/10.1016/j.scienta.2011.09.014>
- Edreva, A., 2005. Pathogenesis-related proteins: research progress in the last 15 years. *Gen Appl Plant Physiol* 31, 105–24.
- El-Katatny, M.H., Gudelj, M., Robra, K.H., Elnaghy, M.A., Gübitz, G.M., 2001. Characterization of a chitinase and an endo- β -1,3-glucanase from *Trichoderma harzianum* Rifai T24 involved in control of the phytopathogen *Sclerotium rolfsii*. *Appl. Microbiol. Biotechnol.* 56, 137–143. <https://doi.org/10.1007/s002530100646>
- Frahm, J.P., Pócs, T., O’Shea, B., Koponen, T., Piipo, S., Enroth, J., Rao, P., Fang, Y., 2003. Manual of tropical bryology. *Trop. Bryol.* 23, 1–196. <https://doi.org/10.11646/bde.23.1.1>
- Fukamizo, T., 2000. Chitinolytic Enzymes: Catalysis, Substrate Binding, and their Application. *Curr. Protein Pept. Sci.* 1, 105–124. <https://doi.org/10.2174/1389203003381450>
- Gontcharov, A.A., Melkonian, M., 2008. In search of monophyletic taxa in the family Desmidiaceae (Zygnematophyceae, Viridiplantae): The genus *Cosmarium*. *Am. J. Bot.* 95, 1079–1095. <https://doi.org/10.3732/ajb.0800046>
- Guiry, M.D., 2012. How many species of algae are there? *J. Phycol.* 48, 1057–1063. <https://doi.org/10.1111/j.1529-8817.2012.01222.x>
- Hamid, R., Khan, M., Ahmed, M., Ahmad, M.M., Abdin, M.Z., Musarrat, J., Javed, S., 2013. Chitinases: An update. *J Pharm Bioallied Sci* 5, 21–29. <https://doi.org/10.4103/0975->

7406.106559

- Iseli, B., Armand, S., Boller, T., Neuhaus, J.M., Henrissat, B., 1996. Plant chitinases use two different hydrolytic mechanisms. *FEBS Lett.* 382, 186–188.
[https://doi.org/10.1016/0014-5793\(96\)00174-3](https://doi.org/10.1016/0014-5793(96)00174-3)
- Ji, N.Y., Wang, B.G., 2016. Mycochemistry of marine algicolous fungi. *Fungal Divers.* 80, 301–342. <https://doi.org/10.1007/s13225-016-0358-9>
- Jones, D.T., Taylor, W.R., Thornton, J.M., 1992. The rapid generation of mutation data matrices from protein sequences. *Bioinformatics* 8, 275–282.
<https://doi.org/https://doi.org/10.1093/bioinformatics/8.3.275>
- Kaur, R., Neetu, Mudgal, R., Jose, J., Kumar, P., Tomar, S., 2019. Glycan-dependent chikungunya viral infection divulged by antiviral activity of NAG specific chi-like lectin. *Virology* 526, 91–98. <https://doi.org/10.1016/j.virol.2018.10.009>
- Kesari, P., Patil, D.N., Kumar, Pramod, Tomar, S., Sharma, A.K., Kumar, Pravindra, 2015. Structural and functional evolution of chitinase-like proteins from plants. *Proteomics* 15, 1693–1705. <https://doi.org/10.1002/pmic.201400421>
- Khan, F.I., Bisetty, K., Singh, S., Permaul, K., Hassan, M.I., 2015. Chitinase from *Thermomyces lanuginosus* SSBP and its biotechnological applications. *Extremophiles* 19, 1055–1066. <https://doi.org/10.1007/s00792-015-0792-8>
- Kumar, S., Stecher, G., Tamura, K., 2016. MEGA7: Molecular Evolutionary Genetics Analysis Version 7.0 for Bigger Datasets. *Mol. Biol. Evol.* 33, 1870–1874.
<https://doi.org/10.1093/molbev/msw054>
- Lerner, D.R., Raikhel, N. V, 1992. The gene for stinging nettle lectin (*Urtica dioica* agglutinin) encodes both a lectin and a chitinase. *J. Biol. Chem.* 267, 11085–11091.
- Li, J., Liu, J., 2003. A Novel cotton gene encoding a new class of chitinase. *Acta Bot. Sin.* 45, 1489–1496.
- Martínez-Caballero, S., Cano-Sánchez, P., Mares-Mejía, I., Díaz-Sánchez, A.G., Macías-Rubalcava, M.L., Hermoso, J.A., Rodríguez-Romero, A., 2014a. Comparative study of two GH19 chitinase-like proteins from *Hevea brasiliensis*, one exhibiting a novel carbohydrate-binding domain. *FEBS J.* 281, 4535–4554.
<https://doi.org/10.1111/febs.12962>
- Mistry, J., Chuguransky, S., Williams, L., Qureshi, M., Salazar, G.A., Sonnhammer, E.L.L., Tosatto, S.C.E., Paladin, L., Raj, S., Richardson, L.J., Finn, R.D., Bateman, A., 2021. Pfam: The protein families database in 2021. *Nucleic Acids Res.* 49, D412–D419.
<https://doi.org/10.1093/nar/gkaa913>

- Neuhaus, J.-M., Fritig, B., Linthorst, H.J., Meins, J., Mikkelsen, J., Ryals, J., 1996. A Revised Nomenclature for Chitinase Genes. *Plant Mol. Biol. Report.* 14, 102–104. <https://doi.org/https://doi.org/10.1007/BF02684897>
- Nishio, J.N., 2000. Why are higher plants green? Evolution of the higher plant photosynthetic pigment complement. *Plant, Cell Environ.* 23, 539–548. <https://doi.org/10.1046/j.1365-3040.2000.00563.x>
- Ohnuma, T., Numata, T., Osawa, T., Mizuhara, M., Lampela, O., Juffer, A.H., Skriver, K., Fukamizo, T., 2011. A class V chitinase from *Arabidopsis thaliana*: Gene responses, enzymatic properties, and crystallographic analysis. *Planta* 234, 123–137. <https://doi.org/10.1007/s00425-011-1390-3>
- Oyeleye, A., Normi, Y.M., 2018. Chitinase: Diversity, limitations, and trends in Engineering for suitable applications. *Biosci. Rep.* 38, 1–21. <https://doi.org/10.1042/BSR20180323>
- Park, S.M., Kim, D.H., Truong, N.H., Itoh, Y., 2002. Heterologous expression and characterization of class III chitinases from rice (*Oryza sativa* L.). *Enzyme Microb. Technol.* 30, 697–702. [https://doi.org/10.1016/S0141-0229\(02\)00042-X](https://doi.org/10.1016/S0141-0229(02)00042-X)
- Passarinho, P.A., de Vries, S.C., 2002. *Arabidopsis* Chitinases : a Genomic Survey . *Arab. B.* 1, e0023. <https://doi.org/10.1199/tab.0023>
- Peterson, K.J., 1994. The Origin and Early Evolution of the Craniata. *Short Courses Paleontol.* 7, 14–37. <https://doi.org/10.1017/s2475263000001240>
- Pittermann, J., 2010. The evolution of water transport in plants: An integrated approach. *Geobiology* 8, 112–139. <https://doi.org/10.1111/j.1472-4669.2010.00232.x>
- Pryer, K.M., Schneider, H., Smith, A.R., Cranfill, R., Wolf, P.G., Hunt, J.S., Sipes, S.D., 2001. Horsetails and ferns are a monophyletic group and the closest living relatives to seed plants. *Nature* 409, 618–621. <https://doi.org/10.1038/35054555>
- Raghukumar, C., 2006. Algal-fungal interactions in the marine ecosystem: Symbiosis to parasitism. *Recent Adv. Appl. Asp. Indian Mar. Algae with Ref. to Glob. Scenar.* 1, 366–385.
- Rasconi, S., Jobard, M., Sime-Ngando, T., 2011. Parasitic fungi of phytoplankton: Ecological roles and implications for microbial food webs. *Aquat. Microb. Ecol.* 62, 123–137. <https://doi.org/10.3354/ame01448>
- Saitou, N., Nei, M., 1987. The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.* 4, 406–425. <https://doi.org/10.1093/oxfordjournals.molbev.a040454>
- Seidl, V., 2008. Chitinases of filamentous fungi: a large group of diverse proteins with

- multiple physiological functions. *Fungal Biol. Rev.* 22, 36–42.
<https://doi.org/10.1016/j.fbr.2008.03.002>
- Seidl, V., Huemer, B., Seiboth, B., Kubicek, C.P., 2005. A complete survey of *Trichoderma* chitinases reveals three distinct subgroups of family 18 chitinases. *FEBS J.* 272, 5923–5939. <https://doi.org/10.1111/j.1742-4658.2005.04994.x>
- Tobias, P.A., Christie, N., Naidoo, S., Guest, D.I., Külheim, C., 2017. Identification of the *Eucalyptus grandis* chitinase gene family and expression characterization under different biotic stress challenges. *Tree Physiol.* 37, 565–582.
<https://doi.org/10.1093/treephys/tpx010>
- Tryon, R. M., Tryon, A.F., 1982. *Ferns and allied Plants with Special reference to Tropical America*. Springer-Verlag, New York. <https://doi.org/10.2307/1546858>
- Tuno, N., Kohzu, A., Tayasu, I., Nakayama, T., Githeko, A., Yan, G., 2018. An Algal Diet Accelerates Larval Growth of *Anopheles gambiae* (Diptera: Culicidae) and *Anopheles arabiensis* (Diptera: Culicidae). *J. Med. Entomol.* <https://doi.org/10.1093/jme/tjx244>
- Tyler, L., Bragg, J.N., Wu, J., Yang, X., Tuskan, G.A., Vogel, J.P., 2010. Annotation and comparative analysis of the glycoside hydrolase genes in *Brachypodium distachyon*. *BMC Genomics* 11, 600. <https://doi.org/10.1186/1471-2164-11-600>
- Vecchi, V., Barera, S., Bassi, R., Dall'osto, L., 2020. Potential and challenges of improving photosynthesis in algae. *Plants* 9. <https://doi.org/10.3390/plants9010067>
- Wang, X., He, N., Zeng, Q., Xiang, Z., 2015. Identification and expression analyses of chitinase genes in mulberry (*Morus L.*) plants. *Plant Omics* 8, 183–189.
- Yan, Q., Fong, S.S., 2015. Bacterial chitinase: Nature and perspectives for sustainable bioproduction. *Bioresour. Bioprocess.* 2. <https://doi.org/10.1186/s40643-015-0057-5>

Supplementary material

Table S1. Blast2GO results of algae phyla, bryophytes and pteridophytes putative GH18 chitinases

ALGAE – GH18						
SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-EBWI-2035542-Ochromonas sp	Chrysophyta	Chromulinaceae	glycoside hydrolase	1,59292E-18	71.42	IPR001223 (PFAM)
scaffold-IEHF-2003601-Dumontia simplex	Rodophyta	Dumontiaceae	Chitinase	5,62296E-74	58.65	IPR001223 (PFAM)
scaffold-IKIZ-2012657-Grateloupia livida	Rodophyta	Halymeniaceae	Chitinase	9,18487E-100	73.3	IPR001223 (PFAM)
scaffold-JEBK-2004603-Eucheuma denticulatum	Rodophyta	Solieriaceae	glycoside hydrolase family 18 protein	1,61585E-78	72.99	IPR001223 (PFAM)
scaffold-POOW-2046963-Glaucocystis cf. nostochinearum	Glaucophyta	Glaucozystaceae	chitotriosidase-1 precursor	1,80205E-27	68.94	IPR001223 (PFAM)
scaffold-PYDB-2002012-Sinotubimorpha guangdongensis	Rodophyta	Halymeniaceae	probable chitinase 10	1,34808E-42	72.51	IPR001223 (PFAM)
scaffold-PYDB-2020269-Sinotubimorpha guangdongensis	Rodophyta	Halymeniaceae	acidic mammalian chitinase-like	7,6859E-46	83.85	IPR001223 (PFAM)
scaffold-RAPY-2001279-Kjellmaniella crassifolia	Ochrophyta	Laminariaceae	chitinase 4	6,70733E-48	80.81	IPR001223 (PFAM)
scaffold-RAPY-2073914-Kjellmaniella crassifolia	Ochrophyta	Laminariaceae	chitinase 4 precursor	5,90828E-32	71.96	IPR001223 (PFAM)
scaffold-RAPY-2075238-Kjellmaniella crassifolia	Ochrophyta	Laminariaceae	acidic mammalian chitinase-like	1,67502E-73	66.99	IPR001223 (PFAM)
scaffold-RAPY-2078410-Kjellmaniella crassifolia	Ochrophyta	Laminariaceae	Chitinase	1,24398E-42	65.31	IPR001223 (PFAM)
scaffold-TZJQ-2121870-Prorocentrum micans	Dinophyta	Prorocentraceae	Acidic mammalian chitinase	5,74682E-73	57.66	IPR001223 (PFAM)
scaffold-UYFR-2003406-Symphyocladia latiuscula	Rodophyta	Rhodomelaceae	acidic mammalian chitinase-like	8,55986E-137	71.38	IPR001223 (PFAM)
scaffold-UYFR-2004651-Symphyocladia latiuscula	Rodophyta	Rhodomelaceae	chitotriosidase-1	5,84526E-47	75.74	IPR001223 (PFAM)
scaffold-YRMA-2002440-Sargassum thunbergii	Ochrophyta	Sargassaceae	AChain A	1,77261E-38	72.75	IPR001223 (PFAM)
scaffold-YRMA-2005642-Sargassum thunbergii	Ochrophyta	Sargassaceae	acidic mammalian chitinase-like	4,17761E-16	77.42	IPR001223 (PFAM)
scaffold-YRMA-2017852-Sargassum thunbergii	Ochrophyta	Sargassaceae	chitinase-3-like protein 1	2,84134E-165	75.78	IPR001223 (PFAM)
scaffold-YRMA-2097681-Sargassum thunbergii	Ochrophyta	Sargassaceae	probable chitinase 10	1,19293E-53	81.17	IPR001223 (PFAM)
scaffold-YRMA-2102570-Sargassum thunbergii	Ochrophyta	Sargassaceae	chitinase-3-like protein 1	3,97527E-56	59.96	IPR001223 (PFAM)
scaffold-YRMA-2103020-Sargassum thunbergii	Ochrophyta	Sargassaceae	AChain A	4,37781E-68	64.43	IPR001223 (PFAM)
scaffold-YRMA-2103934-Sargassum thunbergii	Ochrophyta	Sargassaceae	chitotriosidase-1 precursor	2,11977E-109	62.96	IPR001223 (PFAM)
scaffold-YRMA-2105318-Sargassum thunbergii	Ochrophyta	Sargassaceae	acidic mammalian chitinase isoform x2	2,49986E-76	57.43	IPR001223 (PFAM)
scaffold-ZJOJ-2054710-Grateloupia filicina	Rodophyta	Halymeniaceae	Chitinase	1,63683E-101	73.14	IPR001223 (PFAM)

GREEN ALGAE – GH18					
SeqName	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-ACRY-2005832-Pteromonas sp	Phacotaceae	acidic mammalian chitinase-like	2,8735E-30	58.2	IPR001223 (PFAM)
scaffold-AEKF-2009302-Penium margaritaceum	Peniaceae	class V chitinase	1,05668E-43	54.91	IPR001223 (PFAM)
scaffold-AEKF-2044086-Penium margaritaceum	Peniaceae	chitinase-like protein 4	3,20758E-10	59.12	IPR001223 (PFAM)
scaffold-BAZF-2007777-Chaetopeltis orbicularis	Chaetopeltidaceae	Chitinase	1,5718E-64	54.95	IPR001223 (PFAM)
scaffold-BAZF-2016149-Chaetopeltis orbicularis	Chaetopeltidaceae	chitotriosidase-1	2,15302E-59	5574	IPR001223 (PFAM)
scaffold-BFIK-2029987-Entransia fimbriat	Klebsormidiaceae	acidic mammalian chitinase-like	1,18834E-39	52.89	IPR001223 (PFAM)
scaffold-BHBK-2048640-Cosmarium tinctum	Desmidiaceae	brain chitinase and chia	9,84079E-102	52.91	IPR001223 (PFAM)
scaffold-DFDS-2000769-Desmidium aptogonum	Desmidiaceae	hevamine-A-like	2,44497E-40	69.4	IPR001223 (PFAM)
scaffold-DFDS-2002461-Desmidium aptogonum	Desmidiaceae	acidic endochitinase	1,09525E-62	61.1	IPR001223 (PFAM)
scaffold-DFDS-2002462-Desmidium aptogonum	Desmidiaceae	hevamine-A-like	4,3324E-57	62.79	IPR001223 (PFAM)
scaffold-DRFX-2014366-Closterium lunula	Closteriaceae	acidic mammalian chitinase-like	8,84728E-36	56.07	IPR001223 (PFAM)
scaffold-DZPJ-2054002-Cylindrocapsa geminella	Mesotaeniaceae	Chitinase	2,88943E-86	55.57	IPR001223 (PFAM)
scaffold-EEJO-2039453-Neochloris oleoabundans	incertae sedis	brain chitinase and chia	5,77463E-142	58.81	IPR001223 (PFAM)
scaffold-ETGN-2057676-Botryococcus braunii	Botryococcaceae	Glycoside hydrolase	4,01752E-85	88.76	IPR001223 (PFAM)
scaffold-FFGR-2004388-Netrium digitus	Mesotaeniaceae	acidic endochitinase	1,59149E-41	63.63	IPR001223 (PFAM)
scaffold-FMVB-2005958-Scherffelia dubia	Chlorodendraceae	Chitinase	3,79911E-59	70.48	IPR001223 (PFAM)
scaffold-FOYQ-2002254-Microspora cf. tumidula	Microsporaceae	glycoside hydrolase	6,12419E-37	52.66	IPR001223 (PFAM)
scaffold-FXHG-2079786-Hafniomonas reticulata	Dunaliellaceae	chitinase	9,13289E-47	48.39	IPR001223 (PFAM)
scaffold-GBGT-2000381-Xanthidium antilopaeum	Desmidiaceae	hevamine-A-like	1,44545E-20	67.86	IPR001223 (PFAM)
scaffold-GBGT-2002008-Xanthidium antilopaeum	Desmidiaceae	acidic mammalian chitinase-like	2,64665E-18	61.55	IPR001223 (PFAM)
scaffold-GBGT-2002009-Xanthidium antilopaeum	Desmidiaceae	acidic mammalian chitinase-like	2,64665E-18	61.55	IPR001223 (PFAM)
scaffold-GBGT-2024109-Xanthidium antilopaeum	Desmidiaceae	acidic mammalian chitinase-like	1,86839E-31	56.96	IPR001223 (PFAM)
scaffold-GBGT-2103934-Xanthidium antilopaeum	Desmidiaceae	class V chitinase-like	1,57782E-27	60.2	IPR001223 (PFAM)
scaffold-GFUR-2016925-Chloromonas subdivisa	Chlamydomonadaceae	acidic mammalian chitinase	1,33954E-68	49.85	IPR001223 (PFAM)
scaffold-GGWH-2009903-Onychonema laeve	Desmidiaceae	Acidic endochitinase	3,07289E-76	63.74	IPR001223 (PFAM)
scaffold-GGWH-2048506-Onychonema laeve	Desmidiaceae	acidic endochitinase-like	2,64921E-76	64.83	IPR001223 (PFAM)
scaffold-GYRP-2000499-Euastrum affine	Desmidiaceae	acidic endochitinase-like	1,22857E-63	63.15	IPR001223 (PFAM)
scaffold-GYRP-2000500-Euastrum affine	Desmidiaceae	acidic endochitinase-like	7,68057E-65	63.71	IPR001223 (PFAM)
scaffold-GYRP-2000501-Euastrum affine	Desmidiaceae	acidic endochitinase-like	1,22857E-63	63.15	IPR001223 (PFAM)
scaffold-GYRP-2000502-Euastrum affine	Desmidiaceae	acidic endochitinase-like	2,9316E-43	63.14	IPR001223 (PFAM)
scaffold-GYRP-2000503-Euastrum affine	Desmidiaceae	acidic endochitinase-like	9,96149E-60	57.95	IPR001223 (PFAM)
scaffold-HAOX-2002128-Spirogyra sp	Zygnemataceae	acidic endochitinase	1,39013E-74	64.29	IPR001223 (PFAM)
scaffold-HAOX-2024957-Spirogyra sp	Zygnemataceae	acidic endochitinase	2,31867E-74	64.45	IPR001223 (PFAM)
scaffold-HHXJ-2002324-Tetraselmis striata	Chlorodendraceae	chitinase	1,77379E-46	67.76	IPR001223 (PFAM)
scaffold-HIDG-2009902-Cosmarium broomei	Desmidiaceae	AChain A	1,58101E-64	57.68	IPR001223 (PFAM)

GREEN ALGAE – GH18					
SeqName	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-HIDG-2009903-Cosmarium broomei	Desmidiaceae	AChain A	9,26631E-65	57.67	IPR001223 (PFAM)
scaffold-HIDG-2009904-Cosmarium broomei	Desmidiaceae	acidic mammalian chitinase-like	2,52567E-52	59.5	IPR001223 (PFAM)
scaffold-HIDG-2018628-Cosmarium broomei	Desmidiaceae	hevamine-A-like	3,17674E-37	63.65	IPR001223 (PFAM)
scaffold-HJVM-2001428-Cosmarium ochthodes	Desmidiaceae	Acidic endochitinase	3,37497E-83	66.04	IPR001223 (PFAM)
scaffold-HJVM-2105402-Cosmarium ochthodes	Desmidiaceae	acidic mammalian chitinase-like	8,89255E-17	60.86	IPR001223 (PFAM)
scaffold-HJVM-2110782-Cosmarium ochthodes	Desmidiaceae	acidic endochitinase-like	2,68544E-77	61.92	IPR001223 (PFAM)
scaffold-HKZW-2012228-Mesotaenium caldariorum	Mesotaeniaceae	acidic mammalian chitinase	4,5789E-54	57.22	IPR001223 (PFAM)
scaffold-HVNO-2013597-Tetraselmis chui	Chlorodendraceae	chitinase	4,62445E-61	67.87	IPR001223 (PFAM)
scaffold-IRYH-2007489-Heterochlamydomonas inaequalis	Chlamydomonadaceae	probable chitinase 3	6,19725E-103	52.09	IPR001223 (PFAM)
scaffold-IRYH-2025398-Heterochlamydomonas inaequalis	Chlamydomonadaceae	acidic mammalian chitinase-like	1,42798E-21	61.8	IPR001223 (PFAM)
scaffold-ISHC-2004481-Staurastrum sebaldi	Desmidiaceae	hevamine-A-like	1,01518E-65	64.11	IPR001223 (PFAM)
scaffold-ISHC-2004482-Staurastrum sebaldi	Desmidiaceae	acidic endochitinase-like	3,14706E-67	63.55	IPR001223 (PFAM)
scaffold-SIM-2003838-Nephroselmis pyriformis	Nephroselmidaceae	acidic mammalian chitinase-like	5,78804E-60	62.12	IPR001223 (PFAM)
scaffold-ISPU-2004539-Volvox globator	Volvocaceae	chitotriosidase-1 isoform X2	1,95846E-137	53.24	IPR001223 (PFAM)
scaffold-ISPU-2004540-Volvox globator	Volvocaceae	chitinase	2,16629E-125	54.03	IPR001223 (PFAM)
scaffold-JKKI-2008476-Lobomonas rostrata	Chlamydomonadaceae	glycoside hydrolase family 18	5,491E-28	54.53	IPR001223 (PFAM)
scaffold-JMTE-2014447-Pseudoscourfieldia marina	Pycnococcaceae	chitotriosidase-1-like	3,6133E-39	74.11	IPR001223 (PFAM)
scaffold-JMTE-2014781-Pseudoscourfieldia marina	Pycnococcaceae	chitotriosidase-1-like isoform X2	1,10422E-33	80.8	IPR001223 (PFAM)
scaffold-JMTE-2015199-Pseudoscourfieldia marina	Pycnococcaceae	acidic endochitinase-like	1,2356E-47	85.06	IPR001223 (PFAM)
scaffold-JOJQ-2005804-Cylindrocystis cushleackae	Zygnemataceae	acidic endochitinase-like	7,69732E-85	66.84	IPR001223 (PFAM)
scaffold-JOJQ-2040355-Cylindrocystis cushleackae	Zygnemataceae	acidic mammalian chitinase-like	2,2866E-59	54.32	IPR001223 (PFAM)
scaffold-KADG-2007209-Ignatius tetrasporus	Ulotrichaceae	related to chitinase	5,96352E-14	57.08	IPR001223 (PFAM)
scaffold-KEYW-2025590-Gonatozygon kinahanii	Gonatozygaceae	Acidic endochitinase	1,58417E-80	65.86	IPR001223 (PFAM)
scaffold-KEYW-2026154-Gonatozygon kinahanii	Gonatozygaceae	acidic mammalian chitinase-like	2,14258E-66	56.52	IPR001223 (PFAM)
scaffold-KMNX-2031577-Nucleotaenium eifelense	Mesotaeniaceae	class V chitinase	8,25201E-12	68.51	IPR001223 (PFAM)
scaffold-KMNX-2035459-Nucleotaenium eifelense	Mesotaeniaceae	AChain A	5,65275E-18	63.27	IPR001223 (PFAM)
scaffold-KUJU-2004605-Gonium pectorale	Goniaceae	chitinase A1	1,88791E-170	54.47	IPR001223 (PFAM)
scaffold-LFVP-2004510-Marchantia paleacea-2 samples combined	Marchantiaceae	hevamine-A-like	4,58662E-151	75.07	IPR001223 (PFAM)
scaffold-LFVP-2009172- Marchantia paleacea-2 samples combined	Marchantiaceae	acidic mammalian chitinase-like	0.0	61.69	IPR001223 (PFAM)
scaffold-LFVP-2009173- Marchantia paleacea-2 samples combined	Marchantiaceae	acidic mammalian chitinase-like	0.0	61.69	IPR001223 (PFAM)
scaffold-LFVP-2081766-Marchantia paleacea-2 samples combined	Marchantiaceae	acidic endochitinase	4,00518E-135	75.0	IPR001223 (PFAM)
scaffold-LFVP-2083179- Marchantia paleacea-2 samples combined	Marchantiaceae	acidic endochitinase-like	7,04516E-93	67.43	IPR001223 (PFAM)

GREEN ALGAE – GH18					
SeqName	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-LFVP-2083339-Marchantia_paleacea-2 samples combined	Marchantiaceae	acidic endochitinase-like	1,12303E-84	66.22	IPR001223 (PFAM)
scaffold-LFVP-2083413-Marchantia_paleacea-2 samples combined	Marchantiaceae	hevamine A precursor-like protein	3,41875E-157	67.63	IPR001223 (PFAM)
scaffold-LFVP-2083566-Marchantia_paleacea-2 samples combined	Marchantiaceae	acidic endochitinase-like	2,3841E-173	69.82	IPR001223 (PFAM)
scaffold-LNIL-2052393-Pteromonas_angulosa	Phacotaceae	chitotriosidase-1 isoform X1	4,53761E-56	51.64	IPR001223 (PFAM)
scaffold-MCHJ-2000297-Micrasterias_fimbriata	Desmidiaceae	acidic mammalian chitinase-like	8,43705E-40	50.59	IPR001223 (PFAM)
scaffold-MCHJ-2001441-Micrasterias_fimbriata	Desmidiaceae	AChain A	2,3481E-57	57.02	IPR001223 (PFAM)
scaffold-MFZO-2017215-Zygnemopsis_sp	Zygnemataceae	glycoside hydrolase family 18	1,94157E-42	74.89	IPR001223 (PFAM)
scaffold-MFZO-2022650-Zygnemopsis_sp	Zygnemataceae	AChain A	1,97128E-49	53.98	IPR001223 (PFAM)
scaffold-MMKU-2006934-Nephroselmis_olivace	Nephroselmidaceae	acidic mammalian chitinase-like	4,55345E-92	62.13	IPR001223 (PFAM)
scaffold-MNCB-2024465-Eremosphaera_viridi	Oocystaceae	AChain A	4,90316E-56	55.57	IPR001223 (PFAM)
scaffold-MNCB-2024466-Eremosphaera_viridi	Oocystaceae	chitotriosidase-1 precursor	1,25264E-56	55.85	IPR001223 (PFAM)
scaffold-MNCB-2024468-Eremosphaera_viridi	Oocystaceae	AChain A	2,18804E-47	53.45	IPR001223 (PFAM)
scaffold-MNNM-2000121-Cosmarium_granatum	Desmidiaceae	Glycoside hydrolase	1,28289E-76	66.58	IPR001223 (PFAM)
scaffold-MNNM-2037700-Cosmarium_granatum	Desmidiaceae	acidic endochitinase-like	3,30384E-45	68.26	IPR001223 (PFAM)
scaffold-MOYY-2009074-Pleurotaenium_trabecul	Desmidiaceae	AChain A	3,69547E-12	60.82	IPR001223 (PFAM)
scaffold-MOYY-2018456-Pleurotaenium_trabecul	Desmidiaceae	acidic mammalian chitinase-like	2,23137E-63	56.61	IPR001223 (PFAM)
scaffold-MOYY-2018457-Pleurotaenium_trabecul	Desmidiaceae	AChain A	1,39958E-65	56.34	IPR001223 (PFAM)
scaffold-MULF-2041340-Chlamydomonas_bilatus-A	Chlamydomonadaceae	related to chitinase	1,48889E-30	55.7	IPR001223 (PFAM)
scaffold-NBYP-2006024-Mesotaenium_kramstei	Mesotaeniaceae	acidic mammalian chitinase-like	2,4348E-64	53.56	IPR001223 (PFAM)
scaffold-OFUE-2010323-Lobochlamys_segnis	Chlamydomonadaceae	bacteriodes thetaiotaomicron symbiotic chitinase	1,24825E-57	50.79	IPR001223 (PFAM)
scaffold-OFUE-2047377-Lobochlamys_segnis	Chlamydomonadaceae	Chitinase 18-5	1,70246E-47	52.93	IPR001223 (PFAM)
scaffold-OTQG-2001226-Ankistrodesmus_sp	Ankistrodesmaceae	acidic mammalian chitinase-like	8,86151E-122	52.83	IPR001223 (PFAM)
scaffold-OTQG-2001227-Ankistrodesmus_sp	Ankistrodesmaceae	acidic mammalian chitinase-like	8,86151E-122	52.83	IPR001223 (PFAM)
scaffold-PFUD-2002294-Geminella_sp	Chlorellaceae	class V chitinase	9,05005E-28	48.85	IPR001223 (PFAM)
scaffold-PFUD-2034986-Geminella_sp	Chlorellaceae	glycoside hydrolase	5,22709E-48	71.92	IPR001223 (PFAM)
scaffold-POIR-2027547-Volvox_aureus-2 samples combined	Volvocaceae	probable chitinase 3	1,73593E-117	54.25	IPR001223 (PFAM)
scaffold-PZBH-2032995-Oltmannsiellopsis_viridis-A	Oltmannsiellopsidaceae	acidic mammalian chitinase-like	4,77263E-33	69.06	IPR001223 (PFAM)
scaffold-PZIF-2015162-Scenedesmus_dimorphus	Scenedesmaceae	chitotriosidase-1 isoform X1	3,04739E-64	52.23	IPR001223 (PFAM)
scaffold-QJYX-2001962-Oltmannsiellopsis_viridis-B	Oltmannsiellopsidaceae	AChain A	3,31738E-74	56.94	IPR001223 (PFAM)
scaffold-QPDY-2002748-Coleochaete_irregularis	Coleochaetaceae	Acidic mammalian chitinase	1,57104E-07	47.14	IPR001223 (PFAM)
scaffold-QPDY-2030822-Coleochaete_irregularis	Coleochaetaceae	acidic mammalian chitinase-like	8,39575E-20	44.73	IPR001223 (PFAM)
scaffold-QPDY-2031501-Coleochaete_irregularis	Coleochaetaceae	chitotriosidase-1 isoform X1	4,91999E-71	53.67	IPR001223 (PFAM)
scaffold-QWFV-2003026-Bambusina_borreri	Desmidiaceae	acidic mammalian chitinase-like	2,3847E-51	55.21	IPR001223 (PFAM)
scaffold-QWFV-2003027-Bambusina_borreri	Desmidiaceae	acidic mammalian chitinase-like	1,00078E-13	55.45	IPR001223 (PFAM)
scaffold-QWFV-2003029-Bambusina_borreri	Desmidiaceae	acidic mammalian chitinase-like	4,65254E-35	54.4	IPR001223 (PFAM)

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SeqName	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-QWFV-2003030-Bambusina borneri	Desmidiaceae	acidic mammalian chitinase-like	2,3847E-51	55.21	IPR001223 (PFAM)
scaffold-QWFV-2005300-Bambusina borneri	Desmidiaceae	acidic endochitinase	1,30932E-67	64.06	IPR001223 (PFAM)
scaffold-QWFV-2005301-Bambusina borneri	Desmidiaceae	acidic endochitinase	1,30932E-67	64.06	IPR001223 (PFAM)
scaffold-QWFV-2012344-Bambusina borneri	Desmidiaceae	hevamine-A-like	3,7377E-53	64.37	IPR001223 (PFAM)
scaffold-WFV-2020226-Bambusina borneri	Desmidiaceae	acidic endochitinase	1,25489E-43	73.67	IPR001223 (PFAM)
scaffold-QWFV-2020306-Bambusina borneri	Desmidiaceae	hevamine-A-like	1,17698E-68	63.88	IPR001223 (PFAM)
scaffold-QYXY-2008419-Botryococcus terribilis	Botryococcaceae	acidic endochitinase	1,50253E-20	64.57	IPR001223 (PFAM)
scaffold-QYXY-2040497-Botryococcus terribilis	Botryococcaceae	glycoside hydrolase	6,12575E-21	64.09	IPR001223 (PFAM)
scaffold-RNAT-2058594-Eudorina elegans	Volvocaceae	acidic mammalian chitinase-like	1,1601E-176	52.66	IPR001223 (PFAM)
scaffold-RPGL-2014644-Cylindrocystis brebissonii-M2853	Mesotaeniaceae	acidic mammalian chitinase-like	9,71076E-19	54.29	IPR001223 (PFAM)
scaffold-RPGL-2027497-Cylindrocystis brebissonii-M2853	Mesotaeniaceae	acidic endochitinase	7,87281E-60	65.83	IPR001223 (PFAM)
scaffold-RPQV-2005631-Phymatodocis nordstedtiana	Desmidiaceae	hevamine-A-like	5,78316E-71	61.06	IPR001223 (PFAM)
scaffold-RPQV-2005632-Phymatodocis nordstedtiana	Desmidiaceae	hevamine-A-like	2,62744E-63	62.28	IPR001223 (PFAM)
scaffold-RPQV-2006998-Phymatodocis nordstedtiana	Desmidiaceae	hevamine-A-like	8,04738E-64	62.21	IPR001223 (PFAM)
scaffold-RPQV-2007705-Phymatodocis nordstedtiana	Desmidiaceae	acidic endochitinase-like	4,18562E-97	66.9	IPR001223 (PFAM)
scaffold-RPQV-2036136-Phymatodocis nordstedtiana	Desmidiaceae	AChain A	9,46972E-69	57.99	IPR001223 (PFAM)
scaffold-RPRU-2009659-Staurodesmus omearii	Desmidiaceae	hevamine-A-like	9,12319E-52	60.64	IPR001223 (PFAM)
scaffold-RPRU-2039277-Staurodesmus omearii	Desmidiaceae	acidic endochitinase-like	5,75826E-89	66.85	IPR001223 (PFAM)
scaffold-RQFE-2000106-Cosmocladium cf. constrictum	Desmidiaceae	acidic mammalian chitinase-like	8,02421E-46	55.18	IPR001223 (PFAM)
scaffold-RQFE-2000107-Cosmocladium cf. constrictum	Desmidiaceae	acidic mammalian chitinase	3,60652E-41	55.07	IPR001223 (PFAM)
scaffold-RQFE-2007301-Cosmocladium cf. constrictum	Desmidiaceae	Acidic endochitinase	5,25319E-84	67.06	IPR001223 (PFAM)
scaffold-RYJX-2062142-Pandorina morum	Volvocaceae	acidic mammalian chitinase-like	8,3005E-150	52.87	IPR001223 (PFAM)
scaffold-SNOX-2004982-Planotaenium ohtanii	Mesotaeniaceae	probable chitinase 3	1,47872E-60	58.17	IPR001223 (PFAM)
scaffold-STKJ-2099024-Zygnema sp.-A	Zygnemataceae	acidic endochitinase-like	8,78342E-76	61.66	IPR001223 (PFAM)
scaffold-STKJ-2099840-Zygnema sp.-A	Zygnemataceae	Acidic endochitinase	3,37497E-83	66.04	IPR001223 (PFAM)
scaffold-UKUC-2105707-Dunaliella salina-UTEX LB 200	Dunaliellaceae	Acidic endochitinase	2,0939E-88	88.48	IPR001223 (PFAM)
scaffold-UKUC-2108261-Dunaliella salina-UTEX LB 200	Dunaliellaceae	probable chitinase 2	8,97051E-32	61.25	IPR001223 (PFAM)
scaffold-UTRE-2015935-Chloromonas tughillensi	Chlamydomonadaceae	acidic mammalian chitinase-like	1,45728E-87	51.26	IPR001223 (PFAM)
scaffold-UTRE-2015936-Chloromonas tughillensi	Chlamydomonadaceae	acidic mammalian chitinase-like	1,48853E-94	52.02	IPR001223 (PFAM)
scaffold-UTRE-2084885-Chloromonas tughillensi	Chlamydomonadaceae	carbohydrate-binding module family 5	3,26042E-36	68.76	IPR001223 (PFAM)
scaffold-VAZE-2000533-Cylindrocystis sp	Mesotaeniaceae	Acidic endochitinase	1,33575E-18	75.66	IPR001223 (PFAM)
scaffold-VAZE-2009036-Cylindrocystis sp	Mesotaeniaceae	acidic mammalian chitinase-like	7,76265E-56	55.78	IPR001223 (PFAM)
scaffold-VBLH-2007028-Cladophora glomerata	Cladophoraceae	acidic mammalian chitinase-like	7,36388E-32	53.6	IPR001223 (PFAM)
scaffold-VHIJ-2008486-Blastophysa cf. rhizopus	Chaetosiphonaceae	chitotriosidase-1 precursor	3,16866E-21	52.23	IPR001223 (PFAM)
scaffold-VHIJ-2008487-Blastophysa cf. rhizopus	Chaetosiphonaceae	chitotriosidase-1 precursor	3,16866E-21	52.23	IPR001223 (PFAM)
scaffold-VIAU-2009074-Carteria crucifera	Carteriaceae	acidic mammalian chitinase-like	1,65806E-21	54.78	IPR001223 (PFAM)
scaffold-VIAU-2009075-Carteria crucifera	Carteriaceae	acidic mammalian chitinase-like	1,74084E-21	54.78	IPR001223 (PFAM)
scaffold-VIAU-2053932-Carteria crucifera	Carteriaceae	acidic mammalian chitinase-like	1,47776E-08	66.93	IPR001223 (PFAM)

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SeqName	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-VQBJ-2003884-Coleochaete scutata	Coleochaetaceae	AChain A	1,49241E-06	50.52	IPR001223 (PFAM)
scaffold-VQBJ-2004216-Coleochaete scutata	Coleochaetaceae	acidic mammalian chitinase-like	1,28633E-12	51.83	IPR001223 (PFAM)
scaffold-VQBJ-2009052-Coleochaete scutata	Coleochaetaceae	acidic mammalian chitinase-like	2,69157E-19	58.73	IPR001223 (PFAM)
scaffold-WCQU-2007350-Staurodesmus convergens	Desmidiaceae	acidic mammalian chitinase-like	6,58893E-50	57.27	IPR001223 (PFAM)
scaffold-WDCW-2045137-Mesotaenium endlicherianum	Mesotaeniaceae	acidic mammalian chitinase-like	2,18734E-39	60.97	IPR001223 (PFAM)
scaffold-WDCW-2046043-Mesotaenium endlicherianum	Mesotaeniaceae	acidic endochitinase-like	1,14709E-78	62.84	IPR001223 (PFAM)
scaffold-WDGV-2049195-Cosmarium subtumidum	Desmidiaceae	acidic mammalian chitinase-like	1,98274E-38	57.85	IPR001223 (PFAM)
scaffold-WDWX-2034378-Dunaliella primolecta-UTEX LB 1000	Dunaliellaceae	Acidic endochitinase	1,25342E-58	88.85	IPR001223 (PFAM)
scaffold-WGMD-3000448-Zygnema sp.-B	Zygnemataceae	acidic endochitinase-like	6,87803E-77	65.33	IPR001223 (PFAM)
scaffold-WGMD-3000449-Zygnema sp.-B	Zygnemataceae	acidic endochitinase-like	6,87803E-77	65.33	IPR001223 (PFAM)
scaffold-WGMD-3000450-Zygnema sp.-B	Zygnemataceae	acidic endochitinase-like	6,87803E-77	65.33	IPR001223 (PFAM)
scaffold-WGMD-3000451-Zygnema sp.-B	Zygnemataceae	Acidic endochitinase	6,2214E-39	73.96	IPR001223 (PFAM)
scaffold-WGMD-3003070-Zygnema sp.-B	Zygnemataceae	chitotriosidase-1 isoform X1	9,17075E-47	59.96	IPR001223 (PFAM)
scaffold-WGMD-3003071-Zygnema sp.-B	Zygnemataceae	chitotriosidase-1 isoform X1	9,17075E-47	59.96	IPR001223 (PFAM)
scaffold-WGMD-3014691-Zygnema sp.-B	Zygnemataceae	acidic mammalian chitinase-like	1,28285E-15	56.35	IPR001223 (PFAM)
scaffold-WRSL-2006874-Volvox aureus-M2242	Volvocaceae	chitinase C	5,35671E-171	55.68	IPR001223 (PFAM)
scaffold-WSJO-2006870-Mesotaenium braunii	Mesotaeniaceae	Acidic mammalian chitinase	1,59936E-25	58.12	IPR001223 (PFAM)
scaffold-WSJO-2048613-Mesotaenium braunii	Mesotaeniaceae	Chitinase	6,36077E-60	50.14	IPR001223 (PFAM)
scaffold-XDLL-2006213-Oogamochlamys gigantea	Chlamydomonadaceae	Chitinase 18-5	1,70246E-47	52.93	IPR001223 (PFAM)
scaffold-XDLL-2006214-Oogamochlamys gigantea	Chlamydomonadaceae	Chitinase	4,44353E-63	52.81	IPR001223 (PFAM)
scaffold-XIVI-2090327-Cymbomonas sp	Pyramimonadaceae	glycoside hydrolase family 18	2,64631E-13	52.64	IPR001223 (PFAM)
scaffold-XRTZ-2004862-Roya obtusa	Mesotaeniaceae	acidic mammalian chitinase-like	1,50556E-57	59.26	IPR001223 (PFAM)
scaffold-YOXI-2002515-Cylindrocystis brebissonii-M2213	Mesotaeniaceae	acidic mammalian chitinase	6,46442E-53	57.26	IPR001223 (PFAM)
scaffold-YOXI-2007181-Cylindrocystis brebissonii-M2213	Mesotaeniaceae	hevamine-A-like	1,67289E-71	66.98	IPR001223 (PFAM)
scaffold-YSQT-2005657-Penium exiguum	Peniaceae	AChain A	1,2438E-63	62.69	IPR001223 (PFAM)
scaffold-YSQT-2032479-Penium exiguum	Peniaceae	glycoside hydrolase family 18	8,51608E-36	62.69	IPR001223 (PFAM)
scaffold-YSQT-2036100-Penium exiguum	Peniaceae	hevamine-A-like	1,8449E-82	66.93	IPR001223 (PFAM)
scaffold-ZNUM-2002100-Leptosira obovata	Leptosiraceae	glycoside hydrolase	2,42083E-39	48.42	IPR001223 (PFAM)
scaffold-ZNUM-2002101-Leptosira obovata	Leptosiraceae	glycoside hydrolase	8,06833E-37	49.16	IPR001223 (PFAM)
scaffold-ZRMT-2003702-Mougeotia sp	Zygnemataceae	acidic endochitinase-like	3,38161E-80	65.93	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-ABCD-2061358-Niphotrichum elongatum	Mosses	Grimmiaceae	AChain A	9,20213E-39	60.27	IPR001223 (PFAM)
scaffold-AEXY-2001889-Blasia sp	Liverworts	Blasiaceae	Chitinase	1,50485E-69	80.81	IPR001223 (PFAM)
scaffold-AEXY-2002225-Blasia sp	Liverworts	Blasiaceae	glycoside hydrolase family 18	1,13991E-49	91.29	IPR001223 (PFAM)
scaffold-AEXY-2002415-Blasia sp	Liverworts	Blasiaceae	glycosyl hydrolase family 18	1,10586E-55	69.44	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-AEXY-2010785-Blasia_sp	Liverworts	Blasiaceae	Chitinase	2,60338E-91	90.89	IPR001223 (PFAM)
scaffold-AEXY-2015542-Blasia_sp	Liverworts	Blasiaceae	chitinase 18-7	3,77538E-127	85.09	IPR001223 (PFAM)
scaffold-AEXY-2142432-Blasia_sp	Liverworts	Blasiaceae	class V chitinase	1,02141E-106	85.45	IPR001223 (PFAM)
scaffold-AEXY-2149299-Blasia_sp	Liverworts	Blasiaceae	class V chitinase ChiB1	7,35261E-150	71.57	IPR001223 (PFAM)
scaffold-AEXY-2151689-Blasia_sp	Liverworts	Blasiaceae	Chitinase	2,4465E-91	100.0	IPR001223 (PFAM)
scaffold-AEXY-2152298-Blasia_sp	Liverworts	Blasiaceae	brain chitinase and chia	2,88425E-15	68.61	IPR001223 (PFAM)
scaffold-AKXB-2067103-Phaeomegaceros_coriaceus	Hornworts	Dendrocerotaceae	acidic mammalian chitinase-like	2,14273E-98	62.56	IPR001223 (PFAM)
scaffold-ANON-2000173-Leiosporoceros_dussii-B	Hornworts	Leiosporocerotaceae	acidic mammalian chitinase-like	4,90038E-76	62.39	IPR001223 (PFAM)
scaffold-AWOI-2003923-Diphyscium_foliosum	Mosses	Diphysciaceae	glycoside hydrolase	1,68908E-39	69.15	IPR001223 (PFAM)
scaffold-AWOI-2069936-Diphyscium_foliosum	Mosses	Diphysciaceae	class V chitinase-like	2,13322E-111	63.27	IPR001223 (PFAM)
scaffold-AWOI-2072168-Diphyscium_foliosum	Mosses	Diphysciaceae	acidic mammalian chitinase-like	5,97531E-162	62.52	IPR001223 (PFAM)
scaffold-BGXB-2075632-Plagiommium_insigne	Mosses	Mielichhoferiaceae	acidic mammalian chitinase-like	1,62616E-86	65.59	IPR001223 (PFAM)
scaffold-BPSG-2088193-Scouleria_aquatica	Mosses	Scouleriaceae	class V chitinase-like	3,56822E-171	64.32	IPR001223 (PFAM)
scaffold-BSNI-2050540-Anthoceros_agrestis-B	Hornworts	Anthoceroceae	acidic mammalian chitinase-like	9,8284E-93	64.35	IPR001223 (PFAM)
scaffold-CHJJ-2002473-Lejeuneaceae_sp	Liverworts	Frullaniaceae	acidic mammalian chitinase	5,44865E-15	64.43	IPR001223 (PFAM)
scaffold-CHJJ-2019067-Lejeuneaceae_sp	Liverworts	Frullaniaceae	acidic mammalian chitinase-like	1,63216E-52	61.5	IPR001223 (PFAM)
scaffold-CHJJ-2135670-Lejeuneaceae_sp	Liverworts	Frullaniaceae	glycoside hydrolase	6,58039E-72	63.36	IPR001223 (PFAM)
scaffold-CHJJ-2137152-Lejeuneaceae_sp	Liverworts	Frullaniaceae	class V chitinase-like	1,35375E-124	58.36	IPR001223 (PFAM)
scaffold-CMEQ-2010358-Orthotrichum_lyellii	Mosses	Orthotrichaceae	acidic mammalian chitinase-like	0.0	62.91	IPR001223 (PFAM)
scaffold-CMEQ-2084544-Orthotrichum_lyellii	Mosses	Orthotrichaceae	Chitinase 1	1,41156E-151	83.41	IPR001223 (PFAM)
scaffold-DHWX-2004311-Fontinalis_antipyretica	Mosses	Fontinalaceae	acidic mammalian chitinase-like	4,93069E-172	61.95	IPR001223 (PFAM)
scaffold-DXOU-2037702-Nothoceros_aenigmaticus	Hornworts	Dendrocerotaceae	acidic mammalian chitinase-like	6,54902E-94	62.44	IPR001223 (PFAM)
scaffold-EEMJ-2001989-Thuidium_delicatulum	Mosses	Thuidiaceae	class V chitinase	3,84084E-152	61.02	IPR001223 (PFAM)
scaffold-EEMJ-2001990-Thuidium_delicatulum	Mosses	Thuidiaceae	class V chitinase	2,89724E-56	60.49	IPR001223 (PFAM)
scaffold-FFPD-2013361-Ceratodon_purpureus	Mosses	Ditrichaceae	acidic mammalian chitinase	1,35708E-151	63.94	IPR001223 (PFAM)
scaffold-FITN-2000573-Treubia_lacunose	Liverworts	Treubiaceae	acidic mammalian chitinase-like	1,89337E-125	67.86	IPR001223 (PFAM)
scaffold-FITN-2000574-Treubia_lacunose	Liverworts	Treubiaceae	acidic mammalian chitinase-like	3,27295E-32	66.86	IPR001223 (PFAM)
scaffold-FITN-2000575-Treubia_lacunose	Liverworts	Treubiaceae	class V chitinase	6,29708E-125	67.04	IPR001223 (PFAM)
scaffold-FITN-2007298-Treubia_lacunose	Liverworts	Treubiaceae	acidic endochitinase	1,42066E-102	75.86	IPR001223 (PFAM)
scaffold-FITN-2007299-Treubia_lacunose	Liverworts	Treubiaceae	acidic endochitinase	3,43287E-87	71.08	IPR001223 (PFAM)
scaffold-FITN-2007300-Treubia_lacunose	Liverworts	Treubiaceae	acidic endochitinase	1,97106E-100	77.93	IPR001223 (PFAM)
scaffold-FITN-2007301-Treubia_lacunose	Liverworts	Treubiaceae	acidic endochitinase	3,68093E-101	78.23	IPR001223 (PFAM)
scaffold-FITN-2013117-Treubia_lacunose	Liverworts	Treubiaceae	class V chitinase	4,77637E-84	57.67	IPR001223 (PFAM)
scaffold-FITN-2013118-Treubia_lacunose	Liverworts	Treubiaceae	class V chitinase	2,46802E-74	59.6	IPR001223 (PFAM)
scaffold-FITN-2016127-Treubia_lacunose	Liverworts	Treubiaceae	class V chitinase	2,20471E-103	65.88	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-FITN-2017805-Treubia_lacunose	Liverworts	Treubiaceae	Glycosyl hydrolase family protein with chitinase insertion domain-containing	2,91922E-111	69.16	IPR001223 (PFAM)
scaffold-FITN-2091255-Treubia_lacunose	Liverworts	Treubiaceae	Glycosyl hydrolase family protein with chitinase insertion domain-containing	1,80846E-128	66.44	IPR001223 (PFAM)
scaffold-GOWD-2009968-Sphagnum_lescurii	Mosses	Sphagnaceae	chitotriosidase-1 isoform X1	6,0112E-91	58.66	IPR001223 (PFAM)
scaffold-GOWD-2019400-Sphagnum_lescurii	Mosses	Sphagnaceae	Chitinase-like protein Idgf4	1,01141E-85	80.2	IPR001223 (PFAM)
scaffold-GOWD-2019795-Sphagnum_lescurii	Mosses	Sphagnaceae	Chitinase-like protein Idgf4	3,9289E-133	68.83	IPR001223 (PFAM)
scaffold-GOWD-2079062-Sphagnum_lescurii	Mosses	Sphagnaceae	acidic mammalian chitinase-like	1,601E-57	72.68	IPR001223 (PFAM)
scaffold-GOWD-2082285-Sphagnum_lescurii	Mosses	Sphagnaceae	chitinase-like protein Idgf4	1,60351E-119	79.31	IPR001223 (PFAM)
scaffold-GOWD-2083251-Sphagnum_lescurii	Mosses	Sphagnaceae	glycoside hydrolase family 18	8,89648E-103	75.44	IPR001223 (PFAM)
scaffold-GRKU-2005548-Syntrichia_princeps	Mosses	Pottiaceae	acidic mammalian chitinase	1,04373E-132	66.43	IPR001223 (PFAM)
scaffold-HERT-2006371-Sphaerocarpos_texanus	Liverworts	Sphaerocarpaceae	acidic mammalian chitinase	1,12565E-48	63.46	IPR001223 (PFAM)
scaffold-HERT-2038302-Sphaerocarpos_texanus	Liverworts	Sphaerocarpaceae	acidic mammalian chitinase	5,30488E-43	63.79	IPR001223 (PFAM)
scaffold-HMHL-2009920-Marchantia_paleacea-non_mycorrhizal	Liverworts	Marchantiaceae	acidic mammalian chitinase-like	0.0	61.69	IPR001223 (PFAM)
scaffold-HMHL-2052653-Marchantia_paleacea-non_mycorrhizal	Liverworts	Marchantiaceae	hevamine A precursor-like protein	3,41875E-157	67.63	IPR001223 (PFAM)
scaffold-HMHL-2053338-Marchantia_paleacea-non_mycorrhizal	Liverworts	Marchantiaceae	hevamine-A-like	4,58662E-151	75.07	IPR001223 (PFAM)
scaffold-HPXA-2012251-Ptilidium_pulcherrimum	Liverworts	Ptilidiaceae	AChain A	6,82885E-74	61.11	IPR001223 (PFAM)
scaffold-HPXA-2018253-Ptilidium_pulcherrimum	Liverworts	Ptilidiaceae	glycoside hydrolase family 18	2,26627E-46	52.85	IPR001223 (PFAM)
scaffold-HPXA-2018545-Ptilidium_pulcherrimum	Liverworts	Ptilidiaceae	Endochitinase 1	6,26944E-98	78.46	IPR001223 (PFAM)
scaffold-HPXA-21 scaffold-22978-Ptilidium_pulcherrimum	Liverworts	Ptilidiaceae	Chitinase 1	1,47149E-26	62.48	IPR001223 (PFAM)
scaffold-HPXA-2133118-Ptilidium_pulcherrimum	Liverworts	Ptilidiaceae	acidic mammalian chitinase-like	3,22663E-97	64.81	IPR001223 (PFAM)
scaffold-HVBQ-2126934-Tetraphis_pellucida	Mosses	Tetraphidaceae	acidic mammalian chitinase-like	1,36213E-64	66.28	IPR001223 (PFAM)
scaffold-IGUH-2002948-Schwetschkeopsis_fabronia	Mosses	Leucodontaceae	class V chitinase-like	0.0	63.17	IPR001223 (PFAM)
scaffold-IGUH-2002949-Schwetschkeopsis_fabronia	Mosses	Leucodontaceae	class V chitinase-like	0.0	63.17	IPR001223 (PFAM)
scaffold-IGUH-2158382-Schwetschkeopsis_fabronia	Mosses	Leucodontaceae	glycoside hydrolase family 18 protein	9,78462E-33	66.58	IPR001223 (PFAM)
scaffold-IGUH-2163943-Schwetschkeopsis_fabronia	Mosses	Leucodontaceae	Chitinase	1,23696E-65	69.43	IPR001223 (PFAM)
scaffold-IGUH-2164561-Schwetschkeopsis_fabronia	Mosses	Leucodontaceae	acidic mammalian chitinase-like	2,81279E-39	62.16	IPR001223 (PFAM)
scaffold-IHWO-2005936-Marchantia_paleacea-mycorrhizal	Liverworts	Marchantiaceae	hevamine-A-like	4,58662E-151	75.07	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-IHWO-2011148-Marchantia_paleacea-mycorrhizal	Liverworts	Marchantiaceae	acidic mammalian chitinase-like	0.0	61.69	IPR001223 (PFAM)
scaffold-IHWO-2065167-Marchantia_paleacea-mycorrhizal	Liverworts	Marchantiaceae	acidic endochitinase-like	7,04516E-93	67.43	IPR001223 (PFAM)
scaffold-IHWO-2065195-Marchantia_paleacea-mycorrhizal	Liverworts	Marchantiaceae	hevamine A precursor-like protein	3,41875E-157	67.63	IPR001223 (PFAM)
scaffold-IHWO-2065225-Marchantia_paleacea-mycorrhizal	Liverworts	Marchantiaceae	acidic endochitinase-like	2,3841E-173	69.82	IPR001223 (PFAM)
scaffold-IHWO-2065363-Marchantia_paleacea-mycorrhizal	Liverworts	Marchantiaceae	acidic endochitinase-like	1,12303E-84	66.22	IPR001223 (PFAM)
scaffold-ILBQ-2043168-Conocephalum_conicum	Liverworts	Conocephalaceae	acidic endochitinase-like	1,52664E-137	73.27	IPR001223 (PFAM)
scaffold-ILBQ-2045545-Conocephalum_conicum	Liverworts	Conocephalaceae	acidic mammalian chitinase-like	4,39239E-174	66.09	IPR001223 (PFAM)
scaffold-IQJU-2000727-Anthoceros_formosae	Hornworts	Anthoceroceae	AChain A	3,31658E-63	57.36	PR001223 (PFAM)
scaffold-IRBN-2013604-Scapania_nemorosa	Liverworts	Scapaniaceae	Chitinase	3,02276E-75	78.75	IPR001223 (PFAM)
scaffold-IRBN-2136833-Scapania_nemorosa	Liverworts	Scapaniaceae	acidic mammalian chitinase-like	7,31856E-49	72.14	IPR001223 (PFAM)
scaffold-IRBN-2151386-Scapania_nemorosa	Liverworts	Scapaniaceae	glycoside hydrolase family 18	1,09715E-58	63.64	IPR001223 (PFAM)
scaffold-IRBN-2152154-Scapania_nemorosa	Liverworts	Scapaniaceae	glycoside hydrolase	8,92567E-60	66.68	IPR001223 (PFAM)
scaffold-IRBN-2161616-Scapania_nemorosa	Liverworts	Scapaniaceae	putative glycoside hydrolase family 18 protein	1,50676E-86	71.44	IPR001223 (PFAM)
scaffold-IRBN-2162788-Scapania_nemorosa	Liverworts	Scapaniaceae	glycoside hydrolase family 18	1,4566E-68	61.46	IPR001223 (PFAM)
scaffold-JADL-2003775-Rhynchostegium_serrulatum	Mosses	Brachytheciaceae	acidic mammalian chitinase-like	0.0	62.26	IPR001223 (PFAM)
scaffold-JHFI-2004290-Pellia_neesiana	Liverworts	Pelliaceae	hevamine-A-like	3,71414E-65	73.13	IPR001223 (PFAM)
scaffold-JHFI-2012415-Pellia_neesiana	Liverworts	Pelliaceae	acidic endochitinase	1,74126E-87	69.23	IPR001223 (PFAM)
scaffold-JHFI-2012990-Pellia_neesiana	Liverworts	Pelliaceae	acidic endochitinase	2,83176E-90	68.52	IPR001223 (PFAM)
scaffold-JHFI-2013319-Pellia_neesiana	Liverworts	Pelliaceae	Acidic endochitinase	1,93413E-24	66.0	IPR001223 (PFAM)
scaffold-JHFI-2013440-Pellia_neesiana	Liverworts	Pelliaceae	acidic endochitinase	1,52649E-107	73.35	IPR001223 (PFAM)
scaffold-JHFI-2095885-Pellia_neesiana	Liverworts	Pelliaceae	class V chitinase	9,55848E-44	65.22	IPR001223 (PFAM)
scaffold-JHFI-2109525-Pellia_neesiana	Liverworts	Pelliaceae	glycoside hydrolase family 18	1,98464E-116	76.47	IPR001223 (PFAM)
scaffold-JHFI-2112286-Pellia_neesiana	Liverworts	Pelliaceae	acidic endochitinase	3,01572E-96	66.99	IPR001223 (PFAM)
scaffold-JMXW-2011418-Bryum_argenteum	Mosses	Bryaceae	acidic mammalian chitinase-like	5,44726E-166	62.49	IPR001223 (PFAM)
scaffold-JPYU-2007981-Marchantia_polymorpha	Liverworts	Marchantiaceae	acidic endochitinase-like	5,88343E-171	76.32	IPR001223 (PFAM)
scaffold-JPYU-2038245-Marchantia_polymorpha	Liverworts	Marchantiaceae	acidic endochitinase-like	0.0	73.7	IPR001223 (PFAM)
scaffold-JPYU-2038298-Marchantia_polymorpha	Liverworts	Marchantiaceae	acidic mammalian chitinase-like	0.0	65.15	IPR001223 (PFAM)
scaffold-KEFD-2052193-Encalypta_streptocarpa	Mosses	Encalyptaceae	acidic mammalian chitinase-like	2,76823E-11	65.38	IPR001223 (PFAM)
scaffold-KEFD-2060547-Encalypta_streptocarpa	Mosses	Encalyptaceae	class V chitinase	0.0	63.16	IPR001223 (PFAM)
scaffold-KRUQ-2106675-Porella_navicularis	Liverworts	Porellaceae	Chitinase	7,28084E-40	81.53	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-KRUQ-2106844-Porella_navicularis	Liverworts	Porellaceae	acidic mammalian chitinase isoform X2	2,96939E-33	70.75	IPR001223 (PFAM)
scaffold-KRUQ-2111018-Porella_navicularis	Liverworts	Porellaceae	glycoside hydrolase	2,93169E-72	62.19	IPR001223 (PFAM)
scaffold-KRUQ-2111802-Porella_navicularis	Liverworts	Porellaceae	glycoside hydrolase	2,28783E-84	65.94	IPR001223 (PFAM)
scaffold-LGOW-2003062-Schistochila_sp	Liverworts	Schistochilaceae	carbohydrate-binding module family 1 protein	1,09654E-158	79.9	IPR001223 (PFAM)
scaffold-LGOW-2012088-Schistochila_sp	Liverworts	Schistochilaceae	putative chitinase	7,98664E-34	78.02	IPR001223 (PFAM)
scaffold-LGOW-2016622-Schistochila_sp	Liverworts	Schistochilaceae	class III chitinase ChiA1	2,77662E-96	64.34	IPR001223 (PFAM)
scaffold-LGOW-2017619-Schistochila_sp	Liverworts	Schistochilaceae	chitinase 18-5	3,96304E-114	85.43	IPR001223 (PFAM)
scaffold-LGOW-2087427-Schistochila_sp	Liverworts	Schistochilaceae	class V chitinase	1,01574E-27	71.7	IPR001223 (PFAM)
scaffold-LGOW-2102714-Schistochila_sp	Liverworts	Schistochilaceae	glycoside hydrolase family 18	1,00455E-142	69.23	IPR001223 (PFAM)
scaffold-LGOW-2103657-Schistochila_sp	Liverworts	Schistochilaceae	hevamine-A-like	7,99851E-86	7054	IPR001223 (PFAM)
scaffold-LNSF-2002884-Hypnum_subimponens	Mosses	Pylaisiaceae	class V chitinase-like	1,07601E-167	62.93	IPR001223 (PFAM)
scaffold-LNSF-2009955-Hypnum_subimponens	Mosses	Pylaisiaceae	glycoside hydrolase family 18	6,86338E-59	68.23	IPR001223 (PFAM)
scaffold-LNSF-2011653-Hypnum_subimponens	Mosses	Pylaisiaceae	glycoside hydrolase family 18	1,38533E-54	63.07	IPR001223 (PFAM)
scaffold-LNSF-2067677-Hypnum_subimponens	Mosses	Pylaisiaceae	class V chitinase	2,81196E-109	59.18	IPR001223 (PFAM)
scaffold-LNSF-2068240-Hypnum_subimponens	Mosses	Pylaisiaceae	glycoside hydrolase family 18	7,96472E-46	63.16	IPR001223 (PFAM)
scaffold-MIRS-2086783-Climacium_dendroides	Mosses	Climaciaceae	class V chitinase-like	5,22273E-162	64.28	IPR001223 (PFAM)
scaffold-NGTD-2089690-Dicranum_scoparium	Mosses	Dicranaceae	glycoside hydrolase	6,22035E-41	70.24	IPR001223 (PFAM)
scaffold-NGTD-2090084-Dicranum_scoparium	Mosses	Dicranaceae	glycoside hydrolase family	6,5183E-19	58.84	IPR001223 (PFAM)
scaffold-NGTD-2094229-Dicranum_scoparium	Mosses	Dicranaceae	glycoside hydrolase	3,19773E-66	69.03	IPR001223 (PFAM)
scaffold-NGTD-2098818-Dicranum_scoparium	Mosses	Dicranaceae	class V chitinase	2,73451E-144	64.23	IPR001223 (PFAM)
scaffold-NRWZ-2005174-Metzgeria_crassipilis	Liverworts	Metzgeriaceae	acidic mammalian chitinase-like	1,75861E-114	59.88	IPR001223 (PFAM)
scaffold-OFTV-2008694-Barbilophozia_barbata	Liverworts	Scapaniaceae	glycoside hydrolase family 18	1,48933E-52	74.63	IPR001223 (PFAM)
scaffold-OFTV-2010947-Barbilophozia_barbata	Liverworts	Scapaniaceae	glycoside hydrolase family 18	4,45478E-60	68.02	IPR001223 (PFAM)
scaffold-OFTV-2010948-Barbilophozia_barbata	Liverworts	Scapaniaceae	glycoside hydrolase family 18	4,45478E-60	68.02	IPR001223 (PFAM)
scaffold-OFTV-2128372-Barbilophozia_barbata	Liverworts	Scapaniaceae	chitinase-3-like protein 1	3,49963E-78	73.16	IPR001223 (PFAM)
scaffold-OFTV-2131944-Barbilophozia_barbata	Liverworts	Scapaniaceae	glycoside hydrolase family 18	1,83953E-135	81.0	IPR001223 (PFAM)
scaffold-ORKS-2003937-Philonotis_fontana	Mosses	Bartramiaceae	probable chitinase 10	7,79284E-37	90.32	IPR001223 (PFAM)
scaffold-ORKS-2003938-Philonotis_fontana	Mosses	Bartramiaceae	probable chitinase 10	2,52236E-44	89.73	IPR001223 (PFAM)
scaffold-ORKS-2055632-Philonotis_fontana	Mosses	Bartramiaceae	probable chitinase 10	9,59857E-59	91.97	IPR001223 (PFAM)
scaffold-ORKS-2056670-Philonotis_fontana	Mosses	Bartramiaceae	chitinase-like protein Idgf4	1,7006E-117	74.39	IPR001223 (PFAM)
scaffold-ORKS-2056948-Philonotis_fontana	Mosses	Bartramiaceae	acidic mammalian chitinase-like	0.0	62.35	IPR001223 (PFAM)
scaffold-PIUF-2010922-Pellia_cf. Epiphylla	Liverworts	Pelliaceae	hevamine-A-like	2,69193E-64	71.95	IPR001223 (PFAM)
scaffold-PIUF-2012805-Pellia_cf. Epiphylla	Liverworts	Pelliaceae	acidic endochitinase	2,71444E-105	72.24	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-PIUF-2082270-Pellia cf. Epiphylla	Liverworts	Pelliaceae	acidic endochitinase	6,01528E-40	63.53	IPR001223 (PFAM)
scaffold-PIUF-2090613-Pellia cf. Epiphylla	Liverworts	Pelliaceae	acidic endochitinase	3,53021E-95	67.35	IPR001223 (PFAM)
scaffold-PIUF-2092517-Pellia cf. Epiphylla	Liverworts	Pelliaceae	acidic mammalian chitinase-like	5,72258E-125	61.84	IPR001223 (PFAM)
scaffold-QKQO-2049619-Pseudotaxiphyllum elegans	Mosses	Plagiotheciaceae	class V chitinase-like	1,18021E-175	63.03	IPR001223 (PFAM)
scaffold-QMWB-2012715-Anomodon attenuatus	Mosses	Myuriaceae	acidic mammalian chitinase-like	2,1312E-170	62.93	IPR001223 (PFAM)
scaffold-QMWB-2060103-Anomodon attenuatus	Mosses	Myuriaceae	putative Chitinase	1,74272E-59	60.94	IPR001223 (PFAM)
scaffold-RCBT-2024866-Sphagnum palustre	Mosses	Sphagnaceae	chitinase-3-like protein 1	1,23818E-133	70.93	IPR001223 (PFAM)
scaffold-RCBT-2029889-Sphagnum palustre	Mosses	Sphagnaceae	chitinase-3-like protein 1	1,1818E-130	70.69	IPR001223 (PFAM)
scaffold-RCBT-2175611-Sphagnum palustre	Mosses	Sphagnaceae	class III chitinase	5,65486E-26	75.36	IPR001223 (PFAM)
scaffold-RCBT-2182330-Sphagnum palustre	Mosses	Sphagnaceae	acidic mammalian chitinase-like	7,42305E-25	57.22	IPR001223 (PFAM)
scaffold-RCBT-2185854-Sphagnum palustre	Mosses	Sphagnaceae	glycoside hydrolase family 18	6,11726E-102	75.53	IPR001223 (PFAM)
scaffold-RDOO-2001513-Racomitrium varium	Mosses	Grimmiaceae	related to endochitinase 2 precursor	9,73557E-76	63.44	IPR001223 (PFAM)
scaffold-RGKI-2011057-Leucobryum glaucum	Mosses	Leucobryaceae	acidic mammalian chitinase-like	5,10235E-127	63.31	IPR001223 (PFAM)
scaffold-RTMU-2003364-Calypogeia fissa	Liverworts	Calypogeiaceae	acidic mammalian chitinase-like	1,17979E-109	66.64	IPR001223 (PFAM)
scaffold-RTMU-2006799-Calypogeia fissa	Liverworts	Calypogeiaceae	acidic mammalian chitinase-like	6,42373E-91	68.38	IPR001223 (PFAM)
scaffold-RTMU-2010105-Calypogeia fissa	Liverworts	Calypogeiaceae	glycoside hydrolase family 18	1,04185E-104	82.17	IPR001223 (PFAM)
scaffold-RTMU-2010106-Calypogeia fissa	Liverworts	Calypogeiaceae	glycoside hydrolase family 18	1,04185E-104	82.17	IPR001223 (PFAM)
scaffold-RTMU-2019205-Calypogeia fissa	Liverworts	Calypogeiaceae	chitin recognition protein	1,73439E-88	66.03	IPR001223 (PFAM)
scaffold-RTMU-2143976-Calypogeia fissa	Liverworts	Calypogeiaceae	acidic mammalian chitinase	2,94563E-36	66.37	IPR001223 (PFAM)
scaffold-RTMU-2175404-Calypogeia fissa	Liverworts	Calypogeiaceae	glycosyl hydrolase family 18	6,6491E-79	82.89	IPR001223 (PFAM)
scaffold-RTMU-2175514-Calypogeia fissa	Liverworts	Calypogeiaceae	acidic mammalian chitinase isoform X4	5,65258E-73	71.23	IPR001223 (PFAM)
scaffold-RTMU-2179298-Calypogeia fissa	Liverworts	Calypogeiaceae	acidic mammalian chitinase-like	2,77308E-82	67.06	IPR001223 (PFAM)
scaffold-RTMU-2179446-Calypogeia fissa	Liverworts	Calypogeiaceae	chitinase-3-like protein 1	8,94988E-100	69.82	IPR001223 (PFAM)
scaffold-RTMU-2179721-Calypogeia fissa	Liverworts	Calypogeiaceae	chitinase-like protein Idgf4	1,94877E-124	75.38	IPR001223 (PFAM)
scaffold-RTMU-2182337-Calypogeia fissa	Liverworts	Calypogeiaceae	glycoside hydrolase	4,39345E-104	70.41	IPR001223 (PFAM)
scaffold-RTMU-2183105-Calypogeia fissa	Liverworts	Calypogeiaceae	class V chitinase-like	8,4533E-110	61.16	IPR001223 (PFAM)
scaffold-RXRQ-2121621-Phaeoceros carolinianus-sporophyte	Hornworts	Notothyladaceae	acidic mammalian chitinase-like isoform X1	1,86137E-29	67.61	IPR001223 (PFAM)
scaffold-RXRQ-2133493-Phaeoceros carolinianus-sporophyte	Hornworts	Notothyladaceae	glycoside hydrolase family 18	1,3225E-145	83.4	IPR001223 (PFAM)
scaffold-RXRQ-2133873-Phaeoceros carolinianus-sporophyte	Hornworts	Notothyladaceae	endochitinase 42	1,11909E-116	84.86	IPR001223 (PFAM)
scaffold-RXRQ-2137557-Phaeoceros carolinianus-sporophyte	Hornworts	Notothyladaceae	chitinase-like protein CG5210	9,53502E-124	89.95	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-RXRQ-2137809-Phaeoceros_carolinianus-sporophyte	Hornworts	Notothyladaceae	chitinase-like protein Idgf4	7,23016E-53	94.66	IPR001223 (PFAM)
scaffold-RXRQ-2139890-Phaeoceros_carolinianus-sporophyte	Mosses	Takakiaceae	class V chitinase-like	2,86542E-80	61.24	IPR001223 (PFAM)
scaffold-SKQD-2073562-Takakia_lepidozioides	Mosses	Takakiaceae	class V chitinase	1,21297E-77	66.48	IPR001223 (PFAM)
scaffold-SKQD-2079574-Takakia_lepidozioides	Mosses	Takakiaceae	acidic mammalian chitinase	6,88966E-137	63.61	IPR001223 (PFAM)
scaffold-SZYG-2012109-Polytrichum_commune-gametophyte	Mosses	Polytrichaceae	class V chitinase-like	1,68946E-116	60.85	IPR001223 (PFAM)
scaffold-SZYG-2040465-Polytrichum_commune-gametophyte	Mosses	Polytrichaceae	acidic mammalian chitinase-like	5,72987E-54	66.79	IPR001223 (PFAM)
scaffold-TAVP-2019781-Calliergon_cordifolium	Mosses	Calliergonaceae	acidic mammalian chitinase-like	1,17084E-164	62.68	IPR001223 (PFAM)
scaffold-TCBC-2013303-Megaceros_vincentianus	Hornworts	Dendrocerotaceae	class V chitinase-like	1,30311E-45	65.2	IPR001223 (PFAM)
scaffold-TFDQ-2006119-Monoclea_gottschei	Liverworts	Monocleaceae	related to endochitinase 2 precursor	1,2314E-98	73.44	IPR001223 (PFAM)
scaffold-TFDQ-2008991-Monoclea_gottschei	Liverworts	Monocleaceae	glycosyl hydrolase family 18	1,10586E-55	69.44	IPR001223 (PFAM)
scaffold-TFDQ-2009959-Monoclea_gottschei	Liverworts	Monocleaceae	glycoside hydrolase family 18	4,19293E-65	79.15	IPR001223 (PFAM)
scaffold-TFDQ-2120835-Monoclea_gottschei	Liverworts	Monocleaceae	Endochitinase 1	0.0	92.51	IPR001223 (PFAM)
scaffold-TFDQ-2120916-Monoclea_gottschei	Liverworts	Monocleaceae	Chitinase	1,16384E-147	87.77	IPR001223 (PFAM)
scaffold-TFDQ-2121018-Monoclea_gottschei	Liverworts	Monocleaceae	related to chitinase	2,88006E-159	93.81	IPR001223 (PFAM)
scaffold-TFDQ-2123325-Monoclea_gottschei	Liverworts	Monocleaceae	related to RF2 protein	2,4972E-131	83.2	IPR001223 (PFAM)
scaffold-TFYI-2000164-Marchantia_emarginata	Liverworts	Marchantiaceae	chitinase-3-like protein 1	2,72805E-55	63.4	IPR001223 (PFAM)
scaffold-TFYI-2002021-Marchantia_emarginata	Liverworts	Marchantiaceae	acidic endochitinase	6,06757E-55	79.36	IPR001223 (PFAM)
scaffold-TFYI-2002959-Marchantia_emarginata	Liverworts	Marchantiaceae	hevamine-A-like	1,24185E-125	74.5	IPR001223 (PFAM)
scaffold-TFYI-2066800-Marchantia_emarginata	Liverworts	Marchantiaceae	acidic mammalian chitinase-like	6,07755E-72	72.82	IPR001223 (PFAM)
scaffold-TFYI-2066998-Marchantia_emarginata	Liverworts	Marchantiaceae	acidic endochitinase	3,08156E-69	80.8	IPR001223 (PFAM)
scaffold-TFYI-2068733-Marchantia_emarginata	Liverworts	Marchantiaceae	Acidic endochitinase	8,881E-44	73.83	IPR001223 (PFAM)
scaffold-TFYI-2073368-Marchantia_emarginata	Liverworts	Marchantiaceae	acidic endochitinase	5,49591E-115	71.76	IPR001223 (PFAM)
scaffold-TFYI-2073779-Marchantia_emarginata	Liverworts	Marchantiaceae	hevamine-A-like	1,27042E-89	68.23	IPR001223 (PFAM)
scaffold-TGKW-2013377-Frullania	Liverworts	Frullaniaceae	acidic mammalian chitinase-like	5,43486E-122	57.48	IPR001223 (PFAM)
scaffold-TGKW-2013378-Frullania	Liverworts	Frullaniaceae	acidic mammalian chitinase-like	5,43486E-122	57.48	IPR001223 (PFAM)
scaffold-TGKW-2125559-Frullania	Liverworts	Frullaniaceae	acidic mammalian chitinase-like	1,86899E-39	71.31	IPR001223 (PFAM)
scaffold-TMAJ-2010269-Neckera_douglasii	Mosses	Neckeraceae	class V chitinase-like	1,41027E-171	63.33	IPR001223 (PFAM)
scaffold-TMAJ-2010270-Neckera_douglasii	Mosses	Neckeraceae	class V chitinase-like	1,41027E-171	63.33	IPR001223 (PFAM)
scaffold-TMAJ-2010271-Neckera_douglasii	Mosses	Neckeraceae	class V chitinase-like	1,41027E-171	63.33	IPR001223 (PFAM)
scaffold-TMAJ-2021574-Neckera_douglasii	Mosses	Neckeraceae	family 18 glycoside hydrolase	2,44549E-45	62.72	IPR001223 (PFAM)
scaffold-TMAJ-2023280-Neckera_douglasii	Mosses	Neckeraceae	class V chitinase-like	3,54382E-83	62.71	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-TMAJ-2166782-Neckera_douglasii	Mosses	Neckeraceae	glycoside hydrolase	3,73592E-35	65.55	IPR001223 (PFAM)
scaffold-TMAJ-2174094-Neckera_douglasii	Mosses	Neckeraceae	glycoside hydrolase family 18	4,96823E-64	78.0	IPR001223 (PFAM)
scaffold-TWUW-2049624-Anthoceros_agrestis-A	Hornworts	Anthoceroceae	acidic mammalian chitinase-like	5,54997E-70	65.44	IPR001223 (PFAM)
scaffold-TXVB-2013493-Lunularia_cruciata	Liverworts	Lunulariaceae	Acidic endochitinase	4,32158E-132	71.89	IPR001223 (PFAM)
scaffold-TXVB-2063412-Lunularia_cruciata	Liverworts	Lunulariaceae	acidic endochitinase	4,40415E-35	74.58	IPR001223 (PFAM)
scaffold-TXVB-2070336-Lunularia_cruciata	Liverworts	Lunulariaceae	acidic endochitinase	1,61757E-68	75.65	IPR001223 (PFAM)
scaffold-TXVB-2072783-Lunularia_cruciata	Liverworts	Lunulariaceae	chitinase family protein	5,44521E-92	67.78	IPR001223 (PFAM)
scaffold-TXVB-2074185-Lunularia_cruciata	Liverworts	Lunulariaceae	acidic mammalian chitinase-like	1,77591E-143	61.89	IPR001223 (PFAM)
scaffold-UCRN-2052015-Megaceros_tosanus	Hornworts	Dendrocerotaceae	class V chitinase-like	1,85221E-95	64.11	IPR001223 (PFAM)
scaffold-UUHD-2149435-Porella_pinnata	Liverworts	Porellaceae	acidic mammalian chitinase-like	1,62164E-125	61.68	IPR001223 (PFAM)
scaffold-VBMM-2011897-Anomodon_rostratus	Mosses	Brachytheciaceae	class V chitinase-like	7,63365E-169	62.94	IPR001223 (PFAM)
scaffold-VMXJ-2127708-Leucobryum_albidum	Mosses	Leucobryaceae	acidic mammalian chitinase-like	1,09517E-101	65.44	IPR001223 (PFAM)
scaffold-WCZB-2004735-Phaeoceros_carolinianus-gametophyte	Hornworts	Notothyladaceae	glycoside hydrolase family 18 protein	1,29928E-119	80.06	IPR001223 (PFAM)
scaffold-WCZB-2004736-Phaeoceros_carolinianus-gametophyte	Hornworts	Notothyladaceae	glycoside hydrolase family 18 protein	1,29928E-119	80.06	IPR001223 (PFAM)
scaffold-WCZB-2104761-Phaeoceros_carolinianus-gametophyte	Hornworts	Notothyladaceae	AChain A	3,27928E-68	96.78	IPR001223 (PFAM)
scaffold-WCZB-2114497-Phaeoceros_carolinianus-gametophyte	Hornworts	Notothyladaceae	chitinase-like protein Idgf2	6,73286E-43	87.2	IPR001223 (PFAM)
scaffold-WCZB-2116905-Phaeoceros_carolinianus-gametophyte	Hornworts	Notothyladaceae	chitinase-like protein Idgf4	7,23016E-53	94.66	IPR001223 (PFAM)
scaffold-WCZB-2116970-Phaeoceros_carolinianus-gametophyte	Hornworts	Notothyladaceae	chitinase-3-like protein 1	6,12448E-87	63.7	IPR001223 (PFAM)
scaffold-WCZB-2119571-Phaeoceros_carolinianus-gametophyte	Hornworts	Notothyladaceae	class V chitinase-like	3,01785E-81	62.26	IPR001223 (PFAM)
scaffold-WEEQ-2089607-Phaeoceros_carolinianus-gametophyte 1575	Hornworts	Notothyladaceae	class V chitinase-like	4,39733E-84	60.93	IPR001223 (PFAM)
scaffold-WJLO-2007388-Riccia_berychiana	Liverworts	Ricciaceae	Acidic endochitinase	6,85753E-112	70.28	IPR001223 (PFAM)
scaffold-WJLO-2007710-Riccia_berychiana	Liverworts	Ricciaceae	acidic mammalian chitinase-like	3,69849E-171	63.68	IPR001223 (PFAM)
scaffold-WJLO-2035839-Riccia_berychiana	Liverworts	Ricciaceae	acidic endochitinase-like	2,31588E-136	74.4	IPR001223 (PFAM)
scaffold-WNGH-2008645-Aulacomnium_heterostichum	Mosses	Aulacomniaceae	class V chitinase-like	1,03798E-171	62.82	IPR001223 (PFAM)
scaffold-WNGH-2008646-Aulacomnium_heterostichum	Mosses	Aulacomniaceae	class V chitinase-like	2,00165E-66	63.09	IPR001223 (PFAM)
scaffold-WNGH-2014912-Aulacomnium_heterostichum	Mosses	Aulacomniaceae	putative Chitinase	1,28673E-63	60.72	IPR001223 (PFAM)
scaffold-WNGH-2088662-Aulacomnium_heterostichum	Mosses	Aulacomniaceae	glycoside hydrolase	6,85689E-75	63.46	IPR001223 (PFAM)
scaffold-WSPM-2000719-Rhytidiadelphus_loreus	Mosses	Hylocomiaceae	class V chitinase-like	7,8804E-118	65.49	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-WSPM-2000720-Rhytidiadelphus loreus	Mosses	Hylocomiaceae	class V chitinase-like	2,97353E-69	69.62	IPR001223 (PFAM)
scaffold-WZYK-2086114-Bazzania trilobata	Liverworts	Lepidoziaceae	acidic mammalian chitinase-like	5,46974E-67	65.81	IPR001223 (PFAM)
scaffold-XWHK-2008595-Funaria	Mosses	Bryaceae	acidic mammalian chitinase-like	1,84147E-119	67.12	IPR001223 (PFAM)
scaffold-XWHK-2045122-Funaria	Mosses	Bryaceae	acidic mammalian chitinase-like	4,20745E-65	66.69	IPR001223 (PFAM)
scaffold-YBQN-2014678-Odontoschisma prostratum	Liverworts	Cephaloziaceae	glycoside hydrolase family 18	2,40124E-108	63.95	IPR001223 (PFAM)
scaffold-YBQN-2119580-Odontoschisma prostratum	Liverworts	Cephaloziaceae	probable chitinase 3	1,33636E-19	49.95	IPR001223 (PFAM)
scaffold-YBQN-2120139-Odontoschisma prostratum	Liverworts	Cephaloziaceae	putative chitinase 3	3,97672E-26	62.49	IPR001223 (PFAM)
scaffold-YBQN-2120337-Odontoschisma prostratum	Liverworts	Cephaloziaceae	acidic mammalian chitinase-like	2,74304E-15	52.44	IPR001223 (PFAM)
scaffold-YBQN-2121296-Odontoschisma prostratum	Liverworts	Cephaloziaceae	glycoside hydrolase family 18	1,98189E-98	84.9	IPR001223 (PFAM)
scaffold-YBQN-2123041-Odontoschisma prostratum	Liverworts	Cephaloziaceae	glycoside hydrolase family 18	3,86459E-79	84.15	IPR001223 (PFAM)
scaffold-YEPO-2005117-cf. Physcomitrium sp	Mosses	Funariaceae	oviductal secretory glycoprotein	7,16217E-129	54.3	IPR001223 (PFAM)
scaffold-YEPO-2005679-cf. Physcomitrium sp	Mosses	Funariaceae	class V chitinase	0.0	64.27	IPR001223 (PFAM)
scaffold-YWNF-2048821-Hedwigia ciliata	Mosses	Hedwigiaceae	glycoside hydrolase family 18	5,44232E-35	69.65	IPR001223 (PFAM)
scaffold-YWNF-2051820-Hedwigia ciliata	Mosses	Hedwigiaceae	class V chitinase	7,40555E-170	62.66	IPR001223 (PFAM)
scaffold-ZACW-2001860-Leucodon sciuroides	Mosses	Leucodontaceae	acidic mammalian chitinase-like	2,2733E-173	62.5	IPR001223 (PFAM)
scaffold-ZFRE-2107597-Phaeoceros carolinianus-sporophyte_1575	Hornworts	Notothyladaceae	acidic mammalian chitinase-like	3,53694E-42	66.32	IPR001223 (PFAM)
scaffold-ZQRI-2018149-Timmia austriaca	Mosses	Timmiaceae	acidic mammalian chitinase	2,85841E-146	61.03	IPR001223 (PFAM)
scaffold-ZTHV-2012050-Atrichum angustatum	Mosses	Polytrichaceae	Glycosyl hydrolase family protein with chitinase insertion domain-containing protein	3,53703E-65	62.56	IPR001223 (PFAM)
scaffold-ZTHV-2017150-Atrichum angustatum	Mosses	Polytrichaceae	acidic mammalian chitinase-like	1,07991E-139	62.18	IPR001223 (PFAM)
scaffold-ZTHV-2070247-Atrichum angustatum	Mosses	Polytrichaceae	chitinase-3-like protein 1	8,77938E-33	69.83	IPR001223 (PFAM)
scaffold-ZTHV-2075062-Atrichum angustatum	Mosses	Polytrichaceae	class V chitinase-like	9,85696E-32	61.28	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-ABIJ-2006436-Selaginella lepidophylla	Lycophytes	Selaginellaceae	acidic endochitinase	7,37915E-133	81.16	IPR001223 (PFAM)
scaffold-ABIJ-2008792-Selaginella lepidophylla	Lycophytes	Selaginellaceae	class V chitinase	2,20284E-44	73.88	IPR001223 (PFAM)
scaffold-ABIJ-2008890-Selaginella lepidophylla	Lycophytes	Selaginellaceae	class V chitinase-like	1,47507E-107	68.38	IPR001223 (PFAM)
scaffold-ABIJ-2009255-Selaginella lepidophylla	Lycophytes	Selaginellaceae	acidic endochitinase	2,06505E-129	81.3	IPR001223 (PFAM)
scaffold-ABIJ-2038774-Selaginella lepidophylla	Lycophytes	Selaginellaceae	acidic endochitinase	2,56057E-127	79.24	IPR001223 (PFAM)
scaffold-ABIJ-2038934-Selaginella lepidophylla	Lycophytes	Selaginellaceae	class V chitinase-like	3,18407E-174	67.99	IPR001223 (PFAM)
scaffold-ALVQ-2001326-Tmesipteris parva	Eusporangiate Monilophytes	Psilotaceae	acidic mammalian chitinase-like	2,48305E-06	64.34	IPR001223 (PFAM)

PTERIDOPHYTES – GH18						
SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-ALVQ-2001327-Tmesipteris_parva	Eusporangiate Monilophytes	Psilotaceae	acidic mammalian chitinase-like	3,47019E-36	58.48	IPR001223 (PFAM)
scaffold-ALVQ-2016777-Tmesipteris_parva	Eusporangiate Monilophytes	Psilotaceae	class V chitinase	6,52882E-78	63.56	IPR001223 (PFAM)
scaffold-BEGM-2017282-Botrypus_virginianus	Eusporangiate Monilophytes	Ophioglossaceae	class V chitinase	5,28469E-36	60.59	IPR001223 (PFAM)
scaffold-BEGM-2102286-Botrypus_virginianus	Eusporangiate Monilophytes	Ophioglossaceae	acidic endochitinase-like	4,10295E-113	76.97	IPR001223 (PFAM)
scaffold-BEGM-2102306-Botrypus_virginianus	Eusporangiate Monilophytes	Ophioglossaceae	acidic mammalian chitinase-like	1,61725E-74	61.74	IPR001223 (PFAM)
scaffold-CAPN-2006447-Equisetum_diffusum	Eusporangiate Monilophytes	Equisetaceae	chitotriosidase-1	6,87591E-41	68.83	IPR001223 (PFAM)
scaffold-CAPN-2006448-Equisetum_diffusum	Eusporangiate Monilophytes	Equisetaceae	class V chitinase	1,36146E-34	69.07	IPR001223 (PFAM)
scaffold-CAPN-2009848-Equisetum_diffusum	Eusporangiate Monilophytes	Equisetaceae	class V chitinase	3,15691E-81	61.44	IPR001223 (PFAM)
scaffold-CAPN-2037054-Equisetum_diffusum	Eusporangiate Monilophytes	Equisetaceae	acidic mammalian chitinase-like	2,46257E-12	54.61	IPR001223 (PFAM)
scaffold-CAPN-2037262-Equisetum_diffusum	Eusporangiate Monilophytes	Equisetaceae	AChain A	2,51567E-12	61.46	IPR001223 (PFAM)
scaffold-CAPN-2037836-Equisetum_diffusum	Eusporangiate Monilophytes	Equisetaceae	acidic mammalian chitinase-like	1,12237E-23	59.72	IPR001223 (PFAM)
scaffold-CAPN-2038165-Equisetum_diffusum	Eusporangiate Monilophytes	Equisetaceae	class V chitinase-like	2,25621E-31	59.92	IPR001223 (PFAM)
scaffold-CAPN-2039382-Equisetum_diffusum	Eusporangiate Monilophytes	Equisetaceae	Acidic endochitinase	2,18968E-101	73.68	IPR001223 (PFAM)
scaffold-CAPN-2040486-Equisetum_diffusum	Eusporangiate Monilophytes	Equisetaceae	class V chitinase	1,03037E-75	63.15	IPR001223 (PFAM)
scaffold-CBAE-2001838-Huperzia_myrsinites	Lycophytes	Huperziaceae	acidic endochitinase-like	2,05668E-41	69.81	IPR001223 (PFAM)
scaffold-CBAE-2001839-Huperzia_myrsinites	Lycophytes	Huperziaceae	acidic endochitinase	2,51333E-36	74.34	IPR001223 (PFAM)
scaffold-CBAE-2005523-Huperzia_myrsinites	Lycophytes	Huperziaceae	acidic endochitinase	5,70093E-79	66.78	IPR001223 (PFAM)
scaffold-CBAE-2005524-Huperzia_myrsinites	Lycophytes	Huperziaceae	acidic endochitinase	1,00309E-88	71.26	IPR001223 (PFAM)
scaffold-CBAE-2051921-Huperzia_myrsinites	Lycophytes	Huperziaceae	acidic mammalian chitinase-like	1,46167E-45	70.45	IPR001223 (PFAM)
scaffold-CBAE-2060579-Huperzia_myrsinites	Lycophytes	Huperziaceae	class V chitinase	2,92073E-110	66.64	IPR001223 (PFAM)
scaffold-DFHO-2001788-Danaea_nodosa	Eusporangiate Monilophytes	Marattiaceae	acidic mammalian chitinase-like	4,10254E-29	76.1	IPR001223 (PFAM)
scaffold-DFHO-2007268-Danaea_nodosa	Eusporangiate Monilophytes	Marattiaceae	acidic endochitinase	1,63091E-99	72.69	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-DFHO-2044420-Danaea_nodosa	Eusporangiate Monilophytes	Marattiaceae	acidic mammalian chitinase-like	1,47876E-35	70.86	IPR001223 (PFAM)
scaffold-DFHO-2055341-Danaea_nodosa	Eusporangiate Monilophytes	Marattiaceae	class V chitinase-like	1,20678E-91	63.44	IPR001223 (PFAM)
scaffold-EEAQ-2005608-Sceptridium_dissectum	Eusporangiate Monilophytes	Ophioglossaceae	class V chitinase	1,3358E-88	61.98	IPR001223 (PFAM)
scaffold-EEAQ-2005609-Sceptridium_dissectum	Eusporangiate Monilophytes	Ophioglossaceae	class V chitinase	1,3358E-88	61.98	IPR001223 (PFAM)
scaffold-EEAQ-2008990-Sceptridium_dissectum	Eusporangiate Monilophytes	Ophioglossaceae	acidic endochitinase	1,45893E-59	72.41	IPR001223 (PFAM)
scaffold-EEAQ-2008991-Sceptridium_dissectum	Eusporangiate Monilophytes	Ophioglossaceae	acidic endochitinase	4,58592E-44	71.83	IPR001223 (PFAM)
scaffold-EEAQ-2008992-Sceptridium_dissectum	Eusporangiate Monilophytes	Ophioglossaceae	Acidic endochitinase	9,00319E-71	76.08	IPR001223 (PFAM)
scaffold-EEAQ-2082693-Sceptridium_dissectum	Eusporangiate Monilophytes	Ophioglossaceae	acidic mammalian chitinase-like	1,05958E-15	59.68	IPR001223 (PFAM)
scaffold-EEAQ-2086905-Sceptridium_dissectum	Eusporangiate Monilophytes	Ophioglossaceae	class V chitinase	3,78098E-87	62.0	IPR001223 (PFAM)
scaffold-ENQF-2003741-Lycopodium_annotinum	Lycophytes	Lycopodiaceae	class V chitinase	5,81446E-96	69.33	IPR001223 (PFAM)
scaffold-ENQF-2005526-Lycopodium_annotinum	Lycophytes	Lycopodiaceae	Acidic endochitinase	4,58236E-102	7348	IPR001223 (PFAM)
scaffold-ENQF-2012682-Lycopodium_annotinum	Lycophytes	Lycopodiaceae	class V chitinase	4,39319E-109	67.04	IPR001223 (PFAM)
scaffold-ENQF-2015939-Lycopodium_annotinum	Lycophytes	Lycopodiaceae	class V chitinase-like	1,78278E-63	68.18	IPR001223 (PFAM)
scaffold-ENQF-2017435-Lycopodium_annotinum	Lycophytes	Lycopodiaceae	class V chitinase	2,11436E-90	66.8	IPR001223 (PFAM)
scaffold-GAON-2001240-Huperzia_squarrosa	Lycophytes	Lycopodiaceae	acidic endochitinase	2,47711E-42	79.16	IPR001223 (PFAM)
scaffold-GAON-2001241-Huperzia_squarrosa	Lycophytes	Lycopodiaceae	acidic endochitinase	8,43922E-43	74.67	IPR001223 (PFAM)
scaffold-GAON-2007131-Huperzia_squarrosa	Lycophytes	Lycopodiaceae	acidic endochitinase	7,98915E-88	71.16	IPR001223 (PFAM)
scaffold-GAON-2016742-Huperzia_squarrosa	Lycophytes	Lycopodiaceae	class V chitinase	8,04208E-104	66.99	IPR001223 (PFAM)
scaffold-GAON-2055613-Huperzia_squarrosa	Lycophytes	Lycopodiaceae	class V chitinase	6,34137E-63	69.89	IPR001223 (PFAM)
scaffold-GKAG-2011802-Huperzia_lucidula	Lycophytes	Lycopodiaceae	acidic endochitinase	2,93548E-97	72.27	IPR001223 (PFAM)
scaffold-GKAG-2011803-Huperzia_lucidula	Lycophytes	Lycopodiaceae	acidic endochitinase	9,6752E-98	73.45	IPR001223 (PFAM)
scaffold-GKAG-2017355-Huperzia_lucidula	Lycophytes	Lycopodiaceae	class V chitinase	7,49927E-101	64.52	IPR001223 (PFAM)
scaffold-GKAG-2022275-Huperzia_lucidula	Lycophytes	Lycopodiaceae	class V chitinase	1,66475E-96	68.81	IPR001223 (PFAM)
scaffold-GKAG-2097278-Huperzia_lucidula	Lycophytes	Lycopodiaceae	bacteriodes thetaiotaomicron symbiotic chitinase	1,9125E-131	64.36	IPR001223 (PFAM)
scaffold-GTUO-2001847-Huperzia_selago	Lycophytes	Lycopodiaceae	acidic endochitinase-like	9,47804E-34	74.35	IPR001223 (PFAM)
scaffold-GTUO-2025296-Huperzia_selago	Lycophytes	Lycopodiaceae	acidic mammalian chitinase	7,38789E-103	66.63	IPR001223 (PFAM)
scaffold-GTUO-2026196-Huperzia_selago	Lycophytes	Lycopodiaceae	class V chitinase	1,07813E-57	69.46	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-JKAA-2002507-Selaginella_wallacei	Lycophytes	Selaginellaceae	class V chitinase-like	1,00998E-95	621.4	IPR001223 (PFAM)
scaffold-JKAA-2002508-Selaginella_wallacei	Lycophytes	Selaginellaceae	class V chitinase-like	5,93482E-83	65.9	IPR001223 (PFAM)
scaffold-JKAA-2008532-Selaginella_wallacei	Lycophytes	Selaginellaceae	acidic mammalian chitinase-like	5,55276E-18	61.15	IPR001223 (PFAM)
scaffold-JKAA-2013110-Selaginella_wallacei	Lycophytes	Selaginellaceae	acidic endochitinase	8,65916E-94	77.2	IPR001223 (PFAM)
scaffold-JKAA-2158634-Selaginella_wallacei	Lycophytes	Selaginellaceae	class V chitinase	5,47248E-36	65.98	IPR001223 (PFAM)
scaffold-JKAA-2159729-Selaginella_wallacei	Lycophytes	Selaginellaceae	putative class V chitinase	5,17795E-37	77.41	IPR001223 (PFAM)
scaffold-JKAA-2169170-Selaginella_wallacei	Lycophytes	Selaginellaceae	glycoside hydrolase	1,65812E-39	73.95	IPR001223 (PFAM)
scaffold-JKAA-2173170-Selaginella_wallacei	Lycophytes	Selaginellaceae	glycoside hydrolase family 18	1,39492E-60	89.3	IPR001223 (PFAM)
scaffold-JKAA-2174276-Selaginella_wallacei	Lycophytes	Selaginellaceae	glycoside hydrolase family 18	5,40609E-32	63.2	IPR001223 (PFAM)
scaffold-JKAA-2177540-Selaginella_wallacei	Lycophytes	Selaginellaceae	acidic endochitinase-like	1,42583E-123	77.19	IPR001223 (PFAM)
scaffold-JKAA-2178165-Selaginella_wallacei	Lycophytes	Selaginellaceae	glycoside hydrolase	1,57212E-70	83.97	IPR001223 (PFAM)
scaffold-JKAA-2178880-Selaginella_wallacei	Lycophytes	Selaginellaceae	acidic endochitinase	7,27741E-115	78.35	IPR001223 (PFAM)
scaffold-JKAA-2180047-Selaginella_wallacei	Lycophytes	Selaginellaceae	glycoside hydrolase family 18	1,29131E-143	85.45	IPR001223 (PFAM)
scaffold-JKAA-2180911-Selaginella_wallacei	Lycophytes	Selaginellaceae	glycoside hydrolase	7,29076E-72	63.35	IPR001223 (PFAM)
scaffold-JKAA-2181250-Selaginella_wallacei	Lycophytes	Selaginellaceae	glycoside hydrolase	1,30916E-69	64.19	IPR001223 (PFAM)
scaffold-JVSZ-2002787-Equisetum_hymale	Eusporangiate Monilophytes	Equisetaceae	acidic mammalian chitinase-like	2,55616E-78	61.42	IPR001223 (PFAM)
scaffold-JVSZ-2002788-Equisetum_hymale	Eusporangiate Monilophytes	Equisetaceae	class V chitinase	7,93305E-78	61.15	IPR001223 (PFAM)
scaffold-JVSZ-2002789-Equisetum_hymale	Eusporangiate Monilophytes	Equisetaceae	class V chitinase-like	1,12384E-09	64.97	IPR001223 (PFAM)
scaffold-JVSZ-2119489-Equisetum_hymale	Eusporangiate Monilophytes	Equisetaceae	AChain A	5,8541E-15	59.24	IPR001223 (PFAM)
scaffold-KJYC-2002529-Selaginella_willdenowii	Lycophytes	Selaginellaceae	class V chitinase	5,52651E-46	72.41	IPR001223 (PFAM)
scaffold-KJYC-2006273-Selaginella_willdenowii	Lycophytes	Selaginellaceae	class V chitinase	2,39536E-46	69.56	IPR001223 (PFAM)
scaffold-KJYC-2006274-Selaginella_willdenowii	Lycophytes	Selaginellaceae	class V chitinase	2,39536E-46	69.56	IPR001223 (PFAM)
scaffold-KJYC-2007922-Selaginella_willdenowii	Lycophytes	Selaginellaceae	class V chitinase	5,31759E-65	71.76	IPR001223 (PFAM)
scaffold-KJYC-2013372-Selaginella_willdenowii	Lycophytes	Selaginellaceae	acidic endochitinase	1,28264E-86	67.36	IPR001223 (PFAM)
scaffold-KJYC-2044359-Selaginella_willdenowii	Lycophytes	Selaginellaceae	acidic mammalian chitinase-like	1,35468E-36	68.81	IPR001223 (PFAM)
scaffold-KJYC-2047116-Selaginella_willdenowii	Lycophytes	Selaginellaceae	acidic endochitinase	1,38735E-56	77.88	IPR001223 (PFAM)
scaffold-KJYC-2047493-Selaginella_willdenowii	Lycophytes	Selaginellaceae	acidic endochitinase	2,15134E-62	78.68	IPR001223 (PFAM)
scaffold-KJYC-2048999-Selaginella_willdenowii	Lycophytes	Selaginellaceae	class V chitinase-like	4,43578E-93	70.79	IPR001223 (PFAM)
scaffold-KUXM-2001058-Selaginella_selaginoides	Lycophytes	Selaginellaceae	class V chitinase-like	1,09246E-13	67.59	IPR001223 (PFAM)
scaffold-KUXM-2009202-Selaginella_selaginoides	Lycophytes	Selaginellaceae	class V chitinase	2,17918E-48	63.68	IPR001223 (PFAM)
scaffold-KUXM-2043003-Selaginella_selaginoides	Lycophytes	Selaginellaceae	acidic endochitinase	6,68209E-136	77.97	IPR001223 (PFAM)
scaffold-LGDQ-2008640-Selaginella_apoda	Lycophytes	Selaginellaceae	class V chitinase-like	0.0	69.48	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-LGDQ-2008641-Selaginella_apoda	Lycophytes	Selaginellaceae	class V chitinase-like	0.0	69.48	IPR001223 (PFAM)
scaffold-LGDQ-2011793-Selaginella_apoda	Lycophytes	Selaginellaceae	acidic mammalian chitinase	6,76004E-158	65.8	IPR001223 (PFAM)
scaffold-LGDQ-2040164-Selaginella_apoda	Lycophytes	Selaginellaceae	acidic endochitinase	3,67656E-82	80.34	IPR001223 (PFAM)
scaffold-LGDQ-2041235-Selaginella_apoda	Lycophytes	Selaginellaceae	acidic endochitinase	1,43414E-78	77.87	IPR001223 (PFAM)
scaffold-NYBX-2005847-Huperzia_selago-A	Lycophytes	Lycopodiaceae	chitinase-3-like protein 1	5,43986E-38	52.6	IPR001223 (PFAM)
scaffold-NYBX-2006529-Huperzia_selago-A	Lycophytes	Lycopodiaceae	acidic endochitinase	1,02835E-101	74.97	IPR001223 (PFAM)
scaffold-NYBX-2006530-Huperzia_selago-A	Lycophytes	Lycopodiaceae	acidic endochitinase	1,02835E-101	74.97	IPR001223 (PFAM)
scaffold-NYBX-2006531-Huperzia_selago-A	Lycophytes	Lycopodiaceae	acidic endochitinase	1,02835E-101	74.97	IPR001223 (PFAM)
scaffold-NYBX-2021614-Huperzia_selago-A	Lycophytes	Lycopodiaceae	chitinase-3-like protein 1	1,62726E-126	67.53	IPR001223 (PFAM)
scaffold-NYBX-2021615-Huperzia_selago-A	Lycophytes	Lycopodiaceae	chitinase-3-like protein 1	8,32011E-105	68.91	IPR001223 (PFAM)
scaffold-NYBX-2023892-Huperzia_selago-A	Lycophytes	Lycopodiaceae	class V chitinase	3,84357E-42	64.89	IPR001223 (PFAM)
scaffold-NYBX-2120238-Huperzia_selago-A	Lycophytes	Lycopodiaceae	probable chitinase 10	2,3095E-27	83.1	IPR001223 (PFAM)
scaffold-NYBX-2126520-Huperzia_selago-A	Lycophytes	Lycopodiaceae	probable chitinase 10	2,91486E-77	75.64	IPR001223 (PFAM)
scaffold-NYBX-2129604-Huperzia_selago-A	Lycophytes	Lycopodiaceae	class V chitinase-like	4,85702E-71	71.44	IPR001223 (PFAM)
scaffold-NYBX-2130744-Huperzia_selago-A	Lycophytes	Lycopodiaceae	chitinase-3-like protein 1	6,06156E-143	70.8	IPR001223 (PFAM)
scaffold-PKOX-2000311-Isoetes_tetragyniformans	Lycophytes	Isoetaceae	acidic endochitinase	1,69475E-100	78.92	IPR001223 (PFAM)
scaffold-NWWI-2025427-Nephrolepis_exaltata	Leptosporangiate Monilophytes	Lomariopsidaceae	chitotriosidase-1-like	3,83769E-66	62.39	IPR001223 (PFAM)
scaffold-NWWI-2025428-Nephrolepis_exaltata	Leptosporangiate Monilophytes	Lomariopsidaceae	acidic mammalian chitinase-like	3,37917E-66	62.86	IPR001223 (PFAM)
scaffold-NWWI-2005288-Nephrolepis_exaltata	Leptosporangiate Monilophytes	Lomariopsidaceae	class IV chitinase	1,4924E-77	67.87	IPR000726 (PFAM)
scaffold-NWWI-2005289-Nephrolepis_exaltata	Leptosporangiate Monilophytes	Lomariopsidaceae	class IV chitinase	1,81351E-78	67.22	IPR000726 (PFAM)
scaffold-NWWI-2005290-Nephrolepis_exaltata	Leptosporangiate Monilophytes	Lomariopsidaceae	class IV chitinase	2,08408E-74	66.77	IPR000726 (PFAM)
scaffold-NWWI-2005291-Nephrolepis_exaltata	Leptosporangiate Monilophytes	Lomariopsidaceae	class IV chitinase	1,14732E-75	69.2	IPR000726 (PFAM)
scaffold-NWWI-2008181-Nephrolepis_exaltata	Leptosporangiate Monilophytes	Lomariopsidaceae	endochitinase EP3-like	1,03823E-68	57.63	IPR000726 (PFAM)
scaffold-NWWI-2008182-Nephrolepis_exaltata	Leptosporangiate Monilophytes	Lomariopsidaceae	putative class I chitinase	1,16154E-50	65.41	IPR000726 (PFAM)
scaffold-NWWI-2008183-Nephrolepis_exaltata	Leptosporangiate Monilophytes	Lomariopsidaceae	endochitinase EP3-like	9,25697E-64	69.17	IPR000726 (PFAM)
scaffold-NWWI-2011801-Nephrolepis_exaltata	Leptosporangiate Monilophytes	Lomariopsidaceae	putative class I chitinase	2,45443E-35	69.66	IPR000726 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-NWWI-2011802-Nephrolepis_exaltata	Leptosporangiate Monilophytes	Lomariopsidaceae	putative class I chitinase	2,45443E-35	69.66	IPR000726 (PFAM)
scaffold-NWWI-2014583-Nephrolepis_exaltata	Leptosporangiate Monilophytes	Lomariopsidaceae	Glycoside hydrolase	1,04812E-27	56.31	IPR000726 (PFAM)
scaffold-NWWI-2126383-Nephrolepis_exaltata	Leptosporangiate Monilophytes	Lomariopsidaceae	endochitinase EP3-like	1,22529E-54	72.41	IPR000726 (PFAM)
scaffold-PKOX-2093371-Isoetes tegetiformans	Lycophytes	Isoetaceae	chitinase-3-like protein 1	1,44362E-35	60.98	IPR001223 (PFAM)
scaffold-PKOX-2097106-Isoetes tegetiformans	Lycophytes	Isoetaceae	class V chitinase	2,30735E-103	67.04	IPR001223 (PFAM)
scaffold-PKOX-2098297-Isoetes tegetiformans	Lycophytes	Isoetaceae	acidic mammalian chitinase	9,02551E-120	68.13	IPR001223 (PFAM)
scaffold-PQTO-2012178-Lycopodium deuterodensum	Lycophytes	Lycopodiaceae	acidic mammalian chitinase-like	2,35836E-112	66.97	IPR001223 (PFAM)
scaffold-PQTO-2076779-Lycopodium deuterodensum	Lycophytes	Lycopodiaceae	class V chitinase	4,35967E-68	67.06	IPR001223 (PFAM)
scaffold-PQTO-2079202-Lycopodium deuterodensum	Lycophytes	Lycopodiaceae	acidic endochitinase	1,68223E-101	73.76	IPR001223 (PFAM)
scaffold-PYHZ-2003251-Isoetes sp	Lycophytes	Isoetaceae	acidic mammalian chitinase-like	1,91385E-125	67.86	IPR001223 (PFAM)
scaffold-PYHZ-2003252-Isoetes sp	Lycophytes	Isoetaceae	class V chitinase	4,08874E-29	71.38	IPR001223 (PFAM)
scaffold-PYHZ-2003253-Isoetes sp	Lycophytes	Isoetaceae	class V chitinase	1,6321E-112	67.3	IPR001223 (PFAM)
scaffold-PYHZ-2004504-Isoetes sp	Lycophytes	Isoetaceae	acidic endochitinase	1,25255E-99	75.15	IPR001223 (PFAM)
scaffold-PYHZ-2007052-Isoetes sp	Lycophytes	Isoetaceae	class V chitinase	2,76274E-78	57.02	IPR001223 (PFAM)
scaffold-PYHZ-2007053-Isoetes sp	Lycophytes	Isoetaceae	class V chitinase	5,77231E-75	58.66	IPR001223 (PFAM)
scaffold-PYHZ-2007870-Isoetes sp	Lycophytes	Isoetaceae	acidic endochitinase	1,99239E-100	77.93	IPR001223 (PFAM)
scaffold-PYHZ-2007871-Isoetes sp	Lycophytes	Isoetaceae	hevamine-A-like	7,80435E-109	78.57	IPR001223 (PFAM)
scaffold-PYHZ-2007872-Isoetes sp	Lycophytes	Isoetaceae	acidic endochitinase	2,79998E-93	78.34	IPR001223 (PFAM)
scaffold-PYHZ-2009287-Isoetes sp	Lycophytes	Isoetaceae	class V chitinase-like	7,33748E-64	58.66	IPR001223 (PFAM)
scaffold-PYHZ-2071770-Isoetes sp	Lycophytes	Isoetaceae	Glycosyl hydrolase family protein with chitinase insertion domain-containing	4,87098E-114	68.97	IPR001223 (PFAM)
scaffold-PYHZ-2073439-Isoetes sp	Lycophytes	Isoetaceae	class V chitinase	5,71777E-103	66.54	IPR001223 (PFAM)
scaffold-PYHZ-2075391-Isoetes sp	Lycophytes	Isoetaceae	Glycosyl hydrolase family protein with chitinase insertion domain-containing	1,82802E-128	66.44	IPR001223 (PFAM)
scaffold-QVMR-2015945-Psilotum nudum	Eusporangiate Monilophytes	Psilotaceae	class V chitinase	1,30096E-43	66.92	IPR001223 (PFAM)
scaffold-QVMR-2017494-Psilotum nudum	Eusporangiate Monilophytes	Psilotaceae	class V chitinase	1,68136E-91	62.54	IPR001223 (PFAM)
scaffold-UGNK-2002386-Marattia attenuata	Eusporangiate Monilophytes	Marattiaceae	class V chitinase-like	6,23691E-97	63.35	IPR001223 (PFAM)
scaffold-UGNK-2002387-Marattia attenuata	Eusporangiate Monilophytes	Marattiaceae	class V chitinase-like	2,0814E-78	65.28	IPR001223 (PFAM)
scaffold-UGNK-2016335-Marattia attenuata	Eusporangiate Monilophytes	Marattiaceae	acidic mammalian chitinase-like	3,21432E-93	64.19	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-UGNK-2019481-Marattia_attenuata	Eusporangiate Monilophytes	Marattiaceae	class V chitinase-like	4,26556E-53	62.32	IPR001223 (PFAM)
scaffold-ULKT-2008784-Lycopodiella_ apressa	Lycophytes	Lycopodiaceae	acidic endochitinase	2,01495E-94	72.16	IPR001223 (PFAM)
scaffold-ULKT-2011128-Lycopodiella_ apressa	Lycophytes	Lycopodiaceae	class V chitinase	1,52648E-71	67.32	IPR001223 (PFAM)
scaffold-ULKT-2011129-Lycopodiella_ apressa	Lycophytes	Lycopodiaceae	acidic mammalian chitinase	1,77605E-111	67.96	IPR001223 (PFAM)
scaffold-ULKT-2015752-Lycopodiella_ apressa	Lycophytes	Lycopodiaceae	acidic endochitinase	7,00327E-107	72.7	IPR001223 (PFAM)
scaffold-ULKT-2016810-Lycopodiella_ apressa	Lycophytes	Lycopodiaceae	class V chitinase	4,36749E-32	73.85	IPR001223 (PFAM)
scaffold-UPMJ-2005627-Pseudolycopodiella_ caroliniana	Lycophytes	Lycopodiaceae	Acidic endochitinase	6,80861E-104	73.56	IPR001223 (PFAM)
scaffold-UPMJ-2009660-Pseudolycopodiella_ caroliniana	Lycophytes	Lycopodiaceae	class V chitinase	7,30261E-75	68.9	IPR001223 (PFAM)
scaffold-UPMJ-2018058-Pseudolycopodiella_ caroliniana	Lycophytes	Lycopodiaceae	acidic endochitinase	1,28806E-105	72.48	IPR001223 (PFAM)
scaffold-UPMJ-2077280-Pseudolycopodiella_ caroliniana	Lycophytes	Lycopodiaceae	acidic endochitinase	2,28796E-24	71.55	IPR001223 (PFAM)
scaffold-UPMJ-2082325-Pseudolycopodiella_ caroliniana	Lycophytes	Lycopodiaceae	class V chitinase	1,0466E-114	68.74	IPR001223 (PFAM)
scaffold-UXCS-2007028-Marattia_sp	Eusporangiate Monilophytes	Marattiaceae	acidic endochitinase	3,11625E-47	71.83	IPR001223 (PFAM)
scaffold-UXCS-2007029-Marattia_sp	Eusporangiate Monilophytes	Marattiaceae	acidic endochitinase	5,46914E-90	75.27	IPR001223 (PFAM)
scaffold-UXCS-2016006-Marattia_sp	Eusporangiate Monilophytes	Marattiaceae	class V chitinase-like	2,84033E-57	63.13	IPR001223 (PFAM)
scaffold-UXCS-2120386-Marattia_sp	Eusporangiate Monilophytes	Marattiaceae	class V chitinase	1,70031E-07	59.85	IPR001223 (PFAM)
scaffold-VVRN-2010946-Lonchitis_hirsuta	Leptosporangiate Monilophytes	Lonchitidaceae	class V chitinase	1,02648E-17	52.99	IPR001223 (PFAM)
scaffold-VVRN-2013897-Lonchitis_hirsuta	Leptosporangiate Monilophytes	Lonchitidaceae	class V chitinase	1,58384E-80	59.8	IPR001223 (PFAM)
scaffold-VVRN-2022226-Lonchitis_hirsuta	Leptosporangiate Monilophytes	Lonchitidaceae	class V chitinase	9,75699E-67	57.1	IPR001223 (PFAM)
scaffold-VVRN-2095349-Lonchitis_hirsuta	Leptosporangiate Monilophytes	Lonchitidaceae	Acidic mammalian chitinase	2,83484E-28	52.52	IPR001223 (PFAM)
scaffold-VVRN-2096647-Lonchitis_hirsuta	Leptosporangiate Monilophytes	Lonchitidaceae	chitotriosidase-1-like isoform X2	1,71199E-70	60.5	IPR001223 (PFAM)
scaffold-WAFT-2001766-Diphasiastrum_ digitatum	Lycophytes	Lycopodiaceae	class V chitinase	1,1233E-105	66.31	IPR001223 (PFAM)
scaffold-WAFT-2002690-Diphasiastrum_ digitatum	Lycophytes	Lycopodiaceae	Acidic endochitinase	1,78929E-93	72.89	IPR001223 (PFAM)
scaffold-WAFT-2014739-Diphasiastrum_ digitatum	Lycophytes	Lycopodiaceae	class V chitinase	1,48598E-120	66.2	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-WAFT-2068592-Diphasiastrum_digitatum	Lycophytes	Lycopodiaceae	class V chitinase	5,66453E-80	65.2	IPR001223 (PFAM)
scaffold-WCLG-2004660-Adiantum_aleuticum	Leptosporangiate Monilophytes	Pteridaceae	acidic mammalian chitinase-like	4,03347E-19	57.62	IPR001223 (PFAM)
scaffold-WGTU-2061504-Leucostegia_immersa	Leptosporangiate Monilophytes	Hypodematiaceae	acidic mammalian chitinase-like isoform X1	3,14229E-15	66.76	IPR001223 (PFAM)
scaffold-WGTU-2061726-Leucostegia_immersa	Leptosporangiate Monilophytes	Hypodematiaceae	class V chitinase	8,15691E-30	62.6	IPR001223 (PFAM)
scaffold-WGTU-2071532-Leucostegia_immersa	Leptosporangiate Monilophytes	Hypodematiaceae	acidic endochitinase	1,28058E-91	72.31	IPR001223 (PFAM)
scaffold-WGTU-2072225-Leucostegia_immersa	Leptosporangiate Monilophytes	Hypodematiaceae	class V chitinase	3,17767E-54	62.4	IPR001223 (PFAM)
scaffold-WQML-2003973-Cryptogramma_acrostichoides	Leptosporangiate Monilophytes	Pteridaceae	acidic endochitinase	2,26435E-87	75.49	IPR001223 (PFAM)
scaffold-WQML-2007180-Cryptogramma_acrostichoides	Leptosporangiate Monilophytes	Pteridaceae	chitotriosidase-1-like	2,56956E-65	59.85	IPR001223 (PFAM)
scaffold-WTJG-2002514-Ophioglossum_vulgatum	Eusporangiate Monilophytes	Ophioglossaceae	class V chitinase	1,03948E-37	59.52	IPR001223 (PFAM)
scaffold-WTJG-2002515-Ophioglossum_vulgatum	Eusporangiate Monilophytes	Ophioglossaceae	class V chitinase	3,84283E-37	59.94	IPR001223 (PFAM)
scaffold-WTJG-2016964-Ophioglossum_vulgatum	Eusporangiate Monilophytes	Ophioglossaceae	class V chitinase-like	5,74628E-31	70.09	IPR001223 (PFAM)
scaffold-WTJG-2095237-Ophioglossum_vulgatum	Eusporangiate Monilophytes	Ophioglossaceae	class V chitinase	3,25002E-53	63.44	IPR001223 (PFAM)
scaffold-XDDT-2003802-Argyrochosma_nivea	Leptosporangiate Monilophytes	Pteridaceae	chitotriosidase-1-like isoform X2	2,59369E-68	59.2	IPR001223 (PFAM)
scaffold-XDDT-2009479-Argyrochosma_nivea	Leptosporangiate Monilophytes	Pteridaceae	acidic endochitinase	1,50649E-100	74.39	IPR001223 (PFAM)
scaffold-XDDT-2009480-Argyrochosma_nivea	Leptosporangiate Monilophytes	Pteridaceae	acidic endochitinase	2,54611E-100	75.56	IPR001223 (PFAM)
scaffold-XDDT-2073722-Argyrochosma_nivea	Leptosporangiate Monilophytes	Pteridaceae	class V chitinase-like	1,46976E-67	63.43	IPR001223 (PFAM)
scaffold-XXNF-2066461-Dendrolycopodium_obscurum	Lycophytes	Lycopodiaceae	class V chitinase-like	1,70055E-63	70.04	IPR001223 (PFAM)
scaffold-XXHP-2001291-Cystopteris_fragilis-Stewart	Leptosporangiate Monilophytes	Cystopteridaceae	acidic endochitinase-like	1,2233E-83	74.28	IPR001223 (PFAM)
scaffold-XXHP-2013082-Cystopteris_fragilis-Stewart	Leptosporangiate Monilophytes	Cystopteridaceae	class V chitinase	1,55666E-59	60.12	IPR001223 (PFAM)
scaffold-XXHP-2015468-Cystopteris_fragilis-Stewart	Leptosporangiate Monilophytes	Cystopteridaceae	chitinase-like protein EN03	6,81729E-151	66.53	IPR001223 (PFAM)

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SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-XXHP-2015710-Cystopteris_fragilis-Stewart	Leptosporangiate Monilophytes	Cystopteridaceae	acidic mammalian chitinase-like	1,00029E-24	48.85	IPR001223 (PFAM)
scaffold-XXHP-2015711-Cystopteris_fragilis-Stewart	Leptosporangiate Monilophytes	Cystopteridaceae	acidic mammalian chitinase-like	1,32015E-24	48.92	IPR001223 (PFAM)
scaffold-XXHP-2080057-Cystopteris_fragilis-Stewart	Leptosporangiate Monilophytes	Cystopteridaceae	class V chitinase	2,18375E-26	65.19	IPR001223 (PFAM)
scaffold-XXHP-2083486-Cystopteris_fragilis-Stewart	Leptosporangiate Monilophytes	Cystopteridaceae	acidic mammalian chitinase-like	3,22284E-32	57.6	IPR001223 (PFAM)
scaffold-YCKE-2009010-Notholaena_montieliae	Leptosporangiate Monilophytes	Pteridaceae	chitotriosidase-1-like	2,68089E-43	68.81	IPR001223 (PFAM)
scaffold-YCKE-2054510-Notholaena_montieliae	Leptosporangiate Monilophytes	Pteridaceae	class V chitinase-like	4,3756E-18	63.5	IPR001223 (PFAM)
scaffold-YCKE-2060473-Notholaena_montieliae	Leptosporangiate Monilophytes	Pteridaceae	acidic endochitinase	2,77449E-65	73.28	IPR001223 (PFAM)
scaffold-YIXP-2008121-Lindsaea_microphylla	Leptosporangiate Monilophytes	Lindsaeaceae	class V chitinase	9,70482E-28	55.15	IPR001223 (PFAM)
scaffold-YJYJ-2011496-Woodsia_scopulina	Leptosporangiate Monilophytes	Woodsiaceae	class V chitinase	1,94271E-80	58.92	IPR001223 (PFAM)
scaffold-YJYJ-2071484-Woodsia_scopulina	Leptosporangiate Monilophytes	Woodsiaceae	class V chitinase	7,33428E-35	69.06	IPR001223 (PFAM)
scaffold-YJYJ-2075390-Woodsia_scopulina	Leptosporangiate Monilophytes	Woodsiaceae	chitotriosidase-1-like	5,81232E-66	60.35	IPR001223 (PFAM)
scaffold-YJYJ-2075638-Woodsia_scopulina	Leptosporangiate Monilophytes	Woodsiaceae	Acidic endochitinase	1,63225E-71	7221	IPR001223 (PFAM)
scaffold-YJYJ-2076688-Woodsia_scopulina	Leptosporangiate Monilophytes	Woodsiaceae	class V chitinase	1,12501E-66	50.03	IPR001223 (PFAM)
scaffold-YLJA-2065344-Polypodium_amorphum	Leptosporangiate Monilophytes	Polypodiaceae	acidic mammalian chitinase	2,60911E-29	66.38	IPR001223 (PFAM)
scaffold-YLJA-2066129-Polypodium_amorphum	Leptosporangiate Monilophytes	Polypodiaceae	acidic endochitinase-like	2,96103E-46	72.11	IPR001223 (PFAM)
scaffold-YLJA-2066868-Polypodium_amorphum	Leptosporangiate Monilophytes	Polypodiaceae	class V chitinase	1,01873E-37	58.89	IPR001223 (PFAM)
scaffold-YOWV-2000206-Cystopteris_protrusa	Leptosporangiate Monilophytes	Cystopteridaceae	class V chitinase	7,95557E-52	59.57	IPR001223 (PFAM)
scaffold-YOWV-2004880-Cystopteris_protrusa	Leptosporangiate Monilophytes	Cystopteridaceae	class V chitinase	5,30312E-31	56.08	IPR001223 (PFAM)
scaffold-YOWV-2008552-Cystopteris_protrusa	Leptosporangiate Monilophytes	Cystopteridaceae	acidic mammalian chitinase-like	1,25271E-20	50.25	IPR001223 (PFAM)
YOWV-2057388-Cystopteris_protrusa	Leptosporangiate Monilophytes	Cystopteridaceae	class V chitinase	3,46563E-12	65.03	IPR001223 (PFAM)

PTERIDOPHYTES – GH18						
SeqName	Phylum	Family	Description	e-Value	Similarity	InterPro GO IDs
scaffold-YOWV-2066886-Cystopteris_protrusa	Leptosporangiate Monilophytes	Cystopteridaceae	acidic endochitinase	1,11201E-81	74.29	IPR001223 (PFAM)
scaffold-YQEC-2002428-Woodsia_ilvensis	Leptosporangiate Monilophytes	Woodsiaceae	Glycosyl hydrolase family protein with chitinase insertion domain-containing	2,21718E-37	69.64	IPR001223 (PFAM)
scaffold-YQEC-2012943-Woodsia_ilvensis	Leptosporangiate Monilophytes	Woodsiaceae	acidic mammalian chitinase-like	2,82211E-41	65.08	IPR001223 (PFAM)
scaffold-YQEC-2074942-Woodsia_ilvensis	Leptosporangiate Monilophytes	Woodsiaceae	class V chitinase	5,55586E-80	59.74	IPR001223 (PFAM)
scaffold-YQEC-2075825-Woodsia_ilvensis	Leptosporangiate Monilophytes	Woodsiaceae	class V chitinase	2,99339E-67	60.44	IPR001223 (PFAM)
scaffold-ZFGK-2039578-Selaginella_kraussiana	Lycophytes	Selaginellaceae	acidic endochitinase	6,75082E-145	78.43	IPR001223 (PFAM)
scaffold-ZFGK-2039644-Selaginella_kraussiana	Lycophytes	Selaginellaceae	class V chitinase	2,60541E-73	63.17	IPR001223 (PFAM)
scaffold-ZFGK-2040681-Selaginella_kraussiana	Lycophytes	Selaginellaceae	acidic endochitinase	3,32101E-124	76.55	IPR001223 (PFAM)
scaffold-ZFGK-2041140-Selaginella_kraussiana	Lycophytes	Selaginellaceae	class V chitinase	2,11352E-125	63.66	IPR001223 (PFAM)
scaffold-ZQYU-2108391-Polypodium_pectolens	Leptosporangiate Monilophytes	Polypodiaceae	Acidic mammalian chitinase	1,63673E-19	56.31	IPR001223 (PFAM)
scaffold-ZQYU-2112073-Polypodium_pectolens	Leptosporangiate Monilophytes	Polypodiaceae	acidic endochitinase	2,46249E-95	73.84	IPR001223 (PFAM)
scaffold-ZQYU-2112352-Polypodium_pectolens	Leptosporangiate Monilophytes	Polypodiaceae	Chitinase	1,76715E-76	68.5	IPR001223 (PFAM)
scaffold-ZXJO-2006790-Hemionitis_arifolia	Leptosporangiate Monilophytes	Pteridaceae	chitinase-like protein EN03	1,18525E-62	72.39	IPR001223 (PFAM)
scaffold-ZXJO-2006791-Hemionitis_arifolia	Leptosporangiate Monilophytes	Pteridaceae	chitinase-like protein EN03	4,45541E-163	69.01	IPR001223 (PFAM)
scaffold-ZXJO-2015647-Hemionitis_arifolia	Leptosporangiate Monilophytes	Pteridaceae	acidic endochitinase	4,4529E-78	72.95	IPR001223 (PFAM)
scaffold-ZXJO-2070208-Hemionitis_arifolia	Leptosporangiate Monilophytes	Pteridaceae	acidic mammalian chitinase	8,46351E-40	64.78	IPR001223 (PFAM)
scaffold-ZXJO-2070689-Hemionitis_arifolia	Leptosporangiate Monilophytes	Pteridaceae	acidic mammalian chitinase-like	6,31667E-19	57.44	IPR001223 (PFAM)
scaffold-ZXJO-2072539-Hemionitis_arifolia	Leptosporangiate Monilophytes	Pteridaceae	chitotriosidase-1-like isoform X2	7,3608E-67	59.87	IPR001223 (PFAM)
scaffold-ZZOL-2048644-Selaginella_stauntoniana	Lycophytes	Selaginellaceae	acidic endochitinase	3,81683E-57	83.65	IPR001223 (PFAM)
scaffold-ZZOL-2050389-Selaginella_stauntoniana	Lycophytes	Selaginellaceae	class V chitinase	8,68342E-73	72.54	IPR001223 (PFAM)
scaffold-ZZOL-2053046-Selaginella_stauntoniana	Lycophytes	Selaginellaceae	acidic endochitinase	8,78561E-159	80.56	IPR001223 (PFAM)

Figure S1. Class III chitinases alignments of complete sequences identified in green algae

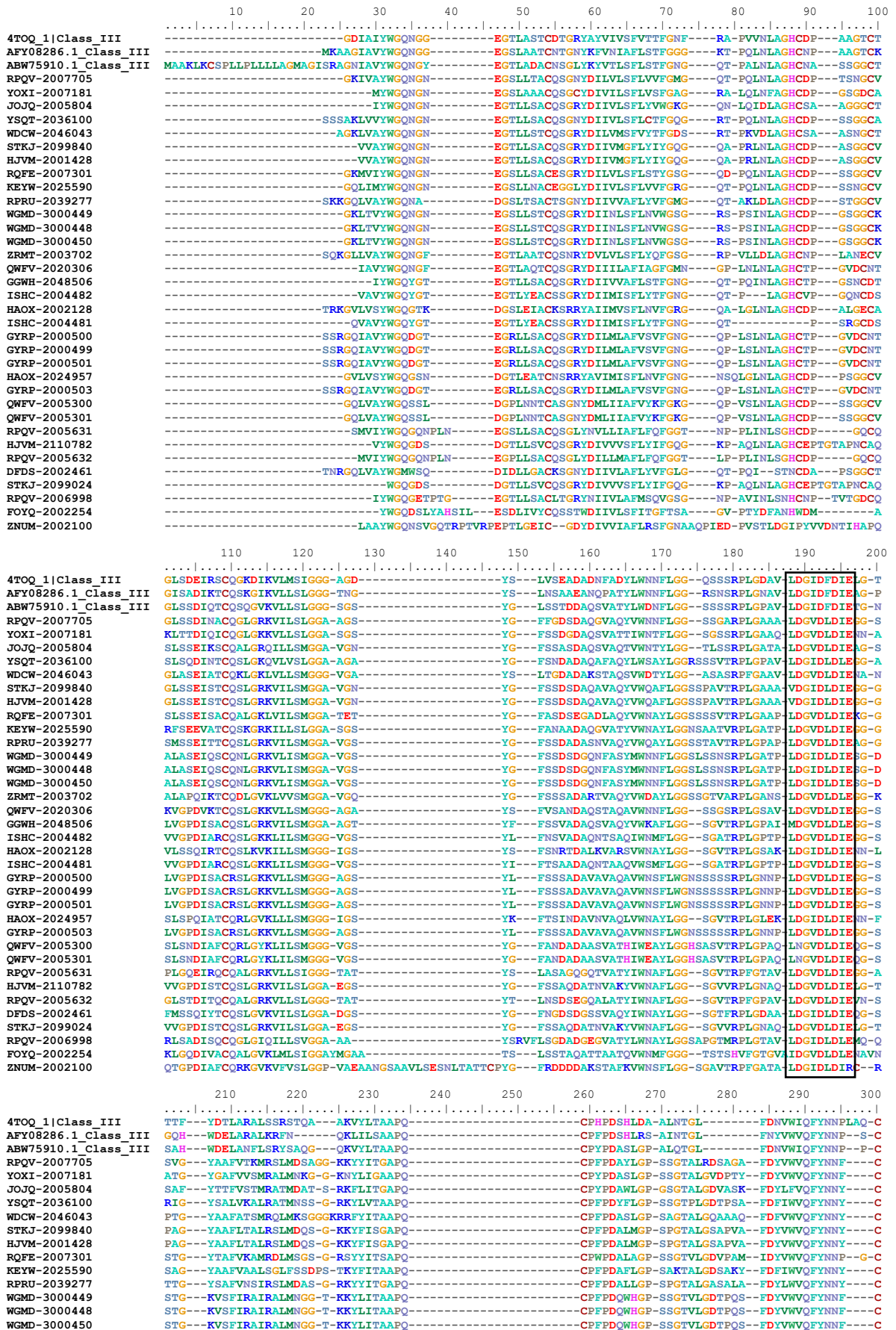
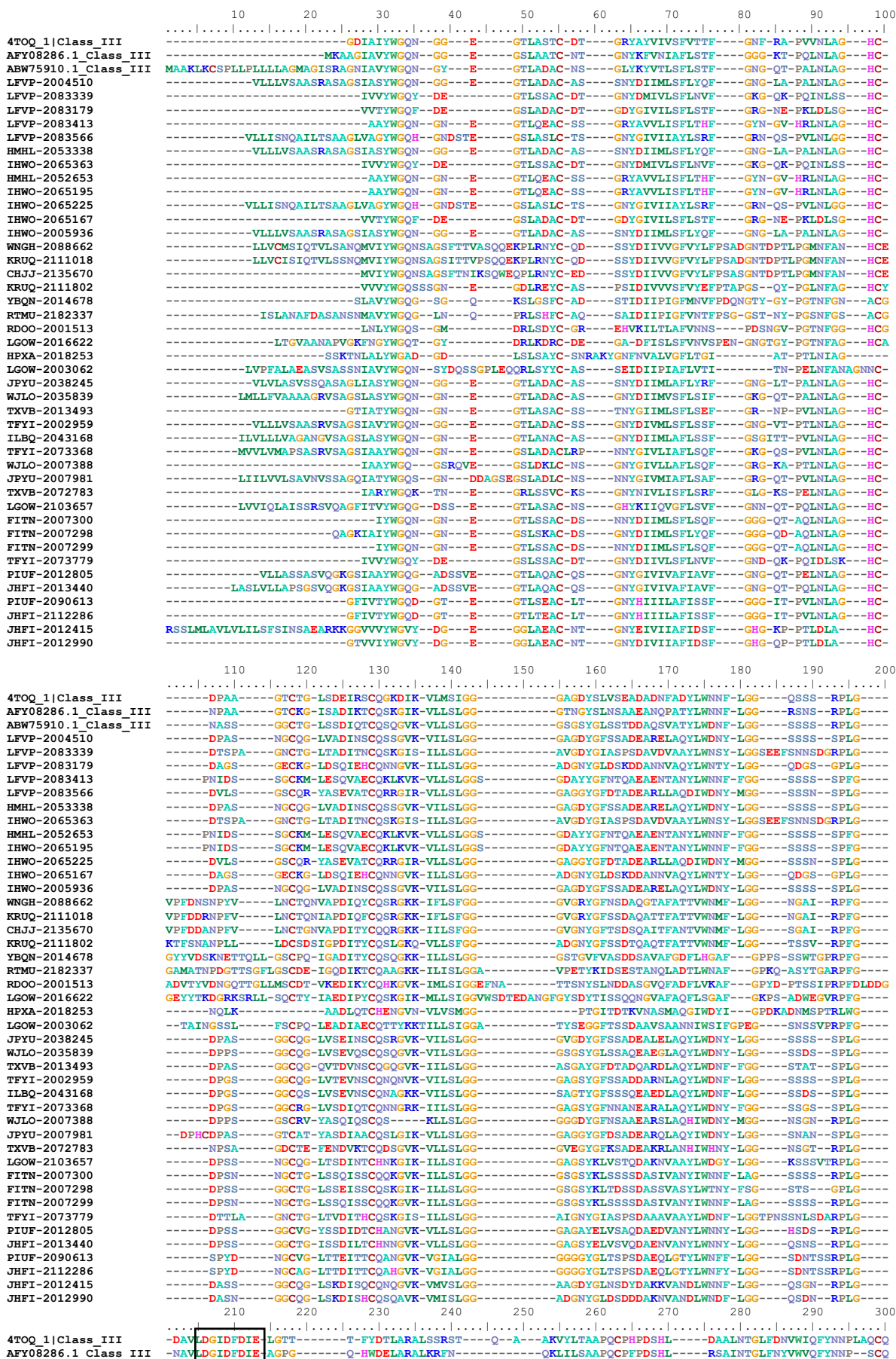
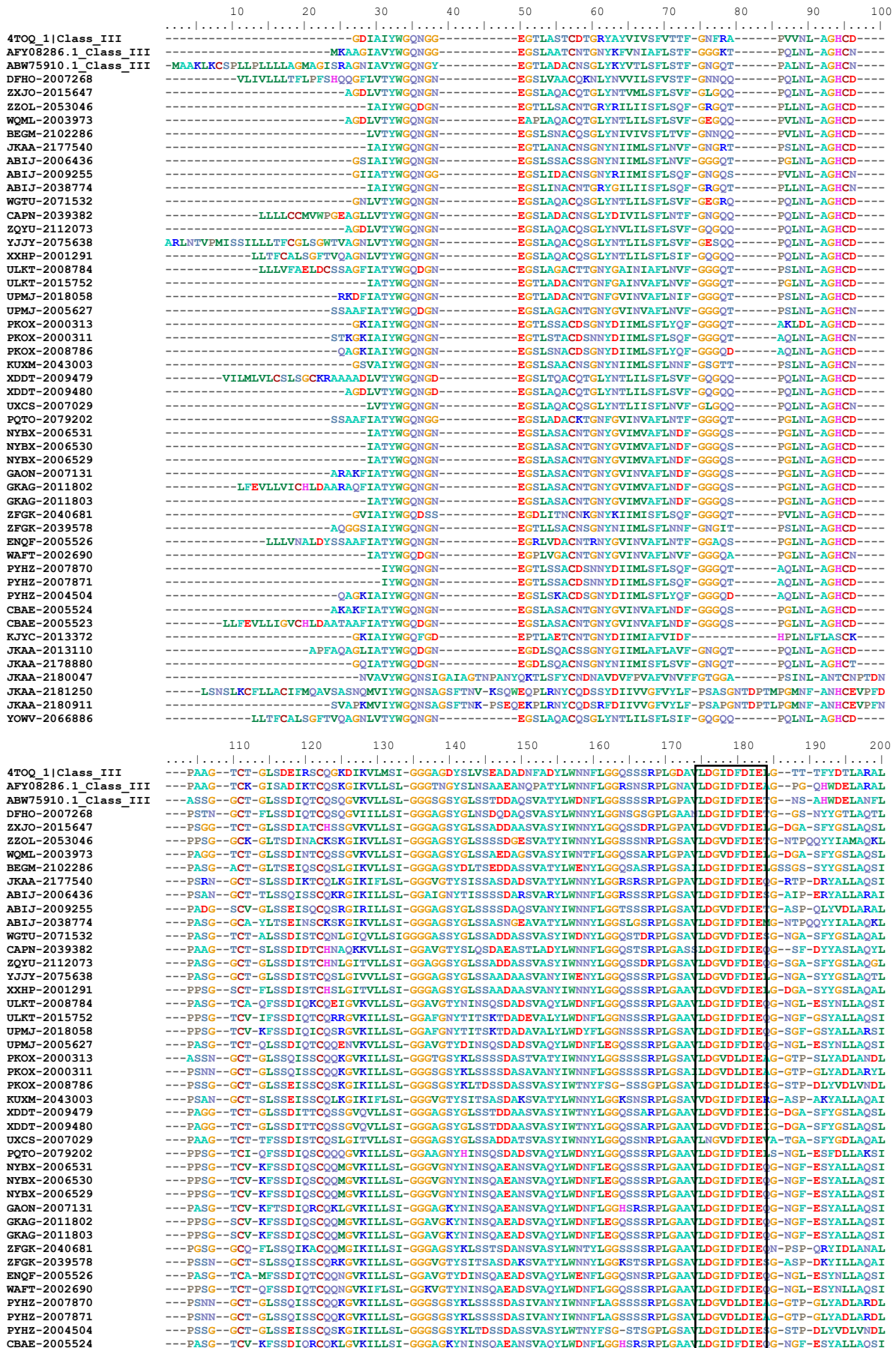


Figure S2. Class III chitinases alignments of complete sequences identified in bryophytes



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HMHL-2052653	NQN-SYSNQISNGI
IHW0-2065195	NQN-SYSNQISNGI
IHW0-2065225	R---TYAASIKSVV
IHW0-2065167	KNP-SYSAAIK---
IHW0-2005936	KENNNYSSQIKSDV
WNGH-2088662	DLNN-----
KRUQ-2111018	DLNN-----
CHJQ-2135670	DMN-----
KRUQ-2111802	-----
YBQN-2014678	-----
RTMU-2182337	-----
RDOO-2001513	-----
LGOW-2016622	-----
HPXA-2018253	-----
LGOW-2003062	-----
JPYU-2038245	KQNNNYSAQIRSGV
WJLO-2035839	KQNDNYSAKIKDSV
TXVB-2013493	IQN-NYSAQIKSAV
TFYI-2002959	RENNNFSAQIRSAV
ILBQ-2043168	QQNNNYSAQIKGSV
TFYI-2073368	RQN-GYSNQIKSHV
WJLO-2007388	-----
JPYU-2007981	-----
TXVB-2072783	T---GYASAIKGA
LGOW-2103657	KQT-GFSSAIASAV
FITN-2007300	KQT-NYAASIRSSI
FITN-2007298	GQT-SYASSIRSSV
FITN-2007299	KQT-TYASSIRSSV
TFYI-2073779	KTS-QFSATIR---
PIUF-2012805	QIT-SFSSSI----
JHFI-2013440	RIT-SFSSSI----
PIUF-2090613	QNP-SYSGEI----
JHFI-2112286	QNP-SYSGEILRAV
JHFI-2012415	-----
JHFI-2012990	-----

Figure S3. Class III chitinases alignments of complete sequences identified in pteridophytes



WAFT-2002690 TSAAGS-GFIPSDVLISSILPQIKSST--KYGGVMVWDYSHD--NGYSSAIK---
 PYHZ-2007870 SAAAGS-GYMSSDVLKSKVLPQIKSSS--KYGGVMFWSVYYDKQTNAAASIRSSI
 PYHZ-2007871 SAAAGS-GYMSSAVLKSQVLPQIKSSS--KYGGVMLWSVYYDKQTSYASSIRSSV
 PYHZ-2004504 SDAAGS-GYMSSDVLKSDMLPTIKGSS--KYGGVMLWSVYYDGTQTSYASSIRSSV
 CBAE-2005524 SSAAGS-GFIPSGVLTSTILPRIKNSP--KFGGVMLWDYSHD--KGYSSSIK---
 CBAE-2005523 SSAAGS-GFIPSGVLTSTILPRIKNSP--KFGGVMLWDYSHD--KGYSSSIK---
 KJYC-2013372 TEAAPSGGYIEPDKFISEVLPASLSP--KYGGVMLWAVYFDLQTKYSSKIRPYV
 JKAA-2013110 TAAAGS-GFIPSDVLSQVLPQIKSSS--KYGGVMLWSVAQD--SGYSAAIKSSV
 JKAA-2178880 PTAAPSGGYIPPSVLTSLRVLPAIKTSP--KYGGIMLYSVAYD-----
 JKAA-2180047 SSAAGS-GYVDINTLSKIAVQMRKFS--SFGGVMLW-----
 JKAA-2181250 SLAAAS-GFVSLSTLQT-ISTAIQNTFPNLYGGMLW-----
 JKAA-2180911 SMAAAS-GFVPISTLKT-ISTTIANTFPNIYGGMMVW-----
 YOWV-2066886 PGAAGS-GFIPASTLISEVLPQIRSTS--NYGGVMLYSYYDETSYSGSTIKSSV

Figure S4. Class V chitinases alignments of complete sequences identified in algae phyla Rhodophyta and Ochrophyta

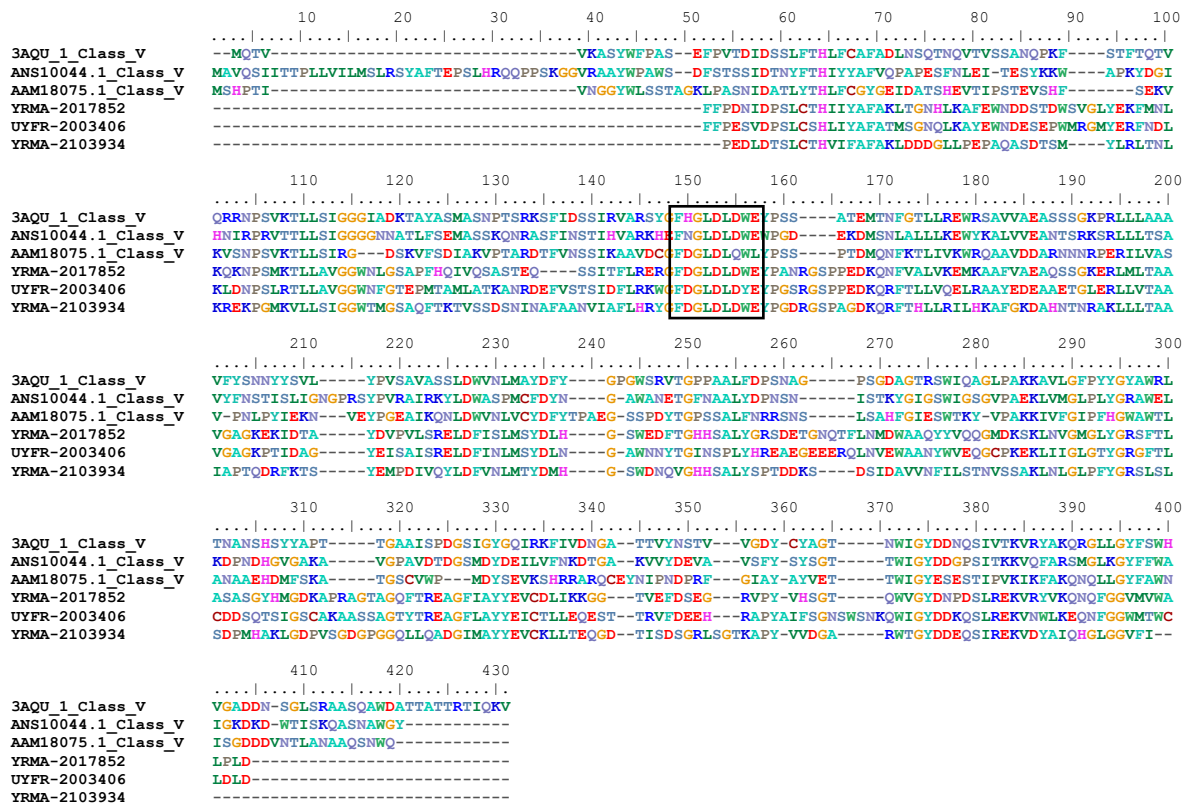
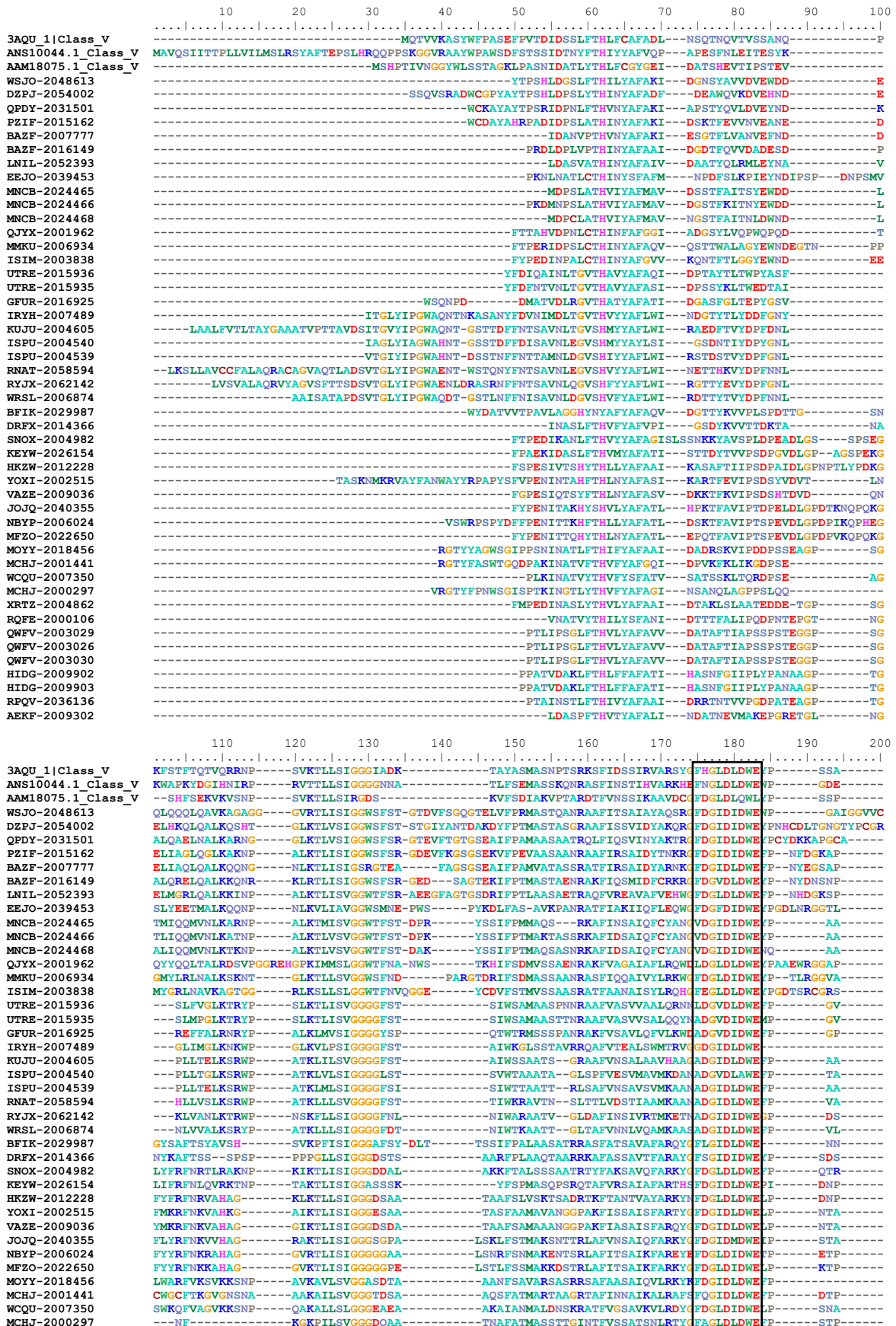


Figure S5. Class V chitinases alignments of complete sequences chitinases identified in green algae




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WCQU-2007350 DATGHSALY-DAAFP-----DQSCSAAQTWISGG-LDPKVIIGLASYGHFWTLQSAKNG-VG---VP---ATDP---R-GIWSYPO
MCHJ-2000297 TITCLHTALY-DTSNP-----TASCQAQVQTFPAGG-VAASKILVMSGLASYGNSWTLKYTTGTC-VGAWWGAA---ATGP--GAAGAITRTA---GILSYFE
KRTZ-2004862 SQTQEGHTALQ-DRNDP-----PLSIAGAIDAWKSAG-MPANKILAGLAFYGHWTWLKSPALHG-VG---AP---ATGA--GAVGAVSQDP---GTLAYFE
RQFE-2000106 TTTHAHTALE-DLALY-----AASCKGAIYAWASGG-LSASKTVMGLASYGVGFTLTPNTITG-MG---AP---AKGG---ADMSYAA
QWV-2003029 SVTQEGHTALY-DPSDP-----GLSCVGGISVWRRGG-MAASKILMGLAAYGRTWTLVVSAGSHG-IG---AA---ASGP--GAPGSQTNAP---GIFSYGE
QWV-2003026 SVTQEGHTALY-DPSDP-----GLSCVGGISVWRRGG-MAASKILMGLAAYGRTWTLVVSAGSHG-IG---AA---ASGP--GAPGSQTNAP---GIFSYGE
QWV-2003030 SVTQEGHTALY-DPSDP-----GLSCVGGISVWRRGG-MAASKILMGLAAYGRTWTLVVSAGSHG-IG---AA---ASGP--GAPGLQTNAP---GVFSYGE
HIDG-2009902 QHTGLHAALE-DPANP-----DASCAGAVQALTGGG-MPASKILMGLAAYGRSWTLKDAADNG-VG---AP---TTGV--GLPGPQTDP---GLLSYAE
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RPQV-2036136 DVTCEGPTALA-DPSNV-----DTSCTGALQAWLGGG-LNPSRFVVLASYGATWTLASARRYG-IG---AK---TVGP--GTPGPATQEA---GILSYGE
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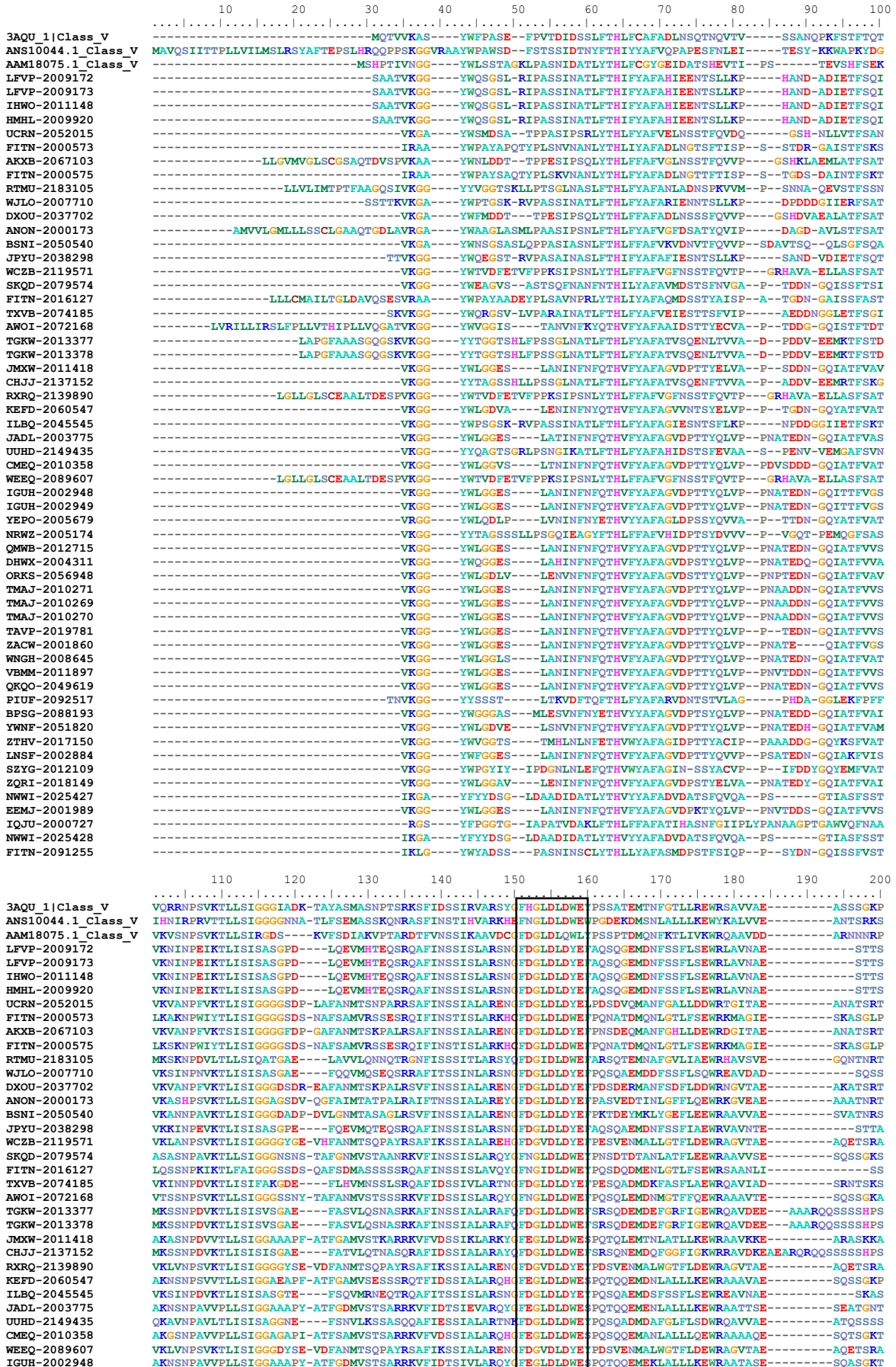
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AAM18075.1_Class_V VLSHRRARQCEY---NIPNDPRF-GIAYAY-VET-TWIGYSESEITPVKIKPAKQ-QLLGYFAWNISGDDVWNTLANAASQSNWQ-----
WSJO-2048613 IERFV---DSG---AIKVDPEV-MAAYAR-YNESVWVGFDAETHRRKMCYARVY-QLGGVFLW-----
DZPJ-2054002 LKRKL---APD---SIKVDKSLLAAYATYDDGQSWAGFDNQEESHRLKMCWARQQ-KLGGVMW-----
QPDY-2031501 INRLV---NKV---DIREDTASLMAAWAP-YGADGWVGFDETEHTHLRKTCTYARVY-RLGGIMVW-----
PZIF-2015162 IKKVY---A---KSIDPVQ-MAAYGV-TTSGHWVGFDETPETHRMKMCYARAR-GISGVMVWDSDDTD-----
BAZF-2007777 IKQLV---PND---AIKVDVSA-MAAYASYKSGTEWVGFDETEHLRKLKCYARSR-GITGAMVWDNMMD-----
BAZF-2016149 IKRKY---PTA---DIRIDDKA-LAAYASFDGGRQWIGFDNVESHRLKMCYARSL-GLGGVMVWASLED-----
LNIL-2052393 IQRAL--QAEGL---AVREDKH-MAAYVVMRGGREWVYDTPSTMQQKVCVARAM-GVGGVFLWTDAD-----
EEJO-2039453 IKRLL---EQGA---QVTIDEGA-MAAYLV-SGS-TWVAFDLPQTIYMKILARQL-GLGGLMVW-----
MNCB-2024465 IKDYL---GSI---QPSVNTLV-ASAYAV-YGSSYVVGVDTEATLGLKVCYARVY-NLGG-----
MNCB-2024466 IKNAL---GSI---QPSVDTVW-ASAYAV-YGSSYVVGFDTEATLALKVCYAKYK-KLGG-----
MNCB-2024468 IKNAL---GSI---QPSVDTVW-ASAYAV-YGSSYVVGFDTEATLALKVCYAKYK-KLGG-----
QJYX-2001962 IRAIL--ERDDP---VYIRDAPY-GVPHFV-YGDNVWSYDDVASIAAKTDFLREK-ALRGGMVWALDLDLA-----
MMKU-2006934 IAAKQ--AAGTA---TAVVDVAR-EAPYAY-WSN-QWVYDQAQSIQKVQLANQN-GLLGAMLWALDLDL-----
ISIM-2003838 VAAVL--ASGAA---TRVFDVAR-GAPYAH-WGD-QWVYDDEESLARKAEVLRAL-GLRGGMVWALDLDL-----
UTRE-2015936 IMAEM--AALGG---AVYKSDAQ-RSAYVY-RGT-LWVGFVQETIYFKMGGAAAF-GLGGVMIWDLSDSDPDQRL-----
UTRE-2015935 LMAQM--AAQGG---SVYKDNQ-RSAYVY-LGT-LWVGFVQETIYAKMGAAAF-GLGGVMIWDLSDQPTDQRL-----
GFOR-2016925 IMAAR--GSAAGSTKAGIFQSTTE-RSAYFV-DGT-SWVGFVDETELWAKIQGATER-GLGGVMIWVSLDTF-----
IRYH-2007489 LMKEQ--VANGG---TVFVDVPE-KSAYVY-RGT-RWVGFDETPETMWLKIQAQQOR-GLGGVMIWDASLDTPVNOQL-----
KUJU-2004605 ILREL--SLRGG---GVFVNDTE-RSAYVY-RGT-RWVGFDETPETMALKIRGAREY-GTGGVMIWDASLDTD-----
ISPU-2004540 IVREF--EVYGG---KILVDATE-RSSYVY-RGT-RWVGFDETEELAAKIQGAREY-GIGGMMIWEASQDT-----
ISPU-2004539 IMSLEL-LTRGG---GIFVGEAE-RSAYVY-RGT-RWVGFDETEESFAAKIQGSRSEY-GIGGVMVWDLSD-----
RNAT-2058594 ILRDL--ATTGG---GVFVDTQ-RSAYVY-YDT-RWVGFDETEELAAKIQGSRSEY-GVGGVMIWDLSDTPD-----
RYJX-20062142 VMRLEL-NYSGG---GVFVDVGE-RSAYVY-SGL-KWVGFVDEVPESMALKIQGTREY-GLAGVMIWDLSDTPSD-----
WRSL-2006874 ILHEL--ATIGG---SVFMDVPQ-RSAYVY-RGT-RWVGFDETEELAAKIQGSRSEY-GVGGVMIWDLSDVRE-----
BFIK-2029987 IKALVTSAKNPW---TSFLDSNT-QCQYAY-GGGNQWVGYDNPPTIIGLKASFAKSK-GLGGYFVWVSLDSD-----
DRFX-2014366 IKSNV--QSGGW---TAKYSPA-TAMYAY-KCK-QWVGYDN-----
SNOX-2004982 ILSML--SLPGT---ITVFDNNT-SSVYSV-NGL-QWVGYDNQPSIRTKVYFKNSIKALGYFFWALDLDL-----
KEYW-2026154 ILNFI--QGGG---TVVFDKAS-TSMYAY-KGT-QWVGYDDVSTIQTKAKYAQSN-GLGGYFWAMDLDL-----
HKZW-2012228 INSFI--RSGA---TVVFNQSK-ASVYAY-KGS-HWVGYDTRTISLKAKYAQSQ-GFGGYFWAMDLDL-----
YOXI-2002515 IVDKF--LKSGA---TAMFDNNT-SSMYAY-KCR-QWVGYDNEKSIKAKYAQSK-GLGGYFWNVDM-----
VAZE-2009036 IVNFF--LKSGA---TAVFDKTT-SSEYAY-KCR-QWVGYDKEKSIKAKYAQSH-GLGGYFWNVDM-----
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MCHJ-2001441 INLFL--RGRGA---VSVFDRKT-TCQYAF-KGN-QWVGYDNTLITTKVRFKMR-GLSGYAFWA-----
WCQU-2007350 IKAFI--KKA---VSNYSAAY-VGQYAY-KGT-QWVGYDNTIATKVKYASR-GIGGYGFW-----
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HIDG-2009902 IAGYV--KGA---VAKLDSAT-SSMYAH-WGT-QWVGYDNTIATKVFASKR-GLGGYAFW-----
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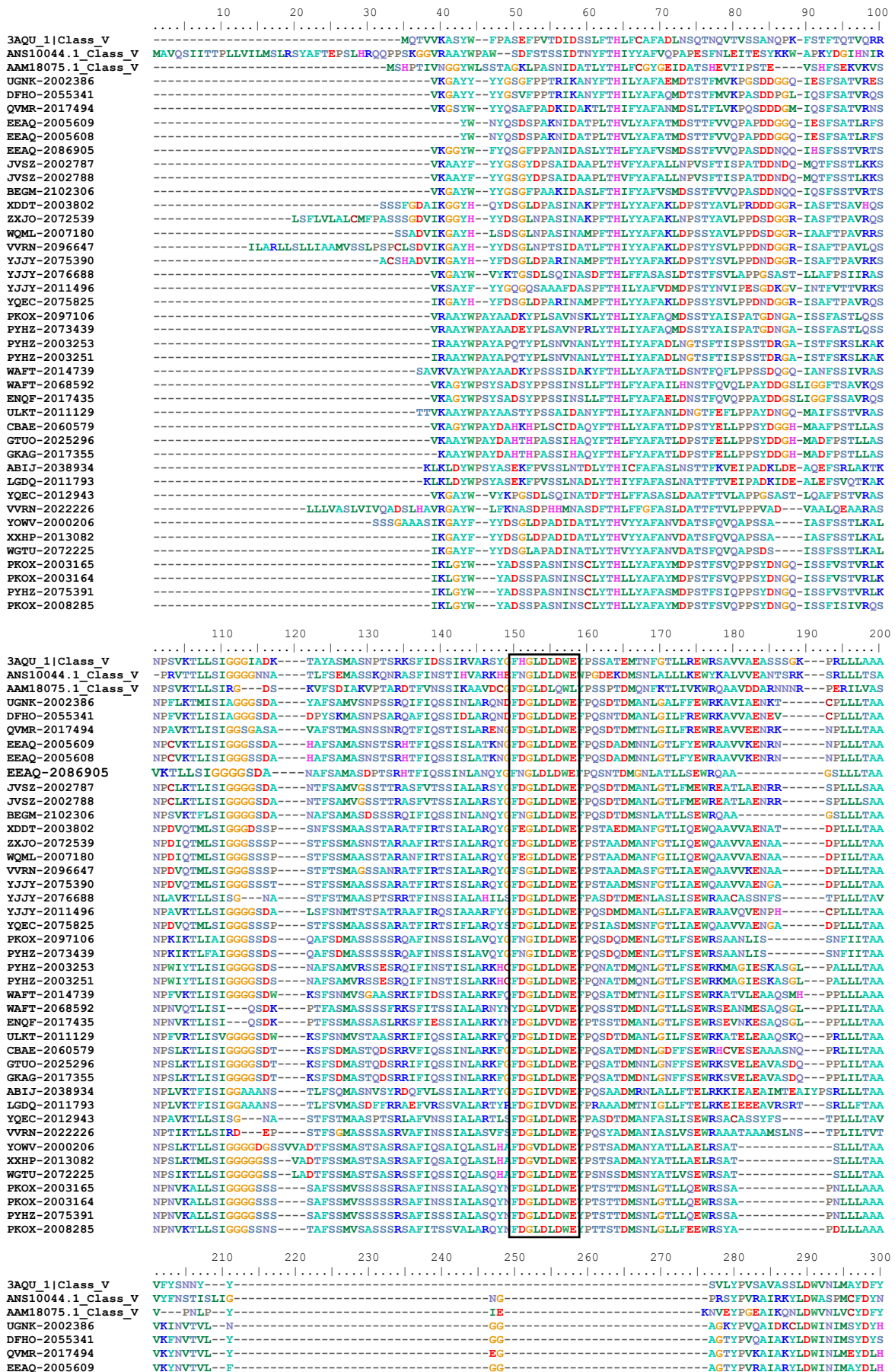
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Figure S6. Class V chitinases alignments of complete sequences identified in bryophytes



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SZYG-2012109	SVCGYFAWNLAQDDSN-----WSLAKAASN-----
ZQRI-2018149	KLLGFFAWALHQDDA-----WSLAS-----
NWWI-2025427	QLAGYFFWRLGLD-----
EEMJ-2001989	ELGGFFAWALYHDTGN-----WSLASAASN-----
IQJU-2000727	GLGGYAFW-----
NWWI-2025428	QLAGYFFWRLGLD-----
FITN-2091255	GLRGYGFWT-----

Figure S7. Class V chitinases alignments of complete sequences identified in pteridophytes




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PKOX-2097106 -DNDATV-VYDNSTVTA-YCYSQ-----TSWIGFDNNLSIAGKVYKLSKLLKGYFFWTSSYDDNKFTLSRTAAYA-----
PYHZ-2073439 -DNDATV-VYDNSTVTA-YCYSQ-----TSWIGFDNNLSIAGKVYKLSKLLKGYFFWTSSYDDNKFTLSRTAAYA-----
PYHZ-2003253 SSSHASL-VFDNNTLCS-YCYSG-----TSWIGFDDRVSIAAKV-----
PYHZ-2003251 -QNHAAL-VFDNNTLCS-YCYSG-----TSWIGFDDRVSIAAKVGFLKSKGLQGYFFWTASYDDKKYTLSTTAAQS-----
WAPT-2014739 -DKHATV-VFDKATVTT-YCYAD-----NLWIGFDPPVSIKKVRYAKQKGLKGYFFWTASFDS-----
WAPT-2068592 -SSNATV-VFDDTTASV-YTYSG-----TIWIGYDNPRISSKVAFLNSKGLLGYFFWTVSFD-----
ENQF-2017435 -SSNATV-VFDETTSSV-YTYSG-----TIWIGYDDPRISISKVAFLNSKGLLGYFFWTVSFD-----
ULKT-2011129 -EKNATV-VFDRFTVTT-YCYAD-----KLWIGFDDPVSIFEKRVYAKKRGKGLKGYFFWTSSFDKWTLSSSAYTA-----
CBAE-2060579 -KKHARV-VFDPTVTT-YCYAH-----NLWVGFDSSVSIKKVRYAKKGLKGYFFWTVSFD-----
GTUO-2025296 -KKHARV-VFDPTVTT-YCYAH-----NVWIGFDDPVSIFEKRVYAKKGLKGYFFWTVSFD-----
GKAG-2017355 -KKHARV-VFDPTVTT-YCYAH-----NLW-QFQ-----SEKRVYAKQKGLKGYFFWTVSFD-----
ABIJ-2038934 -KNNATV-VDDAVTGSA-YAFAG-----DLWVGFDNKVSIKKVRYLKSKEMRGYFFWTSSFD-----
LGDQ-2011793 -EKNATV-VDDAVTGSA-YAFAG-----DFWVGFDNKVSIKKVRYLKSKEMRGYFFWTSSFD-----
YQEC-2012943 -NSKAYC-RDDVVTKAA-FCYGRQLSSGSSNCLWVGFDNPRITATKRVYLNKAKKLRGYFLWLSLGFDDKDLKQAS-----
VVRN-2022226 -SSKAYC-RDDAVTKAS-FCYGRQLSSGSSNCLWVGFDNPRITATKRVYLNKAKKLRGYFLWLSLGFDDKDLKQAS-----
YOWV-2000206 -ANSATCQRDSTITVAS-YCYGS-SGSD---ILWVGFDDAISIAAKVEYLKSKQLGGYFFWRLGFDQNNVLASQASTS-----
XXHP-2013082 -ANSATCQRDSTITVAS-YCYGS-SGSD---ILWVGFDDAISIAAKVEYLKSKQLGGYFFWRLGFDQNNVLASQASTS-----
WGTU-2072225 -SSSTTCQRDSTITVAS-YCYGN-SGSN---VLWVGFDDAISIAAKVYKLSKQLAGYFFWRLGFD-----
PKOX-2003165 -STGATI-VHNATSVST-YAYSG-----RAWVGYDDAWSITRKVNLYLKSGLRGYGFWT-----
PKOX-2003164 -STGATI-VHNATSVST-YAYSG-----RAWVGYDDAWSITRKVNLYLKSGLRGYGFWT-----
PYHZ-2075391 -STGATI-VHNATYVST-YAYSG-----RAWVGYDDAWSITRKVNLYLKSGLRGYGFWT-----
PKOX-2008285 -SSGATV-VHNTSSVST-YAYSG-----ITWVGYDDAWSITRKVNLYLKSGL-RGYGFWTVLSDDSNFTLA-----

```

Figure S8. Phylogenetic tree of GH18 family chitinases identified in algae phyla Rhodophyta and Ochrophyta

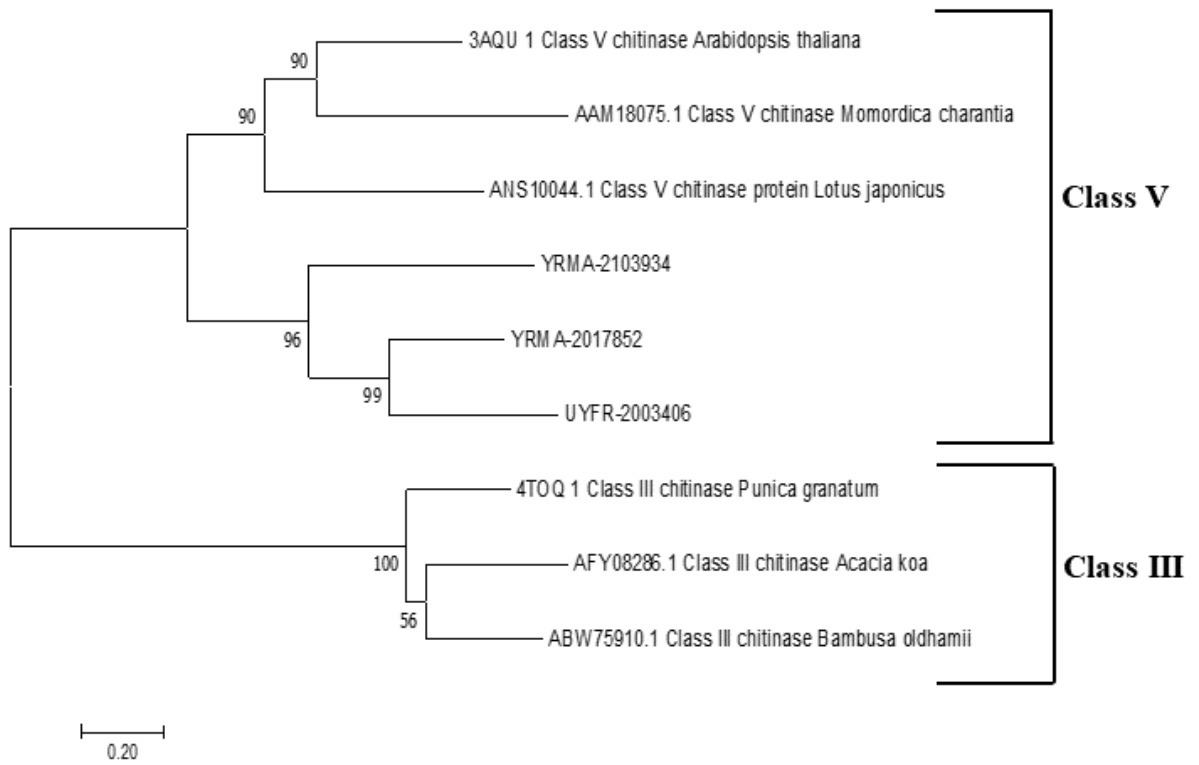


Figure S9. Phylogenetic tree of GH18 family chitinases identified in green algae.

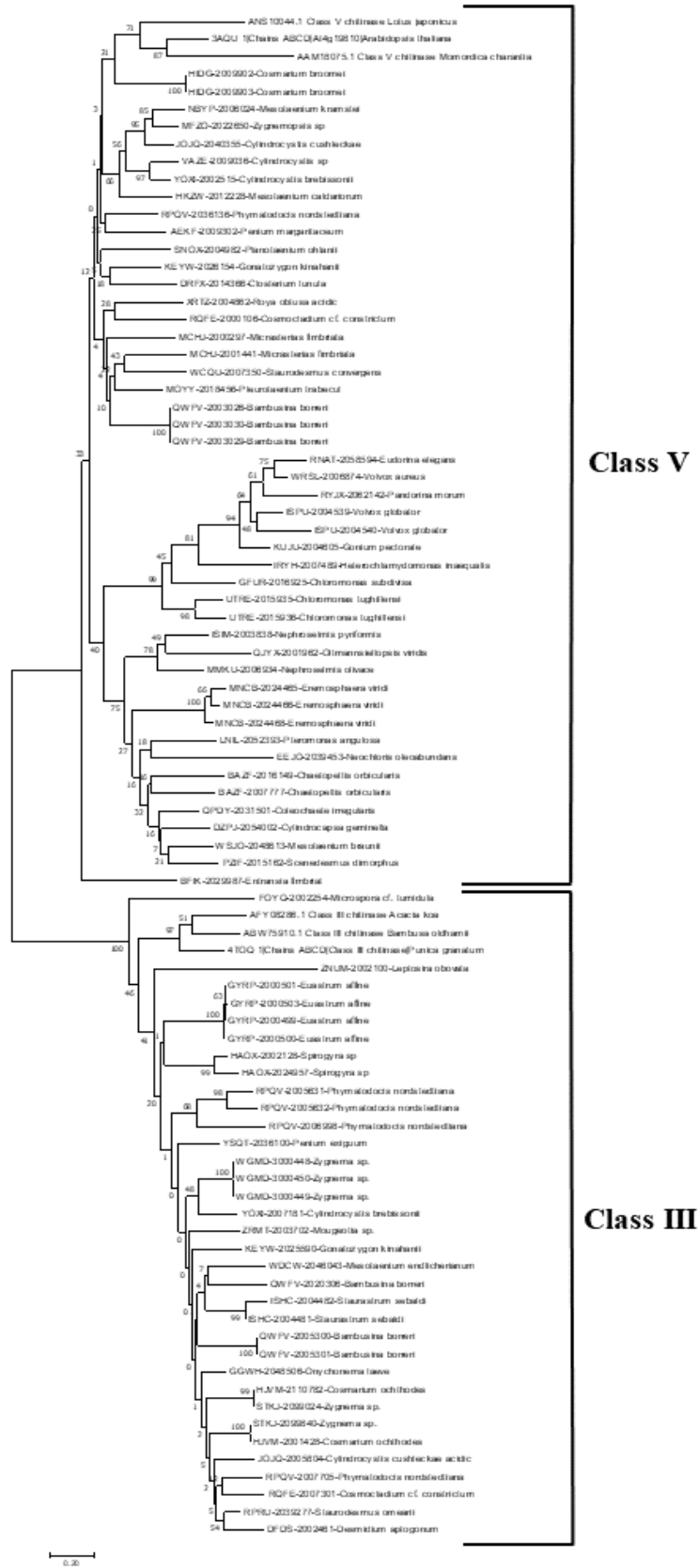


Figure S10. Phylogenetic tree of GH18 family chitinases complete sequences identified in bryophytes

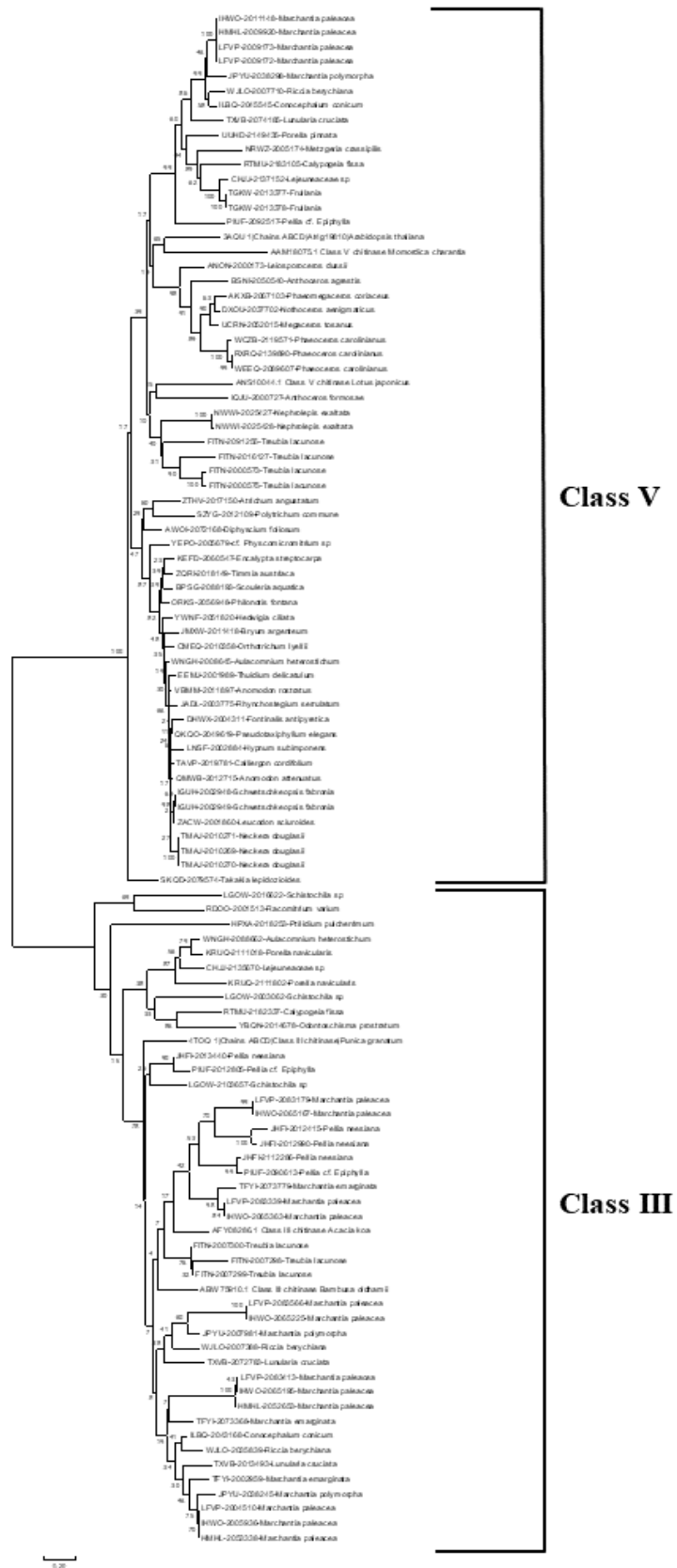


Figure S11. Phylogenetic tree of GH18 family chitinases complete sequences identified in pteridophytes



CONCLUSÃO

As análises conduzidas neste trabalho revelaram a elevada diversidade de quitinases da família GH18 em algas, briófitas e pteridófitas. No total, foram encontradas 648 possíveis quitinases da família GH18, distribuídas em algas, briófitas e pteridófitas. De um modo geral, as quitinases demonstraram ter os domínios catalíticos altamente conservados em todos os grupos analisados. Mas algumas quitinases apresentaram deleções ou substituições em resíduos de aminoácidos fundamentais para a catálise, o que pode interferir na função quitinolítica. A árvore filogenética separou claramente a classe III da classe V, pois apesar de pertencerem a mesma família, essas classes apresentam baixa similaridade entre si. As estruturas tridimensionais de algas verdes, briófitas e pteridófitas demonstraram ter arranjos semelhantes entre si. A identificação e caracterização de quitinases em grupos mais ancestrais das plantas terrestres é importante para compreender a dinâmica dessas proteínas nesses grupos e possibilitar a comparação com quitinases de gimnospermas e angiospermas. Os resultados deste estudo podem também ser úteis para o fornecimento de dados para estudos futuros relacionados a quitinases da família GH18 em plantas.

REFERÊNCIAS

- ADAMS, D. J. Fungal cell wall chitinases and glucanases. **Microbiology**, v. 150, n. 7, p. 2029–2035, 2004. <https://doi.org/10.1099/mic.0.26980-0>
- AMEND, A. et al. Fungi in the marine environment: Open questions and unsolved problems. **mBio**, v. 10, n. 2, p. 1–15, 2019. <https://doi.org/10.1128/mBio.01189-18>
- ANDERSON, J. T.; WILLIS, J. H.; MITCHELL-OLDS, T. Evolutionary genetics of plant adaptation. **Trends Genet**, v. 27, n. 8, p. 258–266, 2011. <https://doi.org/10.1016/j.tig.2011.04.001>
- ARIE, M. et al. Characterization of a basic chitinase which is secreted by cultured pumpkin cells. **Physiologia Plantarum**, v. 110, n. 2, p. 232–239, 2000. <https://doi.org/10.1034/j.1399-3054.2000.110213.x>
- BADARIOTTI, F. et al. The phylogenetically conserved molluscan chitinase-like protein 1 (Cg-Clp1), homologue of human HC-gp39, stimulates proliferation and regulates synthesis of extracellular matrix components of mammalian chondrocytes. **Journal of Biological Chemistry**, v. 281, n. 40, p. 29583–29596, 2006. <https://doi.org/10.1074/jbc.M605687200>
- BASKARAN, X-R, et al. A review of the use of pteridophytes for treating human ailments. **Journal of Zhejiang University: Science B**, v. 19, n. 2, 2018. <https://doi.org/10.1631/jzus.B1600344>
- BENNICI, A. Origin and early evolution of land plants. **Communicative & Integrative Biology**, v. 1, n. 2, p. 212–218, 2008. <https://doi.org/10.4161/cib.1.2.6987>
- BHATTACHARYA, D.; YOON, H. S.; HACKETT, J. D. Photosynthetic eukaryotes unite: Endosymbiosis connects the dots. **BioEssays**, v. 26, n. 1, p. 50–60, 2004. <https://doi.org/10.1002/bies.10376>
- BLACKWELL, W. H. Two Theories of origin of the land-plant sporophyte: Which Is Left Sanding? **The Botanical Review**, v. 69, n. 2, p. 125–148, 2003. Disponível em: link.gale.com/apps/doc/A112542838/AONE?u=capes&sid=AONE&xid=004c651d. Acesso em: 15 nov. 2020.
- BRAMELD, K. A. et al. Substrate assistance in the mechanism of family 18 chitinases: Theoretical studies of potential intermediates and inhibitors. **Journal of Molecular Biology**, v. 280, n. 5, p. 913–923, 1998. <https://doi.org/10.1006/jmbi.1998.1890>
- BRZEZINSKA, M. S. et al. Chitinolytic microorganisms and their possible application in environmental protection. **Current Microbiology**, v. 68, n. 1, p. 71–81, 2014. <https://doi.org/10.1007/s00284-013-0440-4>
- CAMPANA-FILHO, S. P. et al. Extração, estruturas e propriedades de α - e β -quitina. **Química Nova**, v. 30, n. 3, p. 644–650, 2007. <https://doi.org/10.1590/S0100-40422007000300026>

- CAO, J.; TAN, X. Comprehensive analysis of the chitinase family genes in tomato (*Solanum Lycopersicum*). **Plants**, v. 8, n. 3, 2019. <https://doi.org/10.3390/plants8030052>
- CASANOVA, M. T.; NAIRN, L. C. Macroalgae, charophytes and bryophytes. In: CAPON, S., JAMES, A., REID, M. **Vegetation of Australian Riverine Landscapes**, Austrália: CSIRO, 2016. cap. 5, p. 67–103. Disponível em: <https://www.researchgate.net/publication/296396211>. Acesso em: 15 nov. 2020.
- CAVALIER-SMITH, T. The origins of plastids. **Biological Journal of the Linnean Society**, v. 17, n. 3, p. 289–306, 1982. <https://doi.org/10.1111/j.1095-8312.1982.tb02023.x>
- CHEN, L. et al. Enhanced nematicidal potential of the chitinase pachi from *Pseudomonas aeruginosa* in association with Cry21Aa. **Scientific Reports**, v. 5, p. 1–11, 2015. <https://doi.org/10.1038/srep14395>
- CHRISTENHUSZ, M. J. M.; BYNG, J. W. The number of known plants species in the world and its annual increase. **Phytotaxa**, v. 261, n. 3, p. 201–217, 2016. <https://doi.org/10.11646/phytotaxa.261.3.1>
- CHRISTENHUSZ, M. J. M.; CHASE, M. W. Trends and concepts in fern classification. **Annals of Botany**, v. 113, n. 4, p. 571–594, 2014. <https://doi.org/10.1093/aob/mct299>
- CHUANG, H. H.; LIN, H. Y.; LIN, F. P. Biochemical characteristics of C-terminal region of recombinant chitinase from *Bacillus licheniformis* - implication of necessity for enzyme properties. **FEBS Journal**, v. 275, n. 9, p. 2240–2254, 2008. <https://doi.org/10.1111/j.1742-4658.2008.06376.x>
- COOPER, E. D. Overly simplistic substitution models obscure green plant phylogeny. **Trends in Plant Science**, v. 19, n. 9, p. 576–582, 2014. <https://doi.org/10.1016/j.tplants.2014.06.006>
- COTAS, J. et al. A comprehensive review of the nutraceutical and therapeutic applications of red seaweeds (Rhodophyta). **Life**, v. 10, n. 3, 2020. <https://doi.org/10.3390/life10030019>
- EDREVA, A. Pathogenesis-related proteins: Research progress in the last 15 years. **Gen Appl Plant Physiol**, v. 31, n. 1–2, p. 105–24, 2005. Disponível em: http://www.bio21.bas.bg/ipp/gapbfiles/v-31/05_1-2_105-124.pdf. Acesso em: 12 nov. 2020.
- EL-KATATNY, M. H. et al. Characterization of a chitinase and an endo- β -1,3-glucanase from *Trichoderma harzianum* Rifai T24 involved in control of the phytopathogen *Sclerotium rolfsii*. **Applied Microbiology and Biotechnology**, v. 56, n. 1–2, p. 137–143, 2001. <https://doi.org/10.1007/s002530100646>
- EVERT, R. F.; EICHHORN, S. E. **Raven Biologia Vegetal**. 8ª ed. Rio de Janeiro: Guanabara Koogan, 2014. Disponível em: file:///D:/DADOS%20USER/Downloads/LIVRO_2014%20-%20Raven%20Biologia%20Vegetal%208%C3%82%C2%AAEd.%20Fungos%20558.pdf. Acesso em: 10 ago. 2020.
- FANG, L. et al. Evolution of the chlorophyta: Insights from chloroplast phylogenomic analyses. **Journal of Systematics and Evolution**, v. 55, n. 4, p. 322–332, 2017.

<https://doi.org/10.1111/jse.12248>

FRANGEDAKIS et al., 2020. The hornworts: Morphology, evolution and development. **New Phytol**, v. 229, n. 2, p. 735–754. <https://doi.org/10.1111/nph.16874>

FLEURI, L. F.; SATO, H. H. Produção, purificação, clonagem e aplicação de enzimas líticas. **Química Nova**, v. 28, n. 5, p. 871–879, 2005. <https://doi.org/10.1590/S0100-40422005000500026>

FRAHM, J.-P. et al. Manual of tropical bryology. **Bryophyte Diversity and Evolution**, v. 23, n. 1, p. 1–200, 2003. <https://doi.org/10.11646/bde.23.1.1>

FUKAMIZO, T. Chitinolytic enzymes: Catalysis, substrate binding, and their application. **Current Protein & Peptide Science**, v. 1, n. 1, p. 105–124, 2000. <https://doi.org/10.2174/1389203003381450>

GRAHAM, L. E.; COOK, M. E.; BUSSE, J. S. The origin of plants: Body plan changes contributing to a major evolutionary radiation. **Proceedings of the National Academy of Sciences of the United States of America**, v. 97, n. 9, p. 4535–4540, 2000. <https://doi.org/10.1073/pnas.97.9.4535>

GUIRY, M. D. How many species of algae are there? **Journal of Phycology**, v. 48, n. 5, p. 1057–1063, 2012. <https://doi.org/10.1111/j.1529-8817.2012.01222.x>

HAMID, R. et al. Chitinases: An update. **J Pharm Bioallied Sci**, v. 5, n. 1, p. 21–29, 2013. <https://doi.org/10.4103/0975-7406.106559>

HAN, P. et al. Identification and characterization of a novel chitinase with antifungal activity from ‘Baozhu’ pear (*Pyrus ussuriensis* Maxim.). **Food Chemistry**, v. 196, p. 808–814, 2016. <https://doi.org/10.1016/j.foodchem.2015.10.006>

HE-NYGRÉN, X. et al. Illuminating the evolutionary history of liverworts (Marchantiophyta) - Towards a natural classification. **Cladistics**, v. 22, n. 1, p. 1–31, 2006. <https://doi.org/10.1111/j.1096-0031.2006.00089.x>

HO, B. The liverwort genus *Marchantia* L. (Marchantiophyta: Marchantiopsida) in Singapore, with a new species record. **Nature in Singapore**, v. 6, p. 187–190, 2013. Disponível em: <https://www.researchgate.net/publication/260851139>. Acesso em: 10 nov. 2020.

HUTTUNEN, S.; BELL, N.; HEDENÄS, L. The evolutionary diversity of mosses—Taxonomic heterogeneity and its ecological drivers. **Critical Reviews in Plant Sciences**, v. 37, n. 2–3, p. 128–174, 2018. <https://doi.org/10.1080/07352689.2018.1482434>

INGROUILLE, M.; EDDIE, B. **Plants: Evolution and diversity**. New York: Cambridge University Press, 2006. <https://doi.org/10.1017/CBO9780511812972>

ISELI, B. et al. Plant chitinases use two different hydrolytic mechanisms. **FEBS Letters**, v. 382, n. 1–2, p. 186–188, 1996. [https://doi.org/10.1016/0014-5793\(96\)00174-3](https://doi.org/10.1016/0014-5793(96)00174-3)

ISHIZAKI, K. Evolution of land plants: Insights from molecular studies on basal lineages.

Bioscience, Biotechnology and Biochemistry, v. 81, n. 1, p. 73–80, 2017.

<https://doi.org/10.1080/09168451.2016.1224641>

KARLSSON, M.; STENLID, J. Evolution of family 18 glycoside hydrolases: Diversity, domain structures and phylogenetic relationships. **Journal of Molecular Microbiology and Biotechnology**, v. 16, n. 3–4, p. 208–223, 2009. <https://doi.org/10.1159/000151220>

KAUR, R. et al. Glycan-dependent chikungunya viral infection divulged by antiviral activity of NAG specific chi-like lectin. **Virology**, v. 526, p. 91–98, 2019.

<https://doi.org/10.1016/j.virol.2018.10.009>

KAYA, M. et al. On chemistry of γ -chitin. **Carbohydrate Polymers**, v. 176, p. 177–186, 2017. <https://doi.org/10.1016/j.carbpol.2017.08.076>

KESARI, P. et al. Structural and functional evolution of chitinase-like proteins from plants. **Proteomics**, v. 15, n. 10, p. 1693–1705, 2015. <https://doi.org/10.1002/pmic.201400421>

KHAN, F. I. et al. Chitinase from *Thermomyces lanuginosus* SSBP and its biotechnological applications. **Extremophiles**, v. 19, n. 6, p. 1055–1066, 2015.

<https://doi.org/10.1007/s00792-015-0792-8>

LEGER, R. J. S. et al. Characterization and ultrastructural localization of chitinases from *Metarhizium anisopliae*, *M. flavoviride*, and *Beauveria bassiana* during fungal invasion of host (*Manduca sexta*) cuticle. **Applied and Environmental Microbiology**, v. 62, n. 3, p. 907–912, 1996. <https://doi.org/10.1128/AEM.62.3.907-912.1996>

LEWIS, L. A.; MCCOURT, R. M. Green algae and the origin of land plants. **American Journal of Botany**, v. 91, n. 10, p. 1535–1556, 2004. <https://doi.org/10.3732/ajb.91.10.1535>

LOMBARD, V. et al. The carbohydrate-active enzymes database (CAZy) in 2013. **Nucleic Acids Research**, v. 42, n. D1, p. 490–495, 2014. <https://doi.org/10.1093/nar/gkt1178>

MARTIN, W. F.; GARG, S.; ZIMORSKI, V. Endosymbiotic theories for eukaryote origin. **Philosophical transactions of the Royal Society of London, Series B, Biological sciences**, v. 370, n. 1678, p. 1–18, 2015. <https://doi.org/10.1098/rstb.2014.0330>

MARTÍNEZ-CABALLERO, S. et al. Comparative study of two GH19 chitinase-like proteins from *Hevea brasiliensis*, one exhibiting a novel carbohydrate-binding domain. **FEBS Journal**, v. 281, n. 19, p. 4535–4554, 2014. <https://doi.org/10.1111/febs.12962>

MASUDA, T.; ZHAO, G.; MIKAMI, B. Crystal structure of class III chitinase from pomegranate provides the insight into its metal storage capacity. **Bioscience, Biotechnology and Biochemistry**, v. 79, n. 1, p. 45–50, 2015.

<https://doi.org/10.1080/09168451.2014.962475>

MATSUI, M. **Correlações entre estrutura química, superestrutura macromolecular e morfologia das blendas e redes poliméricas à base de quitina e poliuretano**. 2007. Tese (Doutorado em Engenharia) - Programa de Pós-Graduação em Engenharia, Universidade Federal do Paraná, Curitiba, 2007. Disponível em:

<https://acervodigital.ufpr.br/bitstream/handle/1884/16198/MITSUKA%20MATSUI%20.pdf?s>

[equence=1&isAllowed=y](#). Acesso em: 14 ago. 2020.

MCFADDEN, G. I. Origin and evolution of plastids and photosynthesis in eukaryotes. **Cold Spring Harbor Perspectives in Biology**, v. 6, n. 4, p. 1–9, 2014.
<https://doi.org/10.1101/cshperspect.a016105>

MEDEIROS, S. C. et al. Chitinases as antibacterial proteins: A systematic review. **Journal of Young Pharmacists**, v. 10, n. 2, p. 144–148, 2018. <https://doi.org/10.5530/jyp.2018.10.33>

MENDEZ-TEJEDA, R.; ROSADO, J. G. A. Influence of climatic factors on *Sargassum arrivals* to the coasts of the Dominican Republic. **Journal of Oceanography and Marine Science**, v. 10, n. 2, p. 22–32, 2019. <https://doi.org/10.5897/JOMS2019.0156>

MOURA, C. et al. Quitina e quitosana produzidas a partir de resíduos de camarão e siri: avaliação do processo em escala piloto. **Vetor**, v. 16, n. 1–2, p. 37–45, 2006.
<https://periodicos.furg.br/vetor/article/view/294/85>

NEUHAUS, J.-M. et al. A revised nomenclature for chitinase genes. **Plant Molecular Biology Reporter**, v. 14, p. 102–104, 1996. <https://doi.org/10.1007/BF02684897>

NISHIO, J. N. Why are higher plants green? Evolution of the higher plant photosynthetic pigment complement. **Plant, Cell and Environment**, v. 23, n. 6, p. 539–548, 2000.
<https://doi.org/10.1046/j.1365-3040.2000.00563.x>

OHNUMA, T. et al. A class V chitinase from *Arabidopsis thaliana*: Gene responses, enzymatic properties, and crystallographic analysis. **Planta**, v. 234, n. 1, p. 123–137, 2011a.
<https://doi.org/10.1007/s00425-011-1390-3>

OHNUMA, T. et al. Crystal structure and mode of action of a class V chitinase from *Nicotiana tabacum*. **Plant Molecular Biology**, v. 75, n. 3, p. 291–304, 2011b.
<https://doi.org/10.1007/s11103-010-9727-z>

PANAWALA, L. **Difference Between Aerobic and Anaerobic Respiration**, 2017a.
Disponível em:

https://www.researchgate.net/publication/316583908_Difference_Between_Aerobic_and_An_aerobic_respiration. Acesso em: 02 dez. 2020.

PANAWALA, L. **Difference Between Bryophytes and Pteridophytes**, 2017b.
Disponível em:

https://www.researchgate.net/publication/318305486_Difference_Between_Bryophytes_and_Pteridophytes. Acesso em: 02 dez. 2020.

PARK, S. M. et al. Heterologous expression and characterization of class III chitinases from rice (*Oryza sativa* L.). **Enzyme and Microbial Technology**, v. 30, n. 6, p. 697–702, 2002.
[https://doi.org/10.1016/S0141-0229\(02\)00042-X](https://doi.org/10.1016/S0141-0229(02)00042-X)

PATIL, R.; GHORMADE, V.; DESHPANDE, M. Chitinolytic enzymes: An exploration. **Enzyme and Microbial Technology**, v. 26, n. 7, p. 473–483, 2000.
[https://doi.org/10.1016/S0141-0229\(00\)00134-4](https://doi.org/10.1016/S0141-0229(00)00134-4)

- PITTERMANN, J. The evolution of water transport in plants: An integrated approach. **Geobiology**, v. 8, n. 2, p. 112–139, 2010. <https://doi.org/10.1111/j.1472-4669.2010.00232.x>
- PRYER, K. M. et al. Phylogeny and evolution of ferns (monilophytes) with a focus on the early leptosporangiate divergences. **American Journal of Botany**, v. 91, n. 10, p. 1582–1598, 2004. <https://doi.org/10.3732/ajb.91.10.1582>
- PUTTICK, M. N. et al. The interrelationships of land plants and the nature of the ancestral embryophyte. **Current Biology**, v. 28, n. 5, p. 733–745, 2018. <https://doi.org/10.1016/j.cub.2018.01.063>
- RAVEN, J. A. et al. Algal evolution in relation to atmospheric CO₂: Carboxylases, carbon-concentrating mechanisms and carbon oxidation cycles. **Philosophical Transactions of the Royal Society B: Biological Sciences**, v. 367, n. 1588, p. 493–507, 2012. <https://doi.org/10.1098/rstb.2011.0212>
- RAVEN, J. A.; GIORDANO, M. Algae. **Current Biology**, v. 24, n. 13, p. 590–595, 2014. <https://doi.org/10.1016/j.cub.2014.05.039>
- SAMOLOV, E. et al. Biodiversity of algae and cyanobacteria in biological soil crusts collected along a climatic gradient in Chile using an integrative approach. **Microorganisms**, v. 8, n. 7, p. 1–28, 2020. <https://doi.org/10.3390/microorganisms8071047>
- SARMA, K. et al. A comparative proteomic approach to analyse structure, function and evolution of rice chitinases: A step towards increasing plant fungal resistance. **Journal of Molecular Modeling**, v. 18, n. 11, p. 4761–4780, 2012. <https://doi.org/10.1007/s00894-012-1470-8>
- SEIDL, V. et al. A complete survey of Trichoderma chitinases reveals three distinct subgroups of family 18 chitinases. **FEBS Journal**, v. 272, n. 22, p. 5923–5939, 2005. <https://doi.org/10.1111/j.1742-4658.2005.04994.x>
- SEIDL, V. Chitinases of filamentous fungi: A large group of diverse proteins with multiple physiological functions. **Fungal Biology Reviews**, v. 22, n. 1, p. 36–42, 2008. <https://doi.org/10.1016/j.fbr.2008.03.002>
- SELS, J. et al. Plant pathogenesis-related (PR) proteins: A focus on PR peptides. **Plant Physiology and Biochemistry**, v. 46, n. 11, p. 941–950, 2008. <https://doi.org/10.1016/j.plaphy.2008.06.011>
- SU, Y. et al. Identification, phylogeny, and transcript of chitinase family genes in sugarcane. **Scientific Reports**, v. 5, p. 1–15, 2015. <https://doi.org/10.1038/srep10708>
- TRYON, R. M.; TRYON, A. F. **Ferns and allied plants with Special reference to Tropical America**. New York: Springer-Verlag, 1982. <https://doi.org/10.2307/1546858>
- VILLARREAL, J. C. et al. A synthesis of hornwort diversity: Patterns, causes and future work. **Phytotaxa**, v. 9, n. 1, p. 150, 2014. <https://doi.org/10.11646/phytotaxa.9.1.8>

WILLIS, K. J.; MCELWAIN, J. C. The evolution of plants. **Annals of Botany**, v. 90, n. 5, p. 678–679, 2002. <https://doi.org/10.1093/aob/mcf232>

YAN, Q.; FONG, S. S. Bacterial chitinase: Nature and perspectives for sustainable bioproduction. **Bioresources and Bioprocessing**, v. 2, n. 1, 2015. <https://doi.org/10.1186/s40643-015-0057-5>

ANEXOS

ANEXO A - Normas da Revista *Plant Gene*

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