UNIVERSIDADE FEDERAL DE UBERLÂNDIA INSTITUTO DE CIÊNCIAS AGRÁRIAS PROGRAMA DE PÓS-GRADUAÇÃO EM AGRONOMIA

ALEXANDRE MOISÉS ERICSSON DE OLIVEIRA

DISTINCT RECOMBINATION PATTERNS IN GENOMES OF POTYVIRUSES

UBERLÂNDIA MINAS GERAIS – BRASIL 2020

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Tese apresentada à Universidade Federal de Uberlândia, como parte das exigências do Programa de Pós-Graduação em Agronomia, área de concentração em Fitotecnia, para obtenção do título de "Doutor".

Orientador: Prof. Dr^a. Denise Garcia de Santana Coorientador: Prof. Dr. Alison Talis Martins Lima

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RESUMO

A rápida evolução e emergência de vírus de RNA são consequências de suas naturezas propensas à mutação e recombinação. O mecanismo evolutivo de recombinação referese à troca de segmentos de genomas entre vírus distintos durante infecções mistas. A recombinação está frequentemente envolvida no surgimento de vírus com novidades evolutivas, como a capacidade de infectar novos hospedeiros ou suplantar a resistência genética de plantas. Os potyvírus são vírus de RNA de fita simples que causam perdas consideráveis na agricultura em todo o mundo. Estudos indicam que a recombinação intra- e interespecífica pode estar envolvida no surgimento de novas espécies e estirpes de potyvírus. No entanto, se há ou não um padrão de recombinação conservado evolutivamente entre os potyvírus, ainda não foi elucidado. Neste estudo, nós verificamos a existência de "hotspots" de recombinação em genomas de potyvírus. Nove conjuntos de dados de espécies, compostos por aproximadamente 1.500 genomas completos de isolados de potyvírus coletados em todo o mundo, foram obtidos do Genbank. Foram realizadas análises de distribuição de sítios de recombinação para determinar se regiões genômicas específicas são alvo de eventos de recombinação com mais frequência do que esperado ao acaso. O contexto das sequências adjacentes aos "hostpots" de recombinação foi caracterizado por meio de análises de composição de nucleotídeos, predição de estruturas secundárias de RNA e regiões intrinsecamente desordenadas de seus produtos proteicos. "Hotspots" de recombinação foram detectados em três dos nove conjuntos de dados de espécies de potyvírus. Eles foram localizados nas sequências codificadoras das proteínas P3, CI e HC-Pro em genomas de isolados pertencentes às espécies Potato virus Y (PVY), Sugarcane mosaic virus (SCMV) e Turnip mosaic virus (TuMV), respectivamente. As sequências de RNA adjacentes aos "hotspots" apresentaram perfis semelhantes de composição nucleotídica e foram altamente estruturadas em todos os genomas de potyvírus analisados neste estudo. A distribuição uniforme dos sítios de recombinação na maioria dos conjuntos de dados de espécies e a detecção de "hotspots" em posições distintas nos genomas do SCMV, TuMV e PVY indicam que os padrões de recombinação não são conservados entre os potyvírus avaliados. Além disso, a composição de sequências, estruturas secundárias de RNA ou regiões proteicas intrinsecamente desordenadas parecem não explicar os agrupamentos de sítios de recombinação em genomas de vírus potyvírus.

Palavras-chave: Bioinformática. Evolução de vírus. Variabilidade genética. Recombinação.

ABSTRACT

The rapid evolution and emergence of RNA viruses are consequences of their mutation and recombination-prone nature. The evolutionary mechanism of recombination refers to the exchange of genome segments between distinct viruses during mixed infections. Recombination is often involved in the emergence of viruses with evolutionary novelties, such as the ability to infect new hosts or overcome the genetic resistance of plants. Potyviruses are single-stranded (ss)RNA plant viruses that cause considerable losses in agriculture worldwide. Studies indicate that intra- and interspecific recombination may be involved in the emergence of novel potyvirus species and strains. However, whether there is or not an evolutionary conserved recombination pattern among potyviruses remains to be elucidated. In this study, we evaluated the existence of recombination hotspots across genomes of potyviruses. Nine species data sets comprised of about 1,500 complete genomes of potyvirus isolates collected from around the world were retrieved from Genbank. Breakpoint distribution analyses were conducted in order to determine if specific genomic regions are targeted by recombination events more frequently than expected by chance. The sequence context at recombination hotspots was further characterized by nucleotide composition analyses, prediction of RNA secondary structures and intrinsically disordered regions of their protein products. Recombination hotspots were detected into three out of nine potyvirus species data sets. The hotspots were located at P3, CI and HC-Pro encoding sequences in genomes of isolates belonging to the species Potato virus Y (PVY), Sugarcane mosaic virus (SCMV) and Turnip mosaic virus (TuMV), respectively. The RNA sequences surrounding the hotspot positions showed similar profiles of nucleotide composition and were highly structured in all potyvirus genomes analyzed in this study. The even distribution of recombination breakpoints in most species data sets and the detection of hotspots at distinct positions in the SCMV, TuMV and PVY genomes indicate that the recombination patterns are not conserved amongst the evaluated potyviruses. In addition, neither sequence composition, RNA secondary nor intrinsically disordered protein regions seem to explain the breakpoints clustering in genomes of potyviruses.

Keywords: Bioinformatics. Virus evolution. Genetic variability. Recombination.

1 Introduction

Potyviruses are widespread ssRNA plant viruses (genus *Potyvirus*, family *Potyviridae*) encapsidated into flexuous and rod-shaped particles. They are able to infect a wide range of mono- and dicotyledonous plant species leading to considerable losses for agricultural production. They are transmitted by aphids in a non-circulative manner, via infected seeds or propagative plant material, which contributes significantly to their worldwide distribution. The viral RNA genome is about 10,000 nucleotides long, replicates via synthesis of a negative-sense RNA template and encodes a large polyprotein that is self-cleaved into 10 multifunctional proteins. The polyprotein of potyviruses is processed through the following proteases encoded by the viruses: P1, HC-Pro and NIa-Pro. After autocleaving of the polyprotein, multifunctional proteins will be produced in equimolar quantities. However, this production is not dependent on the quantity needed requested by the virus, that is, the production of a certain proteins can be generated in excess may cause accumulation in infected cells (CHUNG et al., 2008; DARÒS; CARRINGTON, 1997; GIBBS et al., 2008; MÄKINEN; HAFRÉN, 2014; URCUQUI-INCHIMA; HAENNI; BERNARDI, 2001; VALLI et al., 2015).

The emergence of novel viruses, alterations in vector-virus specificity, overcoming of genetic resistance are frequently associated with recombination, which might provide substantial advantages in virus ecology and pathogenesis (MARTIN et al., 2011; NORA et al., 2007; SANKARANARAYANAN; PALANI; TENNYSON, 2019). Recent studies also indicates that recombination may be directly involved in overcoming genetic resistance in plants (FENG et al., 2014). Recombination is known

as one of the main mechanisms that affect the evolution of plant viruses and refers to the exchange of genome fragments between viruses from same or even from distinct species. (BENTLEY; EVANS, 2018; BUJARSKI, 2013).

Recombination events occurs during mixed infections when the RNA-dependent RNA polymerase (RDRP) switches from a RNA template molecule to another one, a mechanism called 'copy-choice' (KIRKEGAARD; BALTIMORE, 1986). When recombination events occur at same or similar sites in both parental strands, it is called homologous or precise recombination. On the other hand, when they occurs at different sites of the parental strands, it is named non-homologous or imprecise recombination, which is infrequent in RNA viruses (AUSTERMANN-BUSCH; BECHER, 2012; GALLI; BUKH, 2014; SCHEEL et al., 2013).

In positive ssRNA viruses, recombination events occur at variable frequencies, *e.g.* viruses from the families *Picornaviridae* (SAVOLAINEN-KOPRA; BLOMQVIST, 2010); *Bromoviridae* (URBANOWICZ et al., 2005) and *Potyviridae* (VISSER; BELLSTEDT; PIRIE, 2012) are highly recombinant. However, viruses from *Flaviviridae* (TAUCHER; BERGER; MANDL, 2010) and most (if not all) negative ssRNA viruses (CHARE, 2003) show low frequencies of recombination.

Some studies have focused on nucleotide composition and RNA secondary structures as potential contributors of increased recombination frequencies in specific genomic regions, since they might cause interruptions during replication, favoring the template strand exchange (NAGY; OGIELA; BUJARSKI, 1999). A AU- and GC-rich regions in *Brome mosaic virus* and polioviruses were associated with recombination hotspots (RUNCKEL et al., 2013; SHAPKA; NAGY, 2004). Hairpin loops in tombusviruses (family *Tombusviridae*) seem to favor recombination hotspots (CARPENTER et al., 1995). RNA secondary structures also are involved in increased recombination frequencies in flaviviruses and coronaviruses (families *Flaviviridae* and *Coronaviridae*, respectively) (CHUANG; CHEN, 2009; ROWE et al., 1997).

Viral proteins adopt a well-defined three-dimensional conformation after their translation. However, certain protein regions showing an unstable three-dimensional conformation under natural and biological conditions are known as intrinsically disordered regions (IDRs) (CHARON et al., 2018). Potyvirus proteome indicate a high disorder content, and seems is conserved during potyvirus evolution, suggesting functional advantages of IDR and proteins P1, Coat protein (CP) and Viral genome-linked protein (VPg) present a high content of conserved disorder implying strategies of host machinery hijacking (CHARON et al., 2016). The proteome of potyviruses includes intrinsically disordered proteins, *e.g.* the viral genome-linked protein (VPg), which binds specifically to eIF4E, the mRNAcap-binding protein of the eukaryotic translation initiation complex (WALTER et al., 2019a). Due to the weaker structural requirement of IDRs, mutations have lower impact on virus fitness (WALTER et al., 2019b). However, no studies have established an association between IDRs and increased frequencies of recombination.

In this work, we conducted *in silico* analyses to verify the existence of recombination hotspots in genomes of potyvirus isolates belonging to nine distinct species. In addition, we assessed whether the recombination hotspots were associated

with an unusual nucleotide composition, RNA secondary structure or if there was an overlapping with IDRs. Taken together, our results corroborate the hypothesis that the recombination patterns in potyviruses are by-products of the evolutionary mechanism of selection acting against genomes showing recombination breakpoints within important cistrons for virus infection.

2 Material and methods

2.1 Species data sets and multiple sequence alignments

A total of 1,503 complete genomes of *Bean yellow mosaic virus* (BYMV), *Lettuce mosaic virus* (LMV), *Papaya ringspot virus* (PRSV), *Plum pox virus* (PPV), *Potato virus Y* (PVY), *Soybean mosaic virus* (SMV), *Sugarcane mosaic virus* (SCMV), *Turnip mosaic virus* (TuMV) and *Zucchini yellow mosaic virus* (ZYMV) were retrieved from GenBank using Taxonomy Browser (www.ncbi.nlm.nih.gov) on April 2018. Only potyvirus species for which at least 20 sequences were available in GenBank were analyzed in this study (Table 1 and Supplementary Table S1). Each species data set was aligned separately, using Muscle (EDGAR, 2004) and manually corrected in Mega X (KUMAR et al., 2018).

2.2 Recombination analyses and assessment of genetic variability

Multiple sequence alignments were scanned for recombination using Recombination Detection Program version 4 (RDP4) (MARTIN et al., 2015). Statistical significance was inferred by P-values lower than a Bonferroni-corrected $\alpha = 0.05$ cutoff. Only recombination events detected by at least four of the seven analyses methods implemented in RDP: RDP (MARTIN; RYBICKI, 2000), GENECONV (PADIDAM; SAWYER; FAUQUET, 1999), Chimaera (POSADA; CRANDALL, 2001), MaxChi (SMITH, 1992), BootScan (MARTIN et al., 2005), SiScan (GIBBS; ARMSTRONG; GIBBS, 2000), 3Seq (BONI; POSADA; FELDMAN, 2007) were accounted in the subsequent analyses (RDP project files are available from the authors upon request). The average number of nucleotide differences per site (nucleotide diversity, π) for each species data set was calculated using DnaSP v.6 (ROZAS et al., 2017). Pearson's correlation coefficients between the numbers of recombination events and nucleotide diversity indices were calculated using R-project (R FOUNDATION FOR STATISTICAL COMPUTING, 2018).

2.3 Consensus sequence determination

A consensus sequence for each species data set was determined using Jalview (WATERHOUSE et al., 2009). A similarity matrix was built for each species data set, including the consensus sequence, using the Sequence Demarcation Tool (SDT) program (MUHIRE; VARSANI; MARTIN, 2014). The sequence in the species data set that showed the highest similarity with the consensus sequence was chosen as representative for the subsequent analyses.

2.4 Sequence composition, prediction of RNA secondary structures and IDRs

Sequence composition (or base frequencies) was calculated using Seqool (http://www.biossc.de/seqool/download.html) as percentage of AU or CG dinucleotides. RNAstructure implemented in CLC Genomics Workbench 11.0 (http://resources.qiagenbioinformatics.com/manuals/clcgenomicsworkbench/1101/index .php?manual=Introduction_CLC_Genomics_Workbench.html) was employed for RNA secondary structures prediction. In order to predict IDRs, we use IUPred2A which returns a score from zero to one, corresponding to the probability of the given amino acid residue being part of a disordered region (MÉSZÁROS; ERDÖS; DOSZTÁNYI, 2018).

3 Results

3.1 Distinct recombination patterns detected amongst potyviruses

In this study, we investigated the number of recombination events in data sets composed of genome sequences from nine distinct potyvirus species. The data sets with the largest sample sizes (PPV, PVY and TuMV; N = 219, 358 and 495 sequences, respectively; Table 1) showed different numbers of recombination events (3, 17 and 168 recombination events, respectively; a complete characterization of each recombination event detected in this study is provided in Supplementary Table S2). Although studies indicate a recombination-prone nature of potyvirus genomes, it was notable that only three events were detected in the PPV data set. On the other hand, the TuMV data set showed a considerable larger number of events suggesting to be one of the most recombinant data sets analyzed in this work.

Data sets with intermediate sample sizes (SMV, SCMV and ZYMV; N = 102, 100 and 90 sequences) showed from 11 to 19 recombination events, similar to those

data sets with the smallest sample sizes (BYMV, LMV and PRSV; N = 43, 29 and 67 sequences, respectively).

Previous works indicate that the performance of recombination detection methods is strongly affected by the content of genetic variation in the sequence data set (POSADA, 2002). We validated the results by calculating the Pearson's correlation coefficient, which indicated a negligible (r = -0.18), the coefficient used indicates that there is no linear correlation between the content of genetic variation and the number of recombination events detected by the RDP.

Taking into account our conservative threshold, in which only events detected by at least four out of seven recombination detection methods were accounted, we calculated the number of recombination breakpoints in 200-nucleotide sliding windows to verify the existence of recombination hotspots in viral genomes. Statistically supported recombination hotspots were detected in the PVY, SCMV and TuMV data sets, centered at nucleotide positions of 2,430, 4,800 and 1.650; respectively (Figure 1e, 1g-h). These nucleotide positions were located within the cistrons P3, CI and HC-Pro in PVY, SCMV and TuMV genomes, respectively.

3.2 Sequence elements do not explain the hotspots location

In order to verify if any sequence element (nucleotide composition or RNA secondary structures) could favor the occurrence of recombination hotspots in PVY, SCMV and TuMV genomes, we evaluated the frequency of dinucleotides (AU and GC)

and conducted prediction analyses for detecting RNA secondary structures. Such analyses were based on a single representative sequence from each species data set.

Genomic regions frequently target by recombination events may be associated with unusual composition of nucleotides. In all viruses in which a recombination hotspot was detected by RDP, the variation in the frequency of AU or GC dinucleotides was very wide (Figure 2). More importantly, the recombination hotspots were not associated with regions showing unusual patterns of these dinucleotides. Therefore, the nucleotide composition does not to explain the location of hotspots in PVY, SCMV and TuMV genomes.

We also verified the presence of highly structured regions in the RNA genomes that might also favor high frequencies of recombination. Although the recombination hotspot in the SCMV genome was centered in a highly structured region, the hotspots detected in the PVY and TuMV genomes were located in poorly structured regions (Figure 3). Therefore, the degree of secondary structuring of viral genomes also does not seem to be involved in the location of recombination hotspots.

3.3 Recombination hotspots and intrinsically disordered regions (IDRs) in viral proteins

To test the hypothesis that recombination hotspots are located in sequences that encode protein regions with high level of intrinsic disorder, we conducted a prediction analysis for all proteins of the potyvirus genomes analyzed in this study (Figures 4-6 and Supplementary Figure S2-S7). The recombination hotspots detected in PVY and SCMV genomes overlapped at the N- and C-terminals of the P3 and CI cistrons, respectively. Based on the prediction analysis, both proteins showed low propensity to intrinsic disorder throughout of their lengths. Although some short stretches have been more prone to disorder they not overlapped with recombination hotspots.

On the other hand, the recombination hotspot detected in the region equivalent to the HC-Pro cistron in TuMV genomes (Figure 6) overlapped with a region showing intermediate propensity to intrinsic disorder. Therefore, the lack of a consistent pattern of overlapping between IDRs and recombination hotspots suggest that both features are not correlated in the hotspot virus analyzed.

4 Discussion

Over the last few decades, several studies have shown that recombination events occur at high frequencies in genomes of potyviruses (GREEN; BROWN; KARASEV, 2018; OHSHIMA et al., 2007; SEO et al., 2009). In fact, our results corroborate the recombination-prone nature of some potyviruses, especially for TuMV isolates. In a previous study, recombination hotspots were detected in the P1 and CI-VPg cistrons of 239 TuMV isolates (SHEVCHENKO et al., 2018). However, we conducted a more systematical study in which the existence of recombination hotspots were further validated by permutation tests. The complete genomes of 92 TuMV isolates (50 genomes obtained from samples collected in Europe and Asia and 42 from genomic databases) were analyzed in RDP and 24 recombination events were detected (OHSHIMA et al., 2007). These studies corroborate our findings indicating a recombination-prone nature of TuMV genomes.

The recombination seems to play an important role in the diversification of SCMV isolates. In a study assessing the genetic structure of a SCMV population were detected 20 recombination events in 24 SCMV isolates (XIE et al., 2016). In this study, we detected a recombination hotspot in SCMV genomes. The recombination-prone nature of SCMV genomes is corroborated for another study, in which four out of five isolates collected in distinct countries (USA, Australia, Argentina and Iran) were recombinants (MORADI et al., 2016). A recombination origin for SCMV has been shown in a study based on complete genomes of 20 virus isolates (PADHI; RAMU, 2011).

It is important to emphasize that several recombination events were detected even in those potyvirus species data sets without evidence of a hotspot. In such cases, the recombination events were evenly distributed over the genomes, that is, there was no a specific genomic region targeted by more events than expected by chance. For example, 18 recombination events were reported in a analysis based on 109 BYMV isolates (REVERS et al., 1996).

GC-rich (>60 mol% GC content) and AU-rich (>60 mol% AU) regions are known homologous recombination activators which influence the distribution of events across virus genomes (NAGY; OGIELA; BUJARSKI, 1999). The nucleotide composition at the recombination hotspots detected in PVY, SCMV and TuMV genomes did not showed increased frequency of GC or AU dinucleotides compared to other genome regions. Homologous recombination in genomes of bromoviruses, tombusviruses and carmoviruses is strongly affected by the high content of CG and AU dinucleotides in specific regions of their genomes (NAGY, 1998; NAGY; OGIELA; BUJARSKI, 1999; WHITE; MORRIS, 1994). In a study based on genomes of *Torque teno virus* (TTV) a DNA virus from *Anelloviridae* family, *Alphatorquevirus* genus, recombination events tended to occur within GC-rich regions that have a potential to form secondary structures (LEPPIK et al., 2007). Therefore, our results about nucleotide composition at recombination hotspots of PVY, TuMV and SCMV genomes suggest that other factors might to contribute to the increased numbers of recombination breakpoints.

The contribution of nucleotide composition and RNA secondary structures for increased recombination frequencies at specific regions of PVY genomes has already been tested in previous studies providing contradictory results. A analysis conducted on 43 complete genomes of PVY isolates indicated that the stem–loop structures might favor RNA recombination (HU et al., 2009a). In another study also based on PVY isolates indicated that AU-rich regions and/or RNA secondary structures at some genomic regions might affect the recombination frequency in PVY, but were not the main forces driving the hotspot location (HU et al., 2009b).

Intrinsically disordered regions in proteins are essential for many biological processes. The potyvirus VPg displays many features of a intrinsically disordered protein, since it is involved in several protein-protein interactions during virus infection (JIANG; LALIBERTÉ, 2011; WALTER et al., 2019a). Such regions are more tolerant

to mutations and the presence of disordered segments among plant proteins is associated with the rates of genetic recombination of their encoding genes across the chromosomes of *Arabidopsis thaliana* and *Oryza sativa* indicating a stronger molecular-based dependency of protein disorder and G + C content and much weaker dependency between G + C content and recombination rate in plant genomes. (WALTER et al., 2019b; YRUELA; CONTRERAS-MOREIRA, 2013) We tested if IDRs are also more tolerant to recombination by verifying their overlapping with recombination hotspots in virus genomes. The absence of overlapping indicate that such regions do not tend to concentrate an increased number of recombination breakpoints. Therefore, similar to the results obtained for other viruses (LEFEUVRE et al., 2007; MARTIN et al., 2011), we suggested that the distinct recombination patterns might be a by-product of the natural selection acting against recombinant genomes breakpoints within coding regions essential for virus replication.

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References

AUSTERMANN-BUSCH, S.; BECHER, P. RNA structural elements determine frequency and sites of nonhomologous recombination in an animal plus-strand RNA virus. **Journal of Virology**, v. 86, n. 13, p. 7393-7402, 2012. DOI: 10.1128/JVI.00864-12. <u>https://doi.org/10.1128/JVI.00864-12</u>

BENTLEY, K.; EVANS, D. J. Mechanisms and consequences of positive-strand RNA virus recombination. Journal of General Virology, v. 99, n. 10, p. 1345-1356, 2018. DOI: 10.1099/jgv.0.001142. <u>https://doi.org/10.1099/jgv.0.001142</u>

BONI, M. F.; POSADA, D.; FELDMAN, M. W. An exact nonparametric method for inferring mosaic structure in sequence triplets. **Genetics**, v. 176, n. 2, p. 1035-1047, 2007. DOI: 10.1534/genetics.106.068874. <u>https://doi.org/10.1534/genetics.106.068874</u>

BUJARSKI, J. J. Genetic recombination in plant-infecting messenger-sense RNA viruses: overview and research perspectives. **Frontiers in Plant Science**, v. 4, n. MAR, p. 1-10, 2013. DOI: 10.3389/fpls.2013.00068. <u>https://doi.org/10.3389/fpls.2013.00068</u>

CARPENTER, C. D. *et al.* Involvement of a stem-loop structure in the location of junction sites in viral rna recombination. **Journal of Molecular Biology**, v. 245, n. 5,p. 608-622, 1995. <u>https://doi.org/10.1006/jmbi.1994.0050</u>

CHARE, E. R. Phylogenetic analysis reveals a low rate of homologous recombination in negative-sense RNA viruses. Journal of General Virology, v. 84, n. 10, p. 2691-2703, out. 2003. DOI: 10.1099/vir.0.19277-0. <u>https://doi.org/10.1099/vir.0.19277-0</u>

CHARON, J. *et al.* Protein intrinsic disorder within the Potyvirus genus: from proteome-wide analysis to functional annotation. **Molecular BioSystems**, v. 12, n. 2, p. 634-652, 2016. DOI: 10.1039/C5MB00677E. <u>https://doi.org/10.1039/C5MB00677E</u>

CHARON, J. *et al.* First Experimental assessment of protein intrinsic disorder involvement in an RNA virus natural adaptive process. **Molecular Biology and Evolution**, v. 35, n. 1, p. 38-49, 2018. DOI: 10.1093/molbev/msx249 https://doi.org/10.1093/molbev/msx249

CHUANG, C.-K.; CHEN, W.-J. Experimental evidence that RNA recombination occurs in the Japanese encephalitis virus. **Virology**, v. 394, n. 2, p. 286-297, nov. 2009. DOI: 10.1016/j.virol.2009.08.030. <u>https://doi.org/10.1016/j.virol.2009.08.030</u>

CHUNG, B. Y. W. *et al.* An overlapping essential gene in the Potyviridae. **Proceedings of the National Academy of Sciences**, v. 105, n. 15, p. 5897-5902, abr. 2008. DOI: 10.1073/pnas.0800468105. <u>https://doi.org/10.1073/pnas.0800468105</u>

DARÒS, J. A.; CARRINGTON, J. C. RNA binding activity of NIa proteinase of tobacco etch potyvirus. **Virology**, v. 237, n. 2, p. 327-336, 1997. DOI: 10.1006/viro.1997.8802. <u>https://doi.org/10.1006/viro.1997.8802</u>

EDGAR, R. C. MUSCLE: multiple sequence alignment with high accuracy and high throughput. **Nucleic acids research**, v. 32, n. 5, p. 1792-7, 2004. DOI: 10.1093/nar/gkh340. <u>https://doi.org/10.1093/nar/gkh340</u>

FENG, X. *et al.* Recombinants of Bean common mosaic virus (BCMV) and genetic determinants of BCMV involved in overcoming resistance in Common Bean. **Phytopathology**, v. 104, n. 7, p. 786-793, 2014. DOI: 10.1094/PHYTO-08-13-0243-R. <u>https://doi.org/10.1094/PHYTO-08-13-0243-R</u>

GALLI, A.; BUKH, J. Comparative analysis of the molecular mechanisms of recombination in hepatitis C virus. **Trends in Microbiology**, v. 22, n. 6, p. 354-364, 2014. DOI: 10.1016/j.tim.2014.02.005. <u>https://doi.org/10.1016/j.tim.2014.02.005</u>

GIBBS, A. J. *et al.* The prehistory of Potyviruses : their initial radiation was during the dawn of agriculture. **PLoS ONE**, v. 3, n. 6, 2008. DOI: 10.1371/journal.pone.0002523. https://doi.org/10.1371/journal.pone.0002523

GIBBS, M. J.; ARMSTRONG, J. S.; GIBBS, A. J. Sister-scanning: A Monte Carlo procedure for assessing signals in rebombinant sequences. **Bioinformatics**, v. 16, n. 7, p. 573-582, 2000. <u>https://doi.org/10.1093/bioinformatics/16.7.573</u>

GREEN, K. J.; BROWN, C. J.; KARASEV, A. V. Genetic diversity of potato virus Y (PVY): sequence analyses reveal ten novel PVY recombinant structures. Archives of **Virology**, v. 163, n. 1, p. 23-32, 2018. DOI: 10.1007/s00705-017-3568-x. https://doi.org/10.1007/s00705-017-3568-x

HU, X. *et al.* Molecular characterization and detection of recombinant isolates of potato virus Y from China. **Archives of Virology**, v. 154, n. 8, p. 1303-1312, 2009a. DOI: 10.1007/s00705-009-0448-z. <u>https://doi.org/10.1007/s00705-009-0448-z</u>

HU, X. *et al.* Sequence characteristics of potato virus Y recombinants. Journal of General Virology, v. 90, n. 12, p. 3033-3041, 2009b. DOI: 10.1099/vir.0.014142-0. https://doi.org/10.1099/vir.0.014142-0 JIANG, J.; LALIBERTÉ, J. F. The genome-linked protein VPg of plant viruses - A protein with many partners. **Current Opinion in Virology**, v. 1, n. 5, p. 347-354, 2011. DOI: 10.1016/j.coviro.2011.09.010. <u>https://doi.org/10.1016/j.coviro.2011.09.010</u>

KIRKEGAARD, K.; BALTIMORE, D. The mechanism of RNA recombination in poliovirus. **Cell**, v. 47, n. 3, p. 433-443, 1986. DOI: 10.1016/0092-8674(86)90600-8. https://doi.org/10.1016/0092-8674(86)90600-8

KUMAR, S. *et al.* MEGA X: Molecular evolutionary genetics analysis across computing platforms. **Molecular Biology and Evolution**, v. 35, n. 6, p. 1547-1549, 2018. DOI: 10.1093/molbev/msy096. <u>https://doi.org/10.1093/molbev/msy096</u>

LEFEUVRE, P. *et al.* Avoidance of protein fold disruption in natural virus recombinants. **PLoS Pathogens**, v. 3, n. 11, p. e181, 2007. DOI: 10.1371/journal.ppat.0030181. <u>https://doi.org/10.1371/journal.ppat.0030181</u>

LEPPIK, L. *et al.* In vivo and in vitro intragenomic rearrangement of TT viruses. **Journal of Virology**, v. 81, n. 17, p. 9346-9356, 2007. DOI: 10.1128/JVI.00781-07. https://doi.org/10.1128/JVI.00781-07

MÄKINEN, K.; HAFRÉN, A. Intracellular coordination of potyviral RNA functions in infection. **Frontiers in Plant Science**, v. 5, n. MAR, p. 1-13, 2014. DOI: 10.3389/fpls.2014.00110. <u>https://doi.org/10.3389/fpls.2014.00110</u>

MARTIN, D. P. *et al.* A modified bootscan algorithm for automated identification of recombinant sequences and recombination breakpoints. **AIDS Research and Human Retroviruses**, v. 21, n. 1, p. 98-102, 2005. DOI: 10.1089/aid.2005.21.98. https://doi.org/10.1089/aid.2005.21.98

MARTIN, D. P. *et al.* Recombination in eukaryotic single stranded DNA viruses. **Viruses**, v. 3, n. 9, p. 1699-1738, 2011. DOI: 10.3390/v3091699. https://doi.org/10.3390/v3091699

MARTIN, D. P. *et al.* RDP4: Detection and analysis of recombination patterns in virus genomes. **Virus Evolution**, v. 1, n. 1, p. 1-5, 2015. DOI: 10.1093/ve/vev003. https://doi.org/10.1093/ve/vev003

MARTIN, D.; RYBICKI, E. RDP: detection of recombination amongst aligned sequences. **Bioinformatics**, v. 16, n. 6, p. 562-563, 2000. DOI: 10.1093/bioinformatics/16.6.562. <u>https://doi.org/10.1093/bioinformatics/16.6.562</u>

MÉSZÁROS, B.; ERDÖS, G.; DOSZTÁNYI, Z. IUPred2A: context-dependent prediction of protein disorder as a function of redox state and protein binding. Nucleic **Acids Research**, v. 46, n. W1, p. W329-W337, 2018. DOI: 10.1093/nar/gky384. https://doi.org/10.1093/nar/gky384

MORADI, Z. *et al.* The complete genome sequences of two naturally occurring recombinant isolates of Sugarcane mosaic virus from Iran. **Virus Genes**, v. 52, n. 2, p. 270-280, abr. 2016. DOI: 10.1007/s11262-016-1302-5. <u>https://doi.org/10.1007/s11262-016-1302-5</u>

MUHIRE, B. M.; VARSANI, A.; MARTIN, D. P. SDT: a virus classification tool based on pairwise sequence alignment and identity calculation. **PLoS ONE**, v. 9, n. 9, 2014. <u>https://doi.org/10.1371/journal.pone.0108277</u>

NAGY, P. D. Dissecting RNA recombination invitro: role of RNA sequences and the viral replicase. **The EMBO Journal**, v. 17, n. 8, p. 2392-2403, abr. 1998. DOI: 10.1093/emboj/17.8.2392. <u>https://doi.org/10.1093/emboj/17.8.2392</u>

NAGY, P. D.; OGIELA, C.; BUJARSKI, J. J. Mapping sequences active in homologous RNA recombination in brome mosaic virus: Prediction of recombination hot spots. **Virology**, v. 254, n. 1, p. 92-104, 1999. DOI: 10.1006/viro.1998.9545. https://doi.org/10.1006/viro.1998.9545

NORA, T. *et al.* Contribution of recombination to the rvolution of human immunodeficiency viruses expressing resistance to antiretroviral treatment. **Journal of Virology**, v. 81, n. 14, p. 7620-7628, 2007. DOI: 10.1128/JVI.00083-07. https://doi.org/10.1128/JVI.00083-07

OHSHIMA, K. *et al.* Patterns of recombination in turnip mosaic virus genomic sequences indicate hotspots of recombination. Journal of General Virology, v. 88, n. 1, p. 298-315, 2007. DOI: 10.1099/vir.0.82335-0. <u>https://doi.org/10.1099/vir.0.82335-0</u>

PADHI, A.; RAMU, K. Genomic evidence of intraspecific recombination in sugarcane mosaic virus. **Virus Genes**, v. 42, n. 2, p. 282-285, 2011. DOI: 10.1007/s11262-010-0564-6. <u>https://doi.org/10.1007/s11262-010-0564-6</u>

PADIDAM, M.; SAWYER, S.; FAUQUET, C. M. Possible emergence of new geminiviruses by frequent recombination. Virology, v. 265, n. 2, p. 218-225, 1999. DOI: 10.1006/viro.1999.0056. <u>https://doi.org/10.1006/viro.1999.0056</u>

POSADA, D. Evaluation of methods for detecting recombination from DNA sequences: empirical data. **Molecular Biology and Evolution**, v. 19, n. 5, p. 708-717, 2002. <u>https://doi.org/10.1093/oxfordjournals.molbev.a004129</u>

POSADA, D.; CRANDALL, K. A. 2012. Evaluation of methods for detecting recombination from DNA sequences: computer simulations. **Proceedings of the National Academy of Science,** v. 98, n. 24, 2001. DOI: 10.1073/pnas.241370698. https://doi.org/ pnas.org/content/98/24/13757

REVERS, F. *et al.* Frequent occurrence of recombinant potyvirus isolates. Journal of General Virology, v. 77, n. 8, p. 1953-1965, 1996. DOI: 10.1099/0022-1317-77-8-1953. <u>https://doi.org/10.1099/0022-1317-77-8-1953</u>

ROWE, C. L. *et al.* Generation of coronavirus spike deletion variants by high-frequency recombination at regions of predicted RNA secondary structure. **Journal of virology**, v. 71, n. 8, p. 6183-6190, 1997. DOI: 10.1128/JVI.71.8.6183-6190.1997. https://doi.org/10.1128/JVI.71.8.6183-6190.1997

ROZAS, J. *et al.* DnaSP 6: DNA sequence polymorphism analysis of large data sets. **Molecular Biology and Evolution**, v. 34, n. 12, p. 3299-3302, dez. 2017. DOI: 10.1093/molbev/msx248. <u>https://doi.org/10.1093/molbev/msx248</u>

RUNCKEL, C. *et al.* Identification and manipulation of the molecular determinants influencing poliovirus recombination. **PLoS Pathogens**, v. 9, n. 2, 2013. DOI: 10.1371/journal.ppat.1003164. <u>https://doi.org/10.1371/journal.ppat.1003164</u>

SANKARANARAYANAN, R.; PALANI, S. N.; TENNYSON, J. Hubei poty-like virus 1 is likely an interspecies recombinant of sugarcane mosaic virus and putative bean yellow mosaic virus. **Archives of Virology**, v. 164, n. 1, p. 261-265, 2019. DOI: 10.1007/s00705-018-4047-8. <u>https://doi.org/10.1007/s00705-018-4047-8</u>

SAVOLAINEN-KOPRA, C.; BLOMQVIST, S. Mechanisms of genetic variation in polioviruses. **Reviwes in Medical Virology**, n. October, p. 358-371, 2010. DOI: 10.1002/rmv.663. <u>https://doi.org/10.1002/rmv.663</u>

SCHEEL, T. K. H. *et al.* Productive homologous and non-homologous recombination of Hepatitis C virus in cell culture. **Plos Pathogens**, v. 9, n. 3, 2013. DOI: 10.1371/journal.ppat.1003228. <u>https://doi.org/10.1371/journal.ppat.1003228</u>

SEO, J.-K. *et al.* Molecular variability and genetic structure of the population of Soybean mosaic virus based on the analysis of complete genome sequences. Virology,

v. 393, n. 1, p. 91-103, out. 2009. DOI: 10.1016/j.virol.2009.07.007. https://doi.org/10.1016/j.virol.2009.07.007

SHAPKA, N.; NAGY, P. D. The AU-rich RNA recombination hot spot sequence of brome mosaic virus is functional in Tombusviruses: implications for the mechanism of rna recombination. **Journal of Virology**, v. 78, n. 5, p. 2288-2300, mar. 2004. DOI: 10.1128/JVI.78.5.2288-2300.2004. https://doi.org/10.1128/JVI.78.5.2288-2300.2004

SHEVCHENKO, O. *et al.* First evidence of the occurrence of Turnip mosaic virus in Ukraine and molecular characterization of its isolate. **Journal of Phytopathology**, v. 166, n. 6, p. 429-437, 2018. <u>https://doi.org/10.1111/jph.12703</u>

SMITH, J. M. Analyzing the mosaic structure of genes. Journal of Molecular Evolution, v. 34, n. 2, p. 126-129, 1992. DOI: 10.1007/BF00182389. https://doi.org/10.1007/BF00182389

TAUCHER, C.; BERGER, A.; MANDL, C. W. A trans-complementing recombination trap demonstrates a low propensity of flaviviruses for intermolecular recombination. **Journal of Virology**, v. 84, n. 1, p. 599-611, jan. 2010. DOI: 10.1128/JVI.01063-09. https://doi.org/10.1128/JVI.01063-09

URBANOWICZ, A. *et al.* Homologous crossovers among molecules of Brome Mosaic Bromovirus RNA1 or RNA2 segments in vivo. **Journal of Virology**, v. 79, n. 9, p. 5732-5742, maio 2005. DOI: 10.1128/JVI.79.9.5732-5742.2005. https://doi.org/10.1128/JVI.79.9.5732-5742.2005

URCUQUI-INCHIMA, S.; HAENNI, A. L.; BERNARDI, F. Potyvirus proteins: A wealth of functions. Virus Research, v. 74, n. 1-2, p. 157-175, 2001. DOI: 10.1016/S0168-1702(01)00220-9. <u>https://doi.org/10.1016/S0168-1702(01)00220-9</u>

VALLI, A. *et al.* RNA Polymerase slippage as a mechanism for the production of frameshift gene products in plant viruses of the Potyviridae family. **Journal of Virology**, v. 89, n. 13, p. 6965-6967, 2015. DOI: 10.1128/JVI.00337-15. https://doi.org/10.1128/JVI.00337-15

VISSER, J. C.; BELLSTEDT, D. U.; PIRIE, M. D. The recent recombinant evolution of a major crop pathogen, Potato virus Y. **PLoS ONE**, v. 7, n. 11, 2012. DOI: 10.1371/journal.pone.0050631. <u>https://doi.org/10.1371/journal.pone.0050631</u>

WALTER, J. et al. Hydrodynamic behavior of the intrinsically disordered potyvirus protein VPg, of the translation initiation factor eIF4E and of their binary complex.

International Journal of Molecular Sciences, v. 20, n. 7, p. 1794, 2019a. DOI: 10.3390/ijms20071794. <u>https://doi.org/10.3390/ijms20071794</u>

WALTER, J. *et al.* Comparative analysis of mutational robustness of the intrinsically disordered viral protein VPg and of its interactor eIF4E. **PLOS ONE**, v. 14, n. 2, p. e0211725, fev. 2019b. DOI: 10.1371/journal.pone.0211725. https://doi.org/10.1371/journal.pone.0211725

WATERHOUSE, A. M. et al. Jalview Version 2--a multiple sequence alignment editor and analysis workbench. **Bioinformatics**, v. 25, n. 9, p. 1189-1191, maio 2009. DOI: 10.1093/bioinformatics/btp033. <u>https://doi.org/10.1093/bioinformatics/btp033</u>

WHITE, K. A.; MORRIS, T. J. Recombination between defective tombusvirus RNAs generates functional hybrid genomes. **Proceedings of the National Academy of Sciences**, v. 91, n. 9, p. 3642-3646, abr. 1994. DOI: 10.1073/pnas.91.9.3642. https://doi.org/10.1073/pnas.91.9.3642

XIE, X. *et al.* Molecular variability and distribution of Sugarcane mosaic virus in Shanxi, China. **PLoS ONE**, v. 11, n. 3, p. 1-12, 2016. DOI: 10.1371/journal.pone.0151549. <u>https://doi.org/10.1371/journal.pone.0151549</u>

YRUELA, I.; CONTRERAS-MOREIRA, B. Genetic recombination is associated with intrinsic disorder in plant proteomes. **BMC Genomics**, v. 14, n. 1, 2013. DOI: 10.1186/1471-2164-14-772. <u>https://doi.org/10.1186/1471-2164-14-772</u>

Species dataset	Sample size (N)	Recombination events	Nucleotide Diversity (Pi)	Standard deviation
BYMV	43	11	0,11	0,010
LMV	29	11	0,15	0,050
PRSV	67	17	0,19	0,010
PPV	219	3	0,10	0,005
PVY	358	17	0,07	0,002
SMV	102	19	0,68	0,006
SCMV	100	13	0,54	0,020
TuMV	495	168	0,15	0,003
ZYMV	90	11	0,68	0,007

Table 1. Sample size, recombination events and nucleotide diversity of potyvirus species data sets retrieved from Genbank.

BYMV, Bean yellow mosaic virus; LMV, Lettuce mosaic virus; PRSV, Papaya ringspot virus; PPV, Plum pox virus; PVY, Potato virus Y; SMV, Soybean mosaic virus; SCMV, Sugarcane mosaic virus; TuMV, Turnip mosaic virus; ZYMV, Zucchini mosaic virus.

Figure legends

Figure 1. Breakpoints density across complete genomes of potyviruses from nine distinct species: (a) *Bean yellow mosaic virus*, (b) *Lettuce mosaic virus*, (c) *Papaya ringspot virus* (d) *Plum pox virus*, (e) *Potato virus* Y (f) *Soybean mosaic virus* (g) *Sugarcane mosaic virus*, (h) *Turnip mosaic virus* (i) *Zucchini mosaic virus*. A 200-nucleotide window was moved along the alignment, one nucleotide at time, and the breakpoints detected within the window length were counted and plotted (solid line). The upper and lower broken lines, respectively, indicate 99% and 95% confidence thresholds for globally significant breakpoint clusters. The light- and dark-gray areas, respectively, indicate local 99% and 95% breakpoint-clustering thresholds, taking into account local regional differences in sequence diversity that influence the abilities of different methods to detect recombination breakpoints.

Figure 2. Sequence composition (AU and GC dinucleotide frequencies) across genomes of (a) PVY, (b) SCMV and (c) TuMV. The hotspot position is indicated by the sequence position highlighted in red.

Figure 3. Prediction of RNA secondary structures based on complete genomes (**a**, **c** and **e**) and the hotspot surrounding sequences (**b**, **d** and **f**) of PVY (KY847956), SCMV (JX047385) and TuMV (AB194785), respectively. The kind of secondary structure predicted for each genome region in **b**, **d** and **f** is represented by a distinct color: stem (purple), bulge (green), multiloop (blue), interior loop (brown). The nucleotide position of the hotspots are highlighted in red.

Figure 4. Prediction of intrinsically disordered regions (IDRs) conducted for PVY proteins. The red bar indicates the hotspot position.

Figure 5. Prediction of intrinsically disordered regions (IDRs) conducted for SCMV proteins. The red bar indicates the hotspot position.

Figure 6. Prediction of intrinsically disordered regions (IDRs) conducted for TuMV proteins. The red bar indicates the hotspot position.





















Figure 4


Figure 5







Supplementary online Material

Supplementary Table S1. Genome sequences of potyviruses used in this study obtained from GenBank

Virus	GeenBank Acession	Host	Collected date	Geografical Origin
Turnip mosaic virus (TuMV)	AY134473	Brassica. napus	2003	UK
	AF530055	Zantedeschia spp.	2003	Taiwan
	MG200170	Raphanus sativus	2016	South Korea
	MG200169	Raphanus sativus	2016	South Korea
	MG200168	Raphanus sativus	2016	South Korea
	MG200167	Raphanus sativus	2016	South Korea
	MG200166	Raphanus sativus	2016	South Korea
	KX674734	Raphanus sativus	2015	South Korea
	KY190216	Physalis ixocarpa	2016	Mexico
	KY111274	Raphanus sativus	2015	South Korea
	KY111273	Raphanus sativus	2015	South Korea
	KY111272	Raphanus sativus	2015	South Korea
	KY111271	Raphanus sativus	2015	South Korea
	KY111270	Raphanus sativus	2015	South Korea
	KY111269	Raphanus sativus	2015	South Korea
	KY111268	Raphanus sativus	2015	South Korea
	KY111267	Raphanus sativus	2015	South Korea
	KX674733	Raphanus sativus	2015	South Korea
	KX674732	Raphanus sativus	2015	South Korea
	KX674731	Raphanus sativus	2015	South Korea
	KX674730	Raphanus sativus	2015	South Korea
	KX674729	Raphanus sativus	2015	South Korea

KX674728	Raphanus sativus	2015	South Korea
KX674727	Raphanus sativus	2015	South Korea
KX674726	Raphanus sativus	2015	South Korea
LC215859	Raphanus sativus	1/15/2014	South Korea
KX610932	Brassica oleracea	2015	Australia
KX610931	Brassica oleracea	2015	Australia
KX610930	Brassica oleracea	2015	Australia
KX610929	Brassica oleracea	2015	Australia
KX641466	Arabidopsis sp.	14-Apr-15	Australia
KX641465	Chinese cabbage	14-May-97	Australia
KX579486	Raphanus sativus var. hortensis f. raphanistroides	1-Mar-14	South Korea
KX579485	Raphanus sativus var. hortensis f. raphanistroides	1-Mar-14	South Korea
KX579484	Raphanus sativus var. hortensis f. raphanistroides	1-Mar-14	South Korea
KX579483	Raphanus sativus var. hortensis f. raphanistroides	1-Mar-14	South Korea
KX579482	Raphanus sativus var. hortensis f. raphanistroides	1-Mar-14	South Korea
KX579481	Raphanus sativus var. hortensis f. raphanistroides	1-Mar-14	South Korea
KX579480	Raphanus sativus var. hortensis f. raphanistroides	1-Mar-14	South Korea
KX579479	Raphanus sativus var. hortensis f. raphanistroides	1-Mar-14	South Korea
KM094174	Raphanus sativus	1995	Japan
KJ936093	Raphanus raphanistrum	2000	Australia
KJ936092	Rapistrum rugosum	2006	Australia
KJ936091	Hirschfeldia incana	2006	Australia
KJ936090	Rapistrum rugosum	2006	Australia
KJ936089	Cicer arietinum	2006	Australia
KJ936088	Brassica juncea	2006	Australia
KJ936087	Brassica juncea	2006	Australia

KF595121	Brassica napus L. ssp. oleifera (DC.) Metzg.	2009	Croatia
KF246570	Phalaenopsis sp.	2012	China
KC297103	Brassica oleracea	2012	Russia
AB747315	Brassica juncea	2006	Vietnam
AB747314	Brassica juncea	2006	Vietnam
AB747313	Brassica juncea	2006	Vietnam
AB747312	Brassica juncea	2006	Vietnam
AB747311	Brassica juncea	2006	Vietnam
AB747310	Brassica juncea	2006	Vietnam
AB747309	Brassica juncea	2006	Vietnam
AB747308	Brassica juncea	2006	Vietnam
AB747307	Brassica juncea	2006	Vietnam
AB747306	Brassica juncea	2006	Vietnam
AB747305	Brassica juncea	2006	Vietnam
AB747304	Brassica juncea	2006	Vietnam
AB747303	Brassica juncea	2006	Vietnam
AB747302	Brassica juncea	2006	Vietnam
AB747301	Brassica juncea	2006	Vietnam
AB747300	Brassica juncea	2006	Vietnam
AB747299	Brassica juncea	2006	Vietnam
AB747298	Brassica juncea	2006	Vietnam
AB747297	Brassica juncea	2006	Vietnam
AB747296	Raphanus sativus	2006	Vietnam
AB747295	Raphanus sativus	2006	Vietnam
AB747294	Raphanus sativus	2006	Vietnam
AB747293	Raphanus sativus	2006	Vietnam

AB747292	Raphanus sativus	2006	Vietnam
AB747291	Raphanus sativus	2006	Vietnam
AB747290	Raphanus sativus	2006	Vietnam
AB747289	Raphanus sativus	2006	Vietnam
AB747288	Raphanus sativus	2006	Vietnam
AB747287	Raphanus sativus	2006	Vietnam
AB747286	Raphanus sativus	2006	Vietnam
HM544042	Brassica rapa	2007	Australia: Brisbane
AB440239	Brassica deflexa	2003-2008	Iran
AB440238	Brassica deflexa	2003-2008	Iran
EU734434	Raphanus sativus	2008	China
EU734433	Raphanus sativus	2008	China
AF169561	Brassica napus	2000	UK
AY090660	Brassica napus	2002	China
DQ648592	Cochlearia armoracia L.	2004	Poland
DQ648591	Cochlearia armoracia L.	2004	Poland
EU861593	Brassica oleracea var. oleracea	2002	UK
AB362513	Brassicaceae	2006	Turkey
AB362512	Brassicaceae	2006	Turkey
AB105135	Raphanus sativus L	2003	Japan
AB105134	Brassica oleracea L.	2003	Japan
D10927	Brassica napus	1990	Canada
AB252143	Raphanus sativus	2000	Japan
AB252142	Raphanus sativus	2000	Japan
AB252141	Lactuca sativa	2000	Japan
AB252140	Raphanus sativus	2000	Japan

AB252139	Raphanus sativus	2000	Japan
AB252138	Raphanus sativus	2000	Japan
AB252137	Raphanus sativus	2000	Japan
AB252136	Brassica rapa	2000	Japan
AB252135	Ranunculus asiaticus	1997	Italy
AB252134	Raphanus. sativus	2001	Japan
AB252133	Brassica oleracea	1995	Netherlands
AB252132	Raphanus sativus	1998	Japan
AB252131	Raphanus sativus	2000	Japan
AB252130	Raphanus sativus	1998	Japan
AB252129	Raphanus sativus	2000	Japan
AB252128	Raphanus sativus	2000	Japan
AB252127	Raphanus sativus	2001	Japan
AB252126	Raphanus sativus	2000	Japan
AB252125	Brassica rapa	2004	Japan
AB252124	Brassica oleracea	2004	Japan
AB252123	Raphanus sativus	1998	Japan
AB252122	Brassica sp.	1994	Italy
AB252121	Raphanus sativus	2000	Japan
AB252120	Raphanus sativus	2000	Japan
AB252119	Brassica sp.	1998	China
AB252118	Raphanus sativus	1996	Japan
AB252117	Allium sp.	1999	Greece
AB252116	Brassica oleracea	1993	Greece
AB252115	Raphanus sativus	2001	Japan
AB252114	Brassica oleracea	2000	UK

AB252113	Brassica oleracea	1999	UK
AB252112	Brassica oleracea	1987	Germany
AB252111	Brassica pekinensis	1998	Japan
AB252110	Raphanus sativus	2000	Japan
AB252109	Raphanus sativus	2000	Japan
AB252108	Brassica napus	1993	Denmark
AB252107	Brassica. rapa	1994	Czech Republic
AB252106	Brassica campestris	1999	China
AB252105	Raphanus sativus	1999	China
AB252104	Raphanus sativus	2000	China
AB252103	Raphanus sativus	1999	China
AB252102	Eustoma russellianum	1998	Japan
AB252101	Brassica. pekinensis	2000	Japan
AB252100	Raphanus sativus	2000	Japan
AB252099	Raphanus sativus	1998	Japan
AB252098	Raphanus sativus	2002	Japan
AB252097	Raphanus sativus	2002	Japan
AB252096	Raphanus sativus	2002	Japan
AB252095	Raphanus sativus	1998	Japan
AB252094	Raphanus sativus	1998	Japan
AB194802	Raphanus sativus	2004	Spain
AB194801	Raphanus sativus	2004	Spain
AB194800	Raphanus sativus	2004	Spain
AB194799	Raphanus sativus	2004	Spain
AB194798	Raphanus sativus	2004	Spain
AB194797	Raphanus. sativus	2004	Spain

AB194796	Raphanus sativus	2004	Spain
AB194795	Raphanus sativus	2004	Spain
AB194794	Raphanus sativus	2004	Spain
AB194793	Raphanus sativus	2004	Spain
AB194792	Raphanus sativus	2004	Spain
AB194791	Raphanus sativus	2004	Spain
AB194790	Raphanus sativus	2004	Spain
AB194789	Raphanus sativus	2004	Spain
AB194788	Raphanus sativus	2004	Spain
AB194787	Raphanus sativus	2004	Spain
AB194786	Raphanus sativus	2004	Spain
AB194785	Raphanus sativus	2004	Spain
AB093627	Raphanus sativus	1998	China
AB093626	Brassica spp.	1980	China
AB093625	Brassica campestris	1993	Japan
AB093624	Brassica pekinensis	1994	Japan
AB093623	Raphanus sativus	1993	Japan
AB093622	Brassica pekinensis	1994	Japan
AB093621	Raphanus sativus	1998	Japan
AB093620	Raphanus sativus	1996	Japan
AB093619	Raphanus sativus	1998	Japan
AB093618	Raphanus sativus	1998	Japan
AB093617	Raphanus sativus	1998	Japan
AB093616	Raphanus sativus	1997	Japan
AB093615	Raphanus sativus	1998	Japan
AB093614	Calendula officinalis	1997	Japan

AB093613	Raphanus sativus	1998	Japan
AB093612	Brassica pekinensis	1998	New Zealand
AB093611	Brassica oleracea	1996	Brazil
AB093610	Brassica napus	1989	Canada
AB093609	Brassica oleracea	1980	United States
AB093608	Brassica oleracea	1981	Czech Republic
AB093607	Brassica napus	Unknow	Russia
AB093606	Armoracia rusticana	1993	Russia
AB093605	Brassica oleracea	1994	Kenya
AB093604	Brassica napus	1970	Germany
AB093603	Lactuca sativa	1993	Germany
AB093602	Allium ampeloprasum	1993	Israel
AB093601	Calendula officinalis	1979	Italy
AB093600	Raphanus sativus	1994	Italy
AB093599	Anemone coronaria	1991	Italy
AB093598	Alliaria officinalis	1968	Italy
AB093597	Anemone coronaria	1993	Italy
AB093596	Limonium sinuatum	1993	Italy
D83184	Brassica rapa	1996	Japan
KX377967	Raphanus sativus	2017	Spain
AP017890	Brassica oleracea	1993-2012	Turkey
AP017889	Brassica oleracea	1993-2012	Turkey
AP017888	Raphanus raphanistrum	1993-2012	Turkey
AP017887	Raphanus raphanistrum	1993-2012	Turkey
AP017886	Brassica oleracea	1993–2012	Turkey
AP017885	Brassica oleracea	1993-2012	Turkey

AP017884	Raphanus sativus	1993–2012	Turkey
AP017883	Brassica oleracea var. botrytis	1993–2012	Turkey
AP017882	Brassica oleracea	1993–2012	Turkey
AP017881	Brassica oleracea	1993-2012	Turkey
AP017880	Brassica oleracea	1993–2012	Turkey
AP017879	Brassica oleracea	1993–2012	Turkey
AP017878	Raphanus raphanistrum	1993–2012	Turkey
AP017877	Raphanus raphanistrum	1993–2012	Turkey
AP017876	Brassica oleracea	1993–2012	Turkey
AP017875	Brassica oleracea	1993–2012	Turkey
AP017874	Brassica rapa	1993–2012	Turkey
AP017873	Sinapis arvensis	1993–2012	Turkey
AP017872	Raphanus raphanistrum	1993–2012	Turkey
AP017871	Raphanus raphanistrum	1993–2012	Turkey
AP017870	Brassica spp.	1993–2012	Turkey
AP017869	Brassica spp.	1993–2012	Turkey
AP017868	Raphanus raphanistrum	1993–2012	Turkey
AP017867	Raphanus raphanistrum	1993–2012	Turkey
AP017866	Raphanus sativus	1993–2012	Turkey
AP017865	Raphanus sativus	1993–2012	Turkey
AP017864	Raphanus sativus	1993–2012	Turkey
AP017863	Brassica oleracea	1993–2012	Turkey
AP017862	Brassica oleracea	1993–2012	Turkey
AP017861	Brassica oleracea	1993–2012	Turkey
AP017860	Brassica oleracea	1993–2012	Turkey
AP017859	Brassica oleracea	1993-2012	Turkey

AP017858	Brassica oleracea	1993-2012	Turkey
AP017857	Raphanus sativus	1993-2012	Iran
AP017856	Raphanus sativus	1993-2012	Iran
AP017855	Raphanus sativus	1993-2012	Iran
AP017854	Raphanus sativus	1993-2012	Iran
AP017853	Raphanus sativus	1993-2012	Iran
AP017852	Raphanus sativus	1993-2012	Iran
AP017851	Raphanus sativus	1993-2012	Iran
AP017850	Raphanus sativus	1993-2012	Iran
AP017849	Raphanus sativus	1993-2012	Iran
AP017848	Raphanus sativus	1993-2012	Iran
AP017847	Raphanus sativus	1993-2012	Iran
AP017846	Raphanus sativus	1993-2012	Iran
AP017845	Raphanus sativus	1993-2012	Iran
AP017844	Raphanus sativus	1993-2012	Iran
AP017843	Raphanus sativus	1993-2012	Iran
AP017842	Raphanus sativus	1993-2012	Iran
AP017841	Raphanus sativus	1993-2012	Iran
AP017840	Brassica oleracea var. capitata	1993-2012	Greece
AP017839	Brassica oleracea var. capitata	1993-2012	Greece
AP017838	Brassica oleracea var. botrytis	1993-2012	Greece
AP017837	Brassica oleracea var. capitata	1993-2012	Greece
AP017836	Brassica oleracea var. capitata	1993-2012	Greece
AP017835	Brassica oleracea var. botrytis	1993-2012	Greece
AP017834	Brassica oleracea var. botrytis	1993–2012	Greece
AP017833	Brassica oleracea var. capitata	1993-2012	Greece

AP017832	Brassica oleracea var. capitata	1993-2012	Greece
AP017831	Brassica oleracea var. capitata	1993–2012	Greece
AP017830	Brassica oleracea var. botrytis	1993–2012	Greece
AP017829	Brassica oleracea var. botrytis	1993–2012	Greece
AP017828	Brassica oleracea var. botrytis	1993–2012	Greece
AP017827	Brassica oleracea var. capitata	1993–2012	Greece
AP017826	Brassica oleracea var. botrytis	1993–2012	Greece
AP017825	Brassica oleracea var. botrytis	1993–2012	Greece
AP017824	Brassica oleracea var. botrytis	1993–2012	Greece
AP017823	Brassica oleracea var. botrytis	1993-2012	Greece
AP017822	Brassica oleracea var. botrytis	1993-2012	Greece
AP017821	Brassica oleracea var. botrytis	1993-2012	Greece
AP017820	Brassica oleracea var. botrytis	1993-2012	Greece
AP017819	Brassica oleracea var. capitata	1993-2012	Greece
AP017818	Brassica oleracea var. capitata	1993-2012	Greece
AP017817	Brassica oleracea var. capitata	1993-2012	Greece
AP017816	Brassica oleracea var. capitata	1993-2012	Greece
AP017815	Eruca sativa	1993–2012	Turkey
AP017814	Raphanus raphanistrum	1993–2012	Turkey
AP017813	Zinia elagans	1993-2012	Iran
AP017812	Raphanus sativus	1993-2012	Iran
AP017811	Raphanus sativus	1993-2012	Iran
AP017810	Raphanus sativus	1993–2012	Iran
AP017809	Raphanus sativus	1993-2012	Iran
AP017808	Brassica rapa	1993–2012	Iran
AP017807	Brassica rapa	1993-2012	Iran

AP017806	Raphanus sativus	1993-2012	Iran
AP017805	Brassica rapa	1993-2012	Iran
AP017804	Brassica rapa	1993-2012	Iran
AP017803	Rapistrum rugosum	1993-2012	Iran
AP017802	Sisymbrium loeselii	1993-2012	Iran
AP017801	Sisymbrium loeselii	1993-2012	Iran
AP017800	Sisymbrium loeselii	1993-2012	Iran
AP017799	Sisymbrium loeselii	1993-2012	Iran
AP017798	Mattiola sp.	1993-2012	Iran
AP017797	Sisymbrium irio	1993-2012	Iran
AP017796	Impatiens balsamina	1993-2012	Iran
AP017795	Hirschfeldia incana	1993–2012	Iran
AP017794	Sisymbrium loeselii	1993–2012	Iran
AP017793	Raphanus sativus	1993–2012	Iran
AP017792	Brassica oleracea	1993–2012	Iran
AP017791	Sisymbrium loeselii	1993-2012	Iran
AP017790	Sisymbrium loeselii	1993-2012	Iran
AP017789	Sisymbrium loeselii	1993-2012	Iran
AP017788	Sisymbrium loeselii	1993-2012	Iran
AP017787	Raphanus sativus	1993-2012	Iran
AP017786	Raphanus sativus	1993-2012	Iran
AP017785	Rapistrum rugosum	1993-2012	Iran
AP017784	Rapistrum rugosum	1993-2012	Iran
AP017783	Rapistrum rugosum	1993-2012	Iran
AP017782	Raphanus sativus	1993–2012	Iran
AP017781	Raphanus sativus	1993-2012	Iran

AP017780	Rapistrum rugosum	1993-2012	Iran
AP017779	Cheiranthus cheiri	1993–2012	Iran
AP017778	Brassica rapa	1993-2012	Iran
AP017777	Mattiola sp.	1993-2012	Iran
AP017776	Eruca sativa	1993-2012	Iran
AP017775	Eruca sativa	1993–2012	Iran
AP017774	Eruca sativa	1993-2012	Iran
AP017773	Brassica rapa	1993-2012	Iran
AP017772	Brassica rapa	1993-2012	Iran
AP017771	Brassica rapa	1993–2012	Iran
AP017770	Brassica oleracea var. botrytis	1993–2012	Iran
AP017769	Sisymbrium loeselii	1993–2012	Iran
AP017768	Sisymbrium loeselii	1993–2012	Iran
AP017767	Raphanus sativus	1993–2012	Iran
AP017766	Raphanus sativus	1993–2012	Iran
AP017765	Mattiola sp.	1993–2012	Iran
AP017764	Brassica oleracea var. italica	1993–2012	Iran
AP017763	Chrysanthemum sp.	1993–2012	Iran
AP017762	Cheiranthus cheiri	1993–2012	Iran
AP017761	Brassica napus	1993–2012	Iran
AP017760	Mattiola sp.	1993–2012	Iran
AP017759	Raphanus sativus	1993–2012	Iran
AP017758	Raphanus sativus	1993–2012	Iran
AP017757	Raphanus sativus	1993–2012	Iran
AP017756	Wild Allium sp.	1993–2012	Greece
AP017755	Cheiranthus sp.	1993-2012	Greece

AP017754	Mattiola sp.	1993-2012	Iran
AP017753	Mattiola sp.	1993-2012	Iran
AP017752	Mattiola sp.	1993-2012	Iran
AP017751	Spinacia oleracea	1993-2012	Turkey
AP017750	Raphanus raphanistrum	1993-2012	Turkey
AP017749	Eruca sativa	1993-2012	Turkey
AP017748	Raphanus raphanistrum	1993-2012	Turkey
AP017747	Raphanus raphanistrum	1993–2012	Turkey
AP017746	Raphanus raphanistrum	1993–2012	Turkey
AP017745	Raphanus raphanistrum	1993–2012	Turkey
AP017744	Raphanus raphanistrum	1993–2012	Turkey
AP017743	Raphanus raphanistrum	1993–2012	Turkey
AP017742	Brassica rapa	1993–2012	Turkey
AP017741	Eruca sativa	1993–2012	Turkey
AP017740	Eruca sativa	1993–2012	Turkey
AP017739	Brassica rapa	1993–2012	Turkey
AP017738	Eruca sativa	1993–2012	Turkey
AP017737	Brassica rapa	1993–2012	Turkey
AP017736	Raphanus raphanistrum	1993–2012	Turkey
AP017735	Brassica rapa	1993–2012	Turkey
AP017734	Raphanus raphanistrum	1993–2012	Turkey
AP017733	Brassica rapa	1993-2012	Turkey
AP017732	Brassica rapa	1993–2012	Turkey
AP017731	Raphanus raphanistrum	1993–2012	Turkey
AP017730	Eruca sativa	1993–2012	Turkey
AP017729	Raphanus raphanistrum	1993-2012	Turkey

AP017728	Raphanus raphanistrum 1993–2012		Turkey
AP017727	Sinapis arvensis	Sinapis arvensis 1993–2012	
AP017726	Sinapis arvensis	1993-2012	Greece
AP017725	Sinapis arvensis	1993-2012	Greece
AP017724	Eruca sativa	1993-2012	Greece
AP017723	Eruca sativa	1993-2012	Greece
AP017722	Eruca sativa	1993-2012	Greece
AP017721	Eruca sativa	1993-2012	Greece
AP017720	Sinapis arvensis	1993-2012	Greece
AP017719	Eruca sativa	1993-2012	Greece
AP017718	Eruca sativa	1993-2012	Greece
AP017717	Allium neapolitanum	1993-2012	Greece
AP017716	Allium hirsutum	1993-2012	Greece
AP017715	Brassica rapa	1993-2012	Greece
AP017714	Eruca sativa	1993-2012	Greece
AP017713	Eruca sativa	1993-2012	Greece
AP017712	Sinapis arvensis	1993-2012	Greece
KU140422	Nicotiana benthamiana	2014	South Korea
KU140421	Nicotiana benthamiana	2014	South Korea
KU140420	Raphanus sativus	2014	South Korea
KR153040	Raphanus sativus	2014	China
KR153039	Raphanus sativus	2014	China
KR153038	Raphanus sativus	2014	China
KU053508	Arthropodium cirratum	2014	New Zealand
AB989659	Lepidium oleraceum	1994–2011	New Zealand
AB989658	Lepidium oleraceum	1994–2011	New Zealand

AB989657	Lepidium oleraceum	1994–2011	New Zealand
AB989656	Brassica rapa	1994–2011	New Zealand
AB989655	Brassica rapa	1994–2011	New Zealand
AB989654	Brassica rapa	1994–2011	New Zealand
AB989653	Brassica spp.	1994–2011	New Zealand
AB989652	Brassica rapa cv. Marco	1994–2011	New Zealand
AB989651	Brassica rapa cv. Marco	1994–2011	New Zealand
AB989650	Brassica rapa cv. Marco	1994–2011	New Zealand
AB989649	Pachycladon fastigiatum	1994–2011	New Zealand
AB989648	Pachycladon fastigiatum	1994–2011	New Zealand
AB989647	Lepidium oleraceum	1994–2011	New Zealand
AB989646	Brassica napus cv. York Globe	1994–2011	New Zealand
AB989645	Nasturtium officinale	1994–2011	New Zealand
AB989644	Crocus sativus	1994–2011	New Zealand
AB989643	Rapistrum rugosum	1994–2011	Australia
AB989642	Hirschfeldia incana	1994–2011	Australia
AB989641	Brassica rapa	1994–2011	Australia
AB989640	Brassica rapa	1994–2011	Australia
AB989639	Rapistrum raphanistrum	1994–2011	Australia
AB989638	Rapistrum rugosum	1994–2011	Australia
AB989637	Hirschfeldia incana	1994–2011	Australia
AB989636	Rapistrum raphanistrum	1994–2011	Australia
AB989635	Brassica pekinensis	1994–2011	Australia
AB989634	Brassica pekinensis	1994–2011	Australia
AB989633	Rapistrum rugosum	1994–2011	Australia
AB989632	Brassica juncea	1994-2011	Australia

AB989631	Brassica juncea	1994–2011	Australia
AB989630	Rapistrum rugosum	1994–2011	Australia
Unknow	Unknow	Unknow	Unknow
Unknow	Unknow	Unknow	Unknow
AB989629	Cicer arietinum	1994–2011	Australia
AB989628	Hirschfeldia incana	1999	Australia
AB701742	wild B. oleracea	2002	Staithes, Yorkshire, UK
AB701741	Raphanus sativus	2002	USA
AB701740	Raphanus sativus	1993	USA
AB701739	Brassica pekinensis	1986	USA
AB701738	Tulipa gesnerana	1960	USA
AB701737	Sesynibium sp.	1997	USA
AB701736	Utricularia sp.	1983	Germany
AB701735	Tigridia sp.	1983	Germany
AB701734	Tigridia sp.	1934	Germany
AB701733	Brassica sp.	1993	UK
AB701732	Brassica napus oleifera	1993	Poland
AB701731	Papaver somniferum	Not known	Poland
AB701730	Brassica oleracea	1993/1994	Germany
AB701729	Brassica oleracea acephala	1993	Portugal
AB701728	Brassica napus oleifera	1995	Poland
AB701727	Brassica oleracea	1995	Netherlands
AB701726	Cucurbita pepo	1993	Italy
AB701725	Abutilon sp.	1992	Italy
AB701724	Matthiola incana	1990	Italy
AB701723	Brassica ruvo	1990	Italy

AB701722	Brassica rapa	1992	Italy
AB701721	Cheiranthus cheiri	1990	Italy
AB701720	Brassica ruvo	1996	Italy
AB701719	Alliaria petiolata	2002	Hungary
AB701718	Brassica oleracea	2002	UK
AB701717	Brassica oleracea	2000	UK
AB701716	Brassica oleracea	1999	UK
AB701715	Brassica oleracea	1999	UK
AB701714	Brassica oleracea	1999	UK
AB701713	Brassica oleracea	1999	UK
AB701712	Brassica oleracea	1999	UK
AB701711	Brassica oleracea	1999	UK
AB701710	Lunaria annua	1994	UK
AB701709	Rheum rhabarbarum	1993	UK
AB701708	Brassica napus	1994	France
AB701707	Sisymbrium orientale	2001	Spain
AB701706	Eruca sativa	2001	Spain
AB701705	Eruca sativa	1991	Italy
AB701704	Brassica rapa	1986	Denmark
AB701703	Brassica rapa	1978	Denmark
AB701702	Lactuca sativa	1991	Germany
AB701701	Lactuca sativa	1986	Germany
AB701700	Raphanus sativus	1993	Germany
AB701699	Unknown	1976	Germany
AB701698	Rorippa nasturtium-aquaticum	1986	Belgium
AB701697	Allium sp.	1995	Germany

AB701696	Matthiola incana	1989	Greece
AB701695	Lactuca sativa	1994	Germany
AB701694	Alliaria officinalis	1991	Denmark
AB701693	Orchis simia	1981	Germany
AB701692	Orchis morio	1983	Germany
AB701691	Orchis militaris	1981	Germany
AB701690	Orchis militaris	1981	Germany
KC119189	Cabbage	2009	China
KC119188	Cabbage	2010	China
KC119187	Cabbage	2010	China
KC119186	Cabbage	2010	China
KC119185	Cabbage	2010	China
KC119184	Radish	2010	China
HQ446217	Cruciferous plants	1985-1987	China
HQ446216	Brassica	1986-1990	China
HQ637383	Armoracia rusticana	2004	Poland
EF374098	Unknow	Unknow	Poland
AF394602	Unknow	Unknow	Unknow
AF394601	Unknow	Unknow	Unknow
LC314399	Narcissus tazzeta var. chinensis	2009-2015	Japan
LC314398	Narcissus tazzeta var. chinensis	2009-2015	Japan
LC314397	Narcissus tazzeta var. chinensis	2009-2015	Japan
LC314396	Narcissus tazzeta var. chinensis	2009-2015	Japan
LC314395	Narcissus tazzeta var. chinensis	2009-2015	Japan
LC314394	Narcissus tazzeta var. chinensis	2009–2015	China
LC314393	Narcissus tazzeta var. chinensis	2009–2015	Japan

	LC314392	Narcissus tazzeta var. chinensis	2009-2015	Japan
	LC314391	Narcissus tazzeta var. chinensis	2012	Japan
	NC_030391	Wild onion	2012	Turkey
	LC159495	Wild onion	2012	Turkey
	LC159494	Wild onion	2016	Turkey
Sugarcane mosaic virus				
(SCMV)	EU091075	Zea mays	2010	Mexico and Cameroon
	KU886553	Zea mays	2015	China
	JX047431	Zea mays	2011	China
	JX047430	Zea mays	2011	China
	JX047429	Zea mays	2011	China
	JX047428	Zea mays	2011	China
	JX047427	Zea mays	2011	China
	JX047426	Zea mays	2011	China
	JX047425	Zea mays	2011	China
	JX047424	Zea mays	2011	China
	JX047423	Zea mays	2011	China
	JX047422	Zea mays	2011	China
	JX047421	Zea mays	2011	China
	JX047420	Zea mays	2011	China
	JX047419	Zea mays	2011	China
	JX047418	Zea mays	2011	China
	JX047417	Zea mays	2011	China
	JX047416	Zea mays	2011	China
	JX047415	Zea mays	2011	China
	JX047414	Zea mays	2011	China
	JX047413	Zea mays	2010	China

JX047412	Zea mays	2010	China
JX047411	Zea mays	2010	China
JX047410	Zea mays	2010	China
JX047409	Zea mays	2010	China
JX047408	Zea mays	2010	China
JX047407	Zea mays	2010	China
JX047406	Zea mays	2010	China
JX047405	Zea mays	2010	China
JX047404	Zea mays	2010	China
JX047403	Zea mays	2010	China
JX047402	Zea mays	2010	China
JX047401	Zea mays	2010	China
JX047400	Zea mays	2010	China
JX047399	Zea mays	2010	China
JX047398	Zea mays	2010	China
JX047397	Zea mays	2010	China
JX047396	Zea mays	2010	China
JX047395	Zea mays	2010	China
JX047394	Zea mays	2010	China
JX047393	Zea mays	2010	China
JX047392	Zea mays	2010	China
JX047391	Zea mays	2010	China
JX047390	Zea mays	2010	China
JX047389	Zea mays	2010	China
JX047388	Zea mays	2010	China
JX047387	Zea mays	2010	China

JX047386	Zea mays	2010	China
JX047385	Zea mays	2010	China
JX047384	Zea mays	2010	China
JX047383	Zea mays	2010	China
JX047382	Zea mays	2010	China
JX047381	Zea mays	2010	China
KR611114	Zea mays	2013	China
KR611113	Zea mays	2013	China
KR611112	Zea mays	2013	China
KR611111	Zea mays	2012	China
KR611110	Zea mays	2012	China
KR611109	Zea mays	2013	China
KR611108	Zea mays	2012	China
KR611107	Zea mays	2012	China
KR611106	Zea mays	2013	China
KR611105	Zea mays	2012	China
KT895081	Zea mays	2013	Iran
KT895080	Saccharum officinarum	2013	Iran
JX188385	Zea mays	1965	EUA
JX185303	Unknow	Unknow	Germany
JN021933	Zea mays	2010	China
AY569692	Zea mays	2003	China
GU474635	Zea mays	2009	Mexico
AF494510	Unknow	Unknow	China
AM110759	Zea mays	1992 e 1998	Spain
MG932080	Zea mays and Sorghum bicolor	2012-2014	Kenya

MG932079	Sorghum bicolor 2014		Kenya
MG932078	Zea mays	Zea mays 2012	
MG932077	Zea mays	2012	Kenya
MG932076	Zea mays	2012	Kenya
KY548507	Saccharum officinarum	2016	China
KY548506	Saccharum officinarum	2012	China
KY006657	Zea mays	2012	Ecuador
KR108213	Saccharum officinarum	2014	China
KR108212	Saccharum officinarum	2014	China
KP772216	Zea mays	2014	Ethiopia
KP860936	Zea mays	2014	Ethiopia
KP860935	Zea mays	2014	Ethiopia
NC_003398	Zea mays	2000	China
KF744392	Zea mays	2013	Rwanda
KF744391	Zea mays	2013	Rwanda
KF744390	Zea mays	2013	Rwanda
JX237863	Saccharum officinarum	2007	Argentina
JX237862	Saccharum officinarum	2010	Argentina
HM133588	Zea mays	2008	EUA
AY149118	Zea mays	2002	China
AY042184	Zea mays	Unknow	China
AJ278405	Unknow	Unknow	Australia
AJ310105	Zea mays	2000	China
AJ310104	Saccharum officinarum	2000	China
AJ310103	Saccharum officinarum	2000	China
AJ310102	Saccharum officinarum	2000	China

	AJ297628	Zea mays	2000	China
	AM110758	Zea mays	Unknow	Spain
Soybean mosaic virus (SMV)	S42280	Glycine max	1992	EUA
	NC_002634	Glycine max	1988	EUA
	GU015011	Glycine max	2002	EUA
	FJ640982	Glycine max	2003	South Korea
	FJ640981	Glycine max	2003	South Korea
	FJ640980	Glycine max	2003	South Korea
	FJ640979	Glycine max	2003	South Korea
	FJ640978	Glycine max	2003	South Korea
	FJ640977	Glycine max	2003	South Korea
	FJ640976	Glycine max	2006	South Korea
	FJ640975	Glycine max	2006	South Korea
	FJ640974	Glycine max	2006	South Korea
	FJ640973	Glycine max	2006	South Korea
	FJ640972	Glycine max	2006	South Korea
	FJ640971	Glycine max	2006	South Korea
	FJ640970	Glycine max	2006	South Korea
	FJ640969	Glycine max	2006	South Korea
	FJ640968	Glycine max	2006	South Korea
	FJ640967	Glycine max	2006	South Korea
	FJ640966	Glycine max	2006	South Korea
	FJ640965	Glycine max	2006	South Korea
	FJ640964	Glycine max	2006	South Korea
	FJ640963	Glycine max	2006	South Korea
	FJ640962	Glycine max	2006	South Korea

FJ640961	Glycine max	2006	South Korea
FJ640960	Glycine max	2006	South Korea
FJ640959	Glycine max	2006	South Korea
FJ640958	Glycine max	2006	South Korea
FJ640957	Glycine max	2006	South Korea
FJ640956	Glycine max	2006	South Korea
FJ640955	Glycine max	2006	South Korea
FJ640954	Glycine max	2006	South Korea
FJ548849	Glycine max	2006	South Korea
AY216987	Glycine max	2003	EUA
AY216010	Glycine max	2003	EUA
D00507	Glycine max	1988	EUA
KY986929	Vigna angularis	2016	South Korea
KY249378	Passiflora edulis f. edulis	2015	Colombia
KT285170	Glycine max	2004	China
KR065437	Glycine max	2012	China
KP710878	Glycine max	2013	China
KP710877	Glycine max	2013	China
KP710876	Glycine max	2013	China
KP710875	Glycine max	2013	China
KP710874	Glycine max	2013	China
KP710873	Glycine max	2013	China
KP710872	Glycine max	2013	China
KP710871	Glycine max	2013	China
KP710870	Glycine max	2013	China
KP710869	Glycine max	2013	China

KP710868	Glycine max	2010	China
KP710867	Glycine max	2010	China
KP710866	<i>Glvcine max</i>	2013	China
KP710865	<i>Glycine max</i>	2013	China
KP710864	<i>Glycine max</i>	2013	China
KP710863	Glycine max	2013	China
KP710862	Glycine max	2013	China
KP710861	Glycine max	2013	China
KR024718	Glycine max	2014	China
FJ376388	<i>Glycine max</i>	2005	South Korea
KF982784	Pinellia pedatisecta	2011	China
KF297335	Glycine max	2011	Iran
KF135491	Glycine max	2011	Iran
KF135490	Glycine max	2011	Iran
KF135489	Glycine max	2011	Iran
KF135488	Glycine max	2011	Iran
KC845322	Atractylodes macrocephala	2012	China
KC845321	Glycine max	2012	China
HQ845736	Glycine max	2010	EUA
HQ845735	Glycine max	2010	EUA
HQ166266	Glycine max	2009	Canada
HQ166265	Glycine max	2009	Canada
JN416770	Glycine max	2010	Canada
JF833015	Glycine max	2005	China
JF833014	Glycine max	2005	China
JF833013	Glycine max	2005	China

HM590055	Glycine max	2004	China
HM590054	Glycine max	2004	China
AF241739	Glycine max	1999	EUA
AY294045	Glycine max	2002	South Korea
AY294044	Glycine max	2002	South Korea
FJ807701	Glycine max	2005	South Korea
FJ807700	Glycine max	2005	South Korea
EU871725	Glycine max	2005	Canada
EU871724	Glycine max	2005	Canadá
AJ312439	Glycine max	2000	China
AJ310200	Glycine max	2000	China
AJ507388	Pinellia ternata	2002	China
AJ619757	Glycine max	2003	South Korea
AB100443	Unknow	Unknow	Unknow
AB100442	Unknow	Unknow	Unknow
KX096578	Glycine max	2015	China
KX834325	Glycine max	2016	China
KX834324	Glycine max	2016	China
KX834323	Glycine max	2016	China
KX834322	Glycine max	2016	China
KX834321	Glycine max	2016	China
KX834320	Glycine max	2016	China
KX834319	Glycine max	2016	China
LC037232	Uraria crinita	2014	Taiwan
KM979229	Glycine max	2014	india
HQ396725	Glycine max	2004	China

	AJ628750	Pinellia ternata	2003	China
	MF981910	Glycine max	2017	China
Bean yellow mosaic virus				
(BYMV)	JN692500	Vicia faba	2011	India
	KT934334	Sunflower	2012	Iran
	FJ492961	Freesia sp.	Unknow	South Korea
	JX173278	Diuris magnifica	2011	Australia
	AB373203	Unknow	Unknow	Unknow
	KF632713	Diuris sp.	2012	Australia
	KM114059	Gladiolus dalenii cv. Sylvia	2012	India
	KF155409	Gladiolus cv. Aldebaran	2012	India
	KF155414	Gladiolus cv. Tiger flame	2013	India
	KF155419	Gladiolus cv. Snow princess	2013	India
	KF155420	Gladiolus cv. Regency	2013	India
	AB079888	Unknow	Unknow	Unknow
	JX156423	Diuris sp.	2011	Australia
	HG970850	Lupinus cosentinii	2011	Australia
	HG970852	Lupinus angustifolius	2011	Australia
	HG970854	Lupinus angustifolius	2011	Australia
	HG970855	Lupinus angustifolius	1998	Australia
	HG970859	Lupinus angustifolius	2011	Australia
	HG970860	Lupinus angustifolius	2011	Australia
	HG970861	Lupinus angustifolius	2011	Australia
	HG970863	Lupinus angustifolius	2011	Australia
	HG970865	Lupinus angustifolius	2011	Australia
	HG970869	Lupinus angustifolius	2012	Australia
	HG970851	Lupinus angustifolius	2011	Australia

	AM884180	Eustoma russellianum	Unknow	Taiwan
	HG970866	Lupinus pilosus	1998	Australia
	HG970858	Lupinus angustifolius	2011	Australia
	DQ641248	Pisum sativum L	Unknow	USA
	AB439732	Trifolium pratense	1991	Japan
	AB439731	Vicia faba	1990	Japan
	HG970857	Lupinus angustifolius	2011	Australia
	AB439729	Gladiolus hybrid cultivar	Unknow	Japan
	AB439730	Gladiolus hybrid cultivar	Unknow	Japan
	HG970864	Lupinus angustifolius	2011	Australia
	HG970867	Vicia faba	1998	Australia
	HG970868	Vicia faba	1998	Australia
	HG970853	Lupinus angustifolius	2011	Australia
	HG970856	Lupinus angustifolius	2011	Australia
	HG970848	Lupinus angustifolius	2011	Australia
	HG970847	Lupinus cosentinii	2011	Australia
	HG970849	Lupinus angustifolius	2011	Australia
	HG970862	Lupinus angustifolius	2011	Australia
	MG600297	Trifolium pratense L.	2011	Czech Republic
Papaya ringspot virus				
(PRSV)	KP462721	Fevillea cordifolia	2012	Brazil:
	KC345607	Momordica charantia	2001	France
	NC_023175	Momordica charantia	2001	France
	KC345609	Cucurbita pepo	1979	France
	MF356497	Carica papaya (papaya)	2015	India
	KP743981	Carica papaya	Unknow	India
	KC345605	Cucurbita pepo	1981	Guadeloupe

KC345608	Cucurbita pepo	2011	France
MF085000	Cucurbita pepo L.	2016	China
EU475877	Unknow	Unknow	India
DQ374153	Cucurbita pepo	Unknow	Brazil
HQ424465	Carica papaya L.	Jul-1905	China
KF734962	Carica papaya L	2011	China
KT895257	Carica papaya	4-Jul-1905	China
KX998707	Cucurbit	2015	Australia
MF074214	Pumpkin	2016	China
NC_001785	Carica papaya	Unknow	Taiwan
DQ340769	Carica papaya	Unknow	Taiwan
DQ340770	Carica papaya	Unknow	Taiwan
DQ340771	Carica papaya	Unknow	Taiwan
DQ374152	Cucurbita pepo	Unknow	Brazil
JX448369	Carica papaya	Unknow	Taiwan
JX448370	Carica papaya	Unknow	Taiwan
JX448371	Carica papaya	Unknow	Taiwan
JX448372	Carica papaya	Unknow	Taiwan
JX448373	Carica papaya	Unknow	Taiwan
KJ755852	Carica papaya	2013	India
KT275937	Carica papaya Maradol	19-Mar-1905	Colombia
KT275938	Carica papaya Maradol	06-Jul-1905	Colombia
KX655866	Curcurbit	2015	Australia
KY933061	Carica papaya	08-Jul-1905	China
KT633943	Carica papaya	2015	China
KX655872	Curcurbit	2011	Australia

KY996464	Cucurbita pepo	2014	South Korea
AY010722	Unknow	Unknow	Thailand
AY162218	Unknow	Unknow	Thailand
EF183499	Carica papaya L	Unknow	China
AY231130	Carica papaya L.	Unknow	Mexico
EU126128	Unknow	Unknow	USA
EU882728	Carica papaya	2007	Taiwan
KY271954	Carica papaya L. (papaya)	2014	USA
KC345606	Cucurbita moschata	2010	Venezuela
KX655867	Curcurbit	2004	Australia
EF017707	Carica papaya L	Unknow	India
KX655873	Curcurbit	2011	Australia
KX655874	Curcurbit	2011	East Timor
KU355553	Cucurbita pepo	2013	South Africa
KX655860	Curcurbit	2014	Australia
KY039583	Gourd	19-Mar-1905	USA
KX998708	Cucurbit	2015	Australia
KX655870	Curcurbit	2015	Australia
KX655868	Curcurbit	2015	Australia
KX655865	Curcurbit	2015	East Timor
KX655862	Curcurbit	2015	Australia
KX655861	Curcurbit	2015	Australia
KX655864	Curcurbit	2015	East Timor
KX655863	Curcurbit	2015	East Timor
KY623505	Cucumis melo var. flexuosus	1994	Sudan
NC_035459	Cucumis melo var. flexuosus	1994	Sudan

	KX655869	Curcurbit	2015	Australia
	KT633944	Carica papaya	2015	China
	KY623506	Cucumis melo var. agrestis	2003	Sudan
	NC_035458	Cucumis melo var. agrestis	2003	Sudan
	KX655871	Curcurbit	2014	Australia
	KX235326	Junagadh	19-Mar-1905	India
	KY271955	Carica papaya L. (papaya)	2014	USA
	KY271956	Carica papaya L. (papaya)	2014	USA
Lettuce mosaic virus (LMV)	AJ278854	Lettuce	Unknow	Brazil
	AJ306288	Lettuce	Unknow	China
	KF955619	Lettuce	2011	South Korea
	KF268954	Lettuce	2013	Brazil
	KF268955	Lettuce	2013	Brazil
	KJ161173	Lactuca sativa	Unknow	France
	KJ161174	Lactuca sativa	Unknow	Brazil
	KJ161175	Dimorphotheca sp.	2003	Chile
	KJ161176	Lactuca sativa	2005	Chile
	KJ161177	Lactuca virosa	2005	Chile
	KJ161178	Lactuca sativa	2005	Chile
	KJ161179	Lactuca sativa	2006	Chile
	KJ161180	Lactuca virosa	2007	Chile
	KJ161181	Lactuca virosa	2007	Chile
	KJ161182	Lactuca sativa	2007	Chile
	KJ161183	Lactuca virosa	2007	Chile
	KJ161184	Lactuca virosa	2007	Chile
	KJ161186	Dimorphotheca sp.	2008	France

	KJ161187	Dimorphotheca sp.	2002	Tunisia	
	KJ161188	Dimorphotheca sp.	2005	Tunisia	
	KJ161189	Dimorphotheca sp.	2005	Tunisia	
	KJ161190	Dimorphotheca sp.	2005	Tunisia	
	KJ161191	Dimorphotheca sp.	2005	Tunisia	
	KJ161192	Dimorphotheca sp.	2005	Tunisia	
	KJ161193	Dimorphotheca sp.	2005	Tunisia	
	KF268956	Catharanthus roseus	2013	France	
	KJ161172	Lactuca sativa	1984	France	
	KJ161185	Dimorphotheca sp.	2008	Spain	
	KJ161194	Lactuca sativa	Unknow	Yemen	
Zucchini yellow mosaic virus					
(ZYMV)	L31350	Cucúrbita moschata	Unknow	USA	
	DD056806	Cucúrbita moschata	1988-1989	USA	
	KY225556	Cucumis sativus	2015	East Timor	
	KY225555	Cucurbita pepo	2015	Australia	
	KY225554	Cucurbita pepo	2015	Australia	
	KY225553	Cucurbita pepo	2015	Australia	
	KY225552	Cucurbita pepo	2015	Australia	
	KY225551	Cucurbita pepo	2015	Australia	
	KY225550	Cucurbita pepo	2015	Australia	
	KY225549	Cucúrbita moschata	2015	Australia	
	KY225548	Cucumis melo	2011	Australia	
	KY225547	Cucumis melo	2011	Australia	
	KY225546	Cucumis melo	1996	Australia	
	KY225545	Cucúrbita moschata	2015	EastTimor	
	KY225544	Cucúrbita moschata	2015	EastTimor	

KY225543	Cucúrbita moschata	1996	Australia
KY225542	Cucúrbita moschata	1996	Australia
KX664482	Cucumis sativus	2016	China
KX421104	Sesamum indicum	2016	China
KX249747	Luffa aegyptiaca	2014	China
KX499498	Cucumis sativus	2016	Spain
KU528623	Unknow	2013	Iran
KU198853	Cucumis sativus	2013	Iran
KT598222	Cucúrbita moschata	2011	Argentina
KT778297	Cucumis anguria	2012	India
KJ875865	Cucúrbita moschata	2011	EUA
KJ875864	Cucúrbita moschata	2011	EUA
KF976713	Cucúrbita moschata	2014	Slovakia
KF976712	Cucúrbita moschata	2014	Kazakhstan
KJ923769	Cucúrbita moschata	2011	EUA
KJ923768	Cucúrbita moschata	2011	EUA
KJ923767	Cucúrbita moschata	2011	EUA
KC665635	Cucúrbita moschata	2011	EUA
KC665634	Cucúrbita moschata	2011	EUA
KC665633	Cucúrbita moschata	2011	EUA
KC665632	Cucúrbita moschata	2011	EUA
KC665632 KC665631	Cucúrbita moschata Cucúrbita moschata	2011 2011	EUA EUA
KC665632 KC665631 KC665630	Cucúrbita moschata Cucúrbita moschata Cucúrbita moschata	2011 2011 2009	EUA EUA EUA
KC665632 KC665631 KC665630 KC665629	Cucúrbita moschata Cucúrbita moschata Cucúrbita moschata Cucúrbita moschata	2011 2011 2009 2009	EUA EUA EUA EUA
KC665632 KC665631 KC665630 KC665629 KC665628	Cucúrbita moschata Cucúrbita moschata Cucúrbita moschata Cucúrbita moschata Cucúrbita moschata	2011 2011 2009 2009 2010	EUA EUA EUA EUA
JN183062	Cucúrbita moschata	2010	Iran
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JQ716413	Cucúrbita moschata	2006	EUA
JN192428	Cucúrbita moschata	2011	EUA
JN192427	Cucúrbita moschata	2011	EUA
JN192426	Cucúrbita moschata	2011	EUA
JN192425	Cucúrbita moschata	2011	EUA
JN192424	Cucúrbita moschata	2011	EUA
JN192423	Cucúrbita moschata	2011	EUA
JN192422	Cucúrbita moschata	2011	EUA
JN192421	Cucúrbita moschata	2011	EUA
JN192420	Cucúrbita moschata	2007	EUA
JN192419	Cucúrbita moschata	2007	EUA
JN192418	Cucúrbita moschata	2007	EUA
JN192417	Cucúrbita moschata	2007	EUA
JN192416	Cucúrbita moschata	2007	EUA
JN192415	Cucúrbita moschata	2007	EUA
JN192414	Cucúrbita moschata	2007	EUA
JN192413	Cucúrbita moschata	2007	EUA
JN192412	Cucúrbita moschata	2007	EUA
JN192411	Cucúrbita moschata	2007	EUA
JN192410	Cucúrbita moschata	2007	EUA
JN192409	Cucúrbita moschata	2007	EUA
JN192408	Cucúrbita moschata	2007	EUA
JN192407	Cucúrbita moschata	2007	EUA
JN192406	Cucúrbita moschata	2007	EUA
JN192405	Cucúrbita moschata	2007	EUA

	AM422386	Begonia elatior	2006	Taiwan
	AY188994	Cucurbita pepo	1992-2001	France, Italy, Israel
	AY279000	Cucurbita moschata	Unknow	Unknow
	AY278999	Cucurbita pepo	Unknow	Unknow
	AY278998	Cucurbita pepo	Unknow	Unknow
	EF062583	Cucurbita pepo	2000	Israel
	EF062582	Cucurbita pepo	2000	Israel
	DQ124239	Cucurbita pepo	2016	Slovakia
	AB188116	Cucumis sativus	1997	Japan
	AB188115	Cucumis sativus	1997	Japan
	AJ316229	Cucumis melo	2001	China
	AJ316228	Luffa cylindrica	2002	China
	AJ307036	Cucumis sativus	2001	China
	AJ515911	Citrullus lanatus	2002	China
	KX884570	Araneae	2013	China
	KX884565	Astacidea	2014	China
	NC003224	Luffa cylindrica	1999	Taiwan
	AF127929 e AF343979	Luffa cylindrica	2001	Taiwan
	AB369279	Cucurbita pepo	2007	South Korea
	DJ418432	Unknow	Unknow	Unknow
	DI159774	Unknow	Unknow	Unknow
	AF014811	Unknow	Unknow	Singapore
Plum pox virus (PPV)	AM157175	N. benthamiana	2005	Egypt
,	HG964686	Prunus sp.	2008	Canada
	HG964685	Prunus sp.	2008	Canada
	AY028309	Prunus armeniaca	2003	Slovakia

MF346290	Apricot	2016	Turkey
MF346289	Plum	2016	Turkey
MF346288	Apricot	2016	Turkey
MF346287	Apricot	2016	Turkey
MF346286	Apricot	2016	Turkey
MF346285	Apricot	2016	Turkey
MF346284	Apricot	2016	Turkey
MF346283	Apricot	2016	Turkey
MF346282	plum	2016	Turkey
MF346281	Apricot	2016	Turkey
MF346280	Apricot	2016	Turkey
MF346279	Plum	2016	Turkey
MF346278	Plum	2016	Turkey
MF346277	Apricot	2016	Turkey
MF346276	Plum	2016	Turkey
MF346275	Plum	2016	Turkey
MF346274	Apricot	2016	Turkey
MF346273	Plum	2016	Turkey
MF346272	Plum	2016	Turkey
MF346271	Plum	2016	Turkey
MF346270	Apricot	2016	Turkey
MF346269	Plum	2016	Turkey
MF346268	Plum	2016	Turkey
MF346267	Plum	2016	Turkey
MF346266	Apricot	2016	Turkey
MF346265	Apricot	2014	Turkey

ME246264	Ammiaat	2014	Tuelcon
MIF 340204	Apricot	2014	Turkey
MF346263	Apricot	2014	Turkey
MF346262	Apricot	2014	Turkey
MF346261	Apricot	2014	Turkey
MF346260	Apricot	2014	Turkey
MF346259	Apricot	2014	Turkey
MF346258	Apricot	2014	Turkey
MF346257	Apricot	2014	Turkey
MF346256	Apricot	2014	Turkey
MF346255	Apricot	2014	Turkey
MF346254	Apricot	2014	Turkey
MF346253	Apricot	2014	Turkey
MF346252	Apricot	2014	Turkey
MF346251	Apricot	2014	Turkey
MF346250	Apricot	2014	Turkey
MF346249	Apricot	2014	Turkey
MF346248	Apricot	2014	Turkey
MF346247	Apricot	2014	Turkey
MF346246	Apricot	2014	Turkey
MF346245	Plum	2015	Turkey
MF346244	Apricot	2015	Turkey
MF346243	Apricot	2015	Turkey
MF346242	Plum	2015	Turkey
MF346241	Apricot	2015	Turkey
MF346240	Apricot	2015	Turkey
MF346239	Apricot	2015	Turkey

MF346238	Peach	2015	Turkey
MF346237	Apricot	2015	Turkey
MF346236	Plum	2015	Turkey
MF346235	Apricot	2015	Turkey
LC228949	Prunus mume	2016	Japan
MF447180	Prunus cerasus	2015	Russia
MF447179	Prunus cerasus	2015	Russia
MF371004	Plum	2015	Turkey
MF371003	Peach	2015	Turkey
MF371002	Plum	2015	Turkey
MF371001	Plum	2015	Turkey
MF371000	Plum	2015	Turkey
MF370999	Plum	2015	Turkey
MF370998	Plum	2015	Turkey
MF370997	Plum	2015	Turkey
MF370996	Plum	2015	Turkey
MF370995	Peach	2015	Turkey
MF370994	Plum	2015	Turkey
MF370993	Plum	2015	Turkey
MF370992	Plum	2015	Turkey
MF370991	Plum	2015	Turkey
MF370990	Plum	2015	Turkey
MF370989	Plum	2015	Turkey
MF370988	Plum	2015	Turkey
MF370987	Plum	2015	Turkey
MF370986	Apricot	2015	Turkey

MF370985	Apricot	2015	Turkey
MF370984	Plum	2015	Turkey
LC333553	Prunus persica	2017	South Korea
LC333552	Prunus persica	2017	South Korea
LC331298	Prunus mume	2017	South Korea
LC333268	Prunus persica	2017	South Korea
KY221840	Prunus cerasus	2014	Germany
KU508427	Prunus domestica cv. jojo	2012	Germany
KP998124	Prunus persica	2013	Canada
LT158756	Prunus persica	2016	Slovakia
KR028387	Myrobalan Prunus cerasifera	2011	Ukraine
KR028386	Nectarine	2013	Ukraine
KR028385	Peach	2013	Ukraine
KR006730	Prunus domestica	2011	Ukraine
KR006729	Peach	2011	Ukraine
HG916858	Prunus domestica	2015	Russia
HG916857	prunus domestica	2015	Russia
HG916856	Prunus tomentosa	2015	Russia
HG916862	Prunus nigra	2015	Russia
HG916861	Prunus domestica	2015	Russia
HG916860	Prunus domestica	2015	Russia
HG916859	Prunus spinosa	2015	Russia
LN852400	Prunus domestica	2015	Russia
HG916863	Prunus nigra	2015	Russia
HF585103	Prunus persica	2013	Slovakia
HF585102	Prunus persica	2013	Slovakia

HF585101	Prunus persica	2013	Slovakia
HF585100	Prunus persica	2013	Slovakia
HF585099	Prunus domestica	2013	Slovakia
NC_001445	Unknow	Unknow	Unknow
KJ787006	Prunus cerasus	2014	Russia
LN614587	Prunus armeniaca	2014	Slovakia
KJ849228	Japanese plum	1995	Spain
HF674399	Prunus domestica	2010	Albania
KC347608	Prunus domestica	2011	Rússia
KC020126	Prunus cerasus	2010-2012	Rússia
KC020125	Prunus cerasus	2010-2012	Rússia
KC020124	Prunus cerasus	2010-2012	Rússia
HF585104	Prunus persica	2013	Slováquia
HF585098	Prunus persica	2013	Slováquia
JN596110	Prunus spinosa	2011	Ucrânia
HQ670748	Plum	2011	Latvia
HQ670746	Plum	2011	Latvia
HQ670745	Plum	2011	Latvia
JX013532	Prunus domestica	2007	Croatia
JQ794501	Prunus mume	2009	Slováquia
AB576080	Prunus mume cv. Koushuoujuku	2009	Japão
AB576079	Prunus mume cv. Mongakushi	2009	Japão
AB576078	Prunus mume cv. Shishigashira	2009	Japão
AB576077	Prunus mume cv. Kurenai	2009	Japão
	Prunus mume cv. Tenjinbai and cv.		
AB576076	Kusudama	2009	Japão
AB576075	Prunus mume cv. Hamachidori	2009	Japão

AB576074	Prunus mume cv. Hamachidori	2009	Japão
AB576073	Prunus mume cv. Hamachidori	2009	Japão
AB576072	Prunus mume	2009	Japão
AB576071	Prunus mume	2009	Japão
AB576070	Prunus mume	2009	Japão
AB576069	Prunus mume cv. Hamachidori	2009	Japão
AB576068	Prunus mume cv. kurenai	2009	Japão
AB576067	Prunus mume cv. Komukai	2009	Japão
AB576066	Prunus mume	2009	Japão
AB576065	Prunus mume cv. Shirokaga	2009	Japão
AB576064	Prunus persica	2009	Japão
AB576063	Prunus mume	2009	Japão
AB576062	Prunus mume	2009	Japão
AB576061	Prunus mume	2009	Japão
AB576060	Prunus mume cv. Nanko	2009	Japão
AB576059	Prunus mume	2009	Japão
AB576058	Prunus mume cv. Baigo	2008	Japão
AB576057	Prunus mume	2009	Japão
AB576056	Prunus mume	2009	Japão
AB576055	Prunus mume	2009	Japão
AB576054	Prunus mume cv. Nanko	2009	Japão
AB576053	Prunus mume cv. Shirokaga	2009	Japão
AB576052	Prunus domestica	2009	Japão
AB576051	Prunus mume	2009	Japão
AB576050	Prunus mume	2009	Japão
AB576049	Prunus mume	2009	Japão

AB576048	Prunus mume	2009	Japão
AB576047	Prunus mume	2009	Japão
AB576046	Prunus mume	2009	Japão
AB576045	Prunus mume cv. Nanko	2009	Japão
AY912055	Prunus domestica	2003	Canada
HQ840518	Prunus hybrid cultivar OWP-6	2010	Bielorússia
HQ840517	Prunus hybrid cultivar L2	2010	Bielorússia
AB545926	Prunus mume cv. Nanko	2008	Japão
EU734794	Prunus armeniaca	2004	Turquia
GU474956	Prunus persica (peach)	1990	Serbia
GU461890	Prunus domestica (plum)	1996	Slováquia
GU461889	Prunus domestica (plum)	2003	Bulgaria
M92280 X56759	Unknow	Unknow	Unknow
AY184478	Unknow	Unknow	Unknow
EF611248	Peach	2007	USA
EF611247	Peach	2007	USA
EF611246	Peach	2007	USA
EF611245	Peach	2007	USA
EF611244	Peach	2007	USA
EF611243	Peach	2007	USA
EF611242	Peach	2007	USA
EF611241	Peach	2007	USA
DQ465243	Peach	2007	USA
DQ465242	Peach	2007	USA
DQ431465	Prunus armeniaca	1980	Egito
AY953267	P. persica cv. Vinegold peach	Unknow	Canada

AY953266	P. domestica cv. Italian plum	Unknow	Canada
AY953265	P. glandulosacv	Unknow	Canada
AY953264	P. persica	Unknow	Canada
AY953263	P. persica cv. Blushing star peach	Unknow	Canada
AY953262	P. persica cv. Vinegold peach	Unknow	Canada
AY953261	P. domestica cv. Italian plum	Unknow	Canada
AY912058	Prunus persica	Unknow	Canada
AY912057	Prunus persica	Unknow	Canada
AY912056	Prunus persica var. nectarina	Unknow	Canada
X16415	Unknow	Unknow	Unknow
AJ243957	Prunus sp.	1999	Serbia and Montenegro
D13751 D00424	Unknow	Unknow	Unknow
LT600782	Unknow	Unknow	Unknow
LT600781	Unknow	Unknow	Unknow
LT600780	Unknow	Unknow	Unknow
LT600779	Unknow	Unknow	Unknow
KU948432	Plum	2013	Canada
KM273015	Prunus domestica	2013	Russia
KF472134	Prunus cerasifera	2011	Ukraine
AF401296	Prunus domestica	2001	USA
AF401295	Prunus persica	2001	USA
EU117116	Prunus domestica	2007	Poland
FM955843	Unknow	Unknow	Greece
EF640939	Peach	2007	USA
EF640938	Peach	2007	USA
EF640937	Peach	2007	USA

	EF640936	Peach	2007	USA	
	EF640935	Peach	2007	USA	
	EF640934	Peach	2007	USA	
	EF640933	Peach	2007	USA	
	X81083	Unknow	Unknow	Unknow	
	EF569215	Nicotiana clevelandii	Unknow	Unknow	
	EF569214	Prunus persica	Unknow	Unknow	
Potato virus Y	A08776	Unknow	Unknow	Unknow	
	AB185833	Solanum tuberosum	Unknow	Syria	
	AB270705	Solanum tuberosum	2002-07-15	Syria	
	AB461451	Solanum tuberosum	2006	Syria	
	AB461452	Solanum tuberosum	2004	Syria	
	AB461453	Solanum tuberosum	2007	Syria	
	AB702945	Solanum tuberosum	2007	Japan	
	AB711143	Solanum tuberosum	Unknow	Japan	
	AB711144	Solanum tuberosum	Unknow	Japan	
	AB711145	Solanum tuberosum	Unknow	Japan	
	AB711146	Solanum tuberosum	Unknow	Japan	
	AB711147	Solanum tuberosum	Unknow	Japan	
	AB711148	Solanum tuberosum	Unknow	Japan	
	AB711149	Solanum tuberosum	Unknow	Japan	
	AB711150	Solanum tuberosum	Unknow	Japan	
	AB711151	Solanum tuberosum	Unknow	Japan	
	AB711152	Solanum tuberosum	Unknow	Japan	
	AB711153	Solanum tuberosum	Unknow	Japan	
	AB711154	Solanum tuberosum	Unknow	Japan	

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AB711155	Solanum tuberosum	Unknow	Japan
AB714134	Solanum tuberosum	Unknow	Japan
AB714135	Solanum tuberosum	1989	Japan
AF237963	Capsicum spp.	Unknow	Unknow
AF463399	Unknow	Unknow	Unknow
AJ584851	Solanum tuberosum	Unknow	United Kingdom
AJ585195	Solanum tuberosum	Unknow	United Kingdom
AJ585196	Solanum tuberosum	Unknow	United Kingdom
AJ585197	Solanum tuberosum	Unknow	United Kingdom
AJ585198	Solanum tuberosum	Unknow	United Kingdom
AJ585342	Solanum tuberosum	Unknow	Slovenia
AJ889866	Solanum tuberosum	Unknow	Poland
AJ889867	Solanum tuberosum	Unknow	Germany
AJ889868	Solanum tuberosum	Unknow	Germany
AJ890342	Nicotiana tabacum	Unknow	Poland
AJ890343	Nicotiana tabacum	Unknow	Poland
AJ890344	Nicotiana tabacum	Unknow	Poland
AJ890345	Nicotiana tabacum	Unknow	Germany
AJ890346	Nicotiana tabacum	Unknow	Germany
AJ890347	Solanum tuberosum	Unknow	Germany
AJ890348	Solanum tuberosum	Unknow	France
AJ890349	Solanum tuberosum	Unknow	Poland
AJ890350	Solanum tuberosum	Unknow	Germany
AM113988	Solanum tuberosum	Unknow	Germany
AM268435	Solanum tuberosum	Unknow	New Zealand
D00441	Unknow	Unknow	Unknow

DQ309028	Nicotiana tobacum	Unknow	Unknow
EF026074	Solanum tuberosum	2002-06	USA
EF026075	Solanum tuberosum	2002-07-15	USA
EF026076	Solanum tuberosum	2002-07-15	USA
EU563512	Solanum tuberosum	1938	Netherlands
FJ204164	Solanum tuberosum	2007	USA
FJ204165	Solanum tuberosum	2007	USA
FJ204166	Solanum tuberosum	2007	USA
FJ643477	Solanum tuberosum	2004	USA
FJ643478	Solanum tuberosum	2004	USA
FJ643479	Solanum tuberosum	2004	USA
FJ666337	Solanum tuberosum	1970	Poland
GQ200836	Solanum tuberosum	2007-04-27	China
HE608963	Solanum tuberosum	2010-09	Germany
HE608964	Solanum tuberosum	2010-09	Germany
HG810949	Solanum tuberosum	2012	Vietnam
HG810950	Solanum tuberosum	2012	Vietnam
HG810951	Solanum tuberosum	2012	Vietnam
HG810952	Solanum tuberosum	2012	Vietnam
HM367075	Solanum tuberosum	2007	Canada
HM367076	Solanum tuberosum	2007	Canada
HM590405	Nicotiana tabacum	Unknow	Unknow
HM590406	Nicotiana tabacum	Unknow	Unknow
HM590407	Nicotiana tabacum	Unknow	China
HQ631374	Solanum tuberosum	2007-04-27	China
HQ912862	Solanum tuberosum	2006	USA

Solanum tuberosum	2007	USA
Solanum tuberosum	2007	USA
Solanum tuberosum	2000	USA
Solanum tuberosum	2007	USA
Solanum tuberosum	2007	USA
Solanum tuberosum	2007	USA
Solanum tuberosum	2004	USA
Solanum tuberosum	2007	USA
Solanum tuberosum	2007	USA
Solanum tuberosum	2004	USA
Solanum tuberosum	2005	USA
Solanum tuberosum	2007	USA
Solanum tuberosum	2004	USA
Solanum tuberosum	2005	USA
Solanum tuberosum	2005	USA
Solanum tuberosum	2006	USA
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Solanum tuberosum	2005	USA
Solanum tuberosum	2006	USA
Solanum tuberosum	2006	USA
Solanum tuberosum	2004	USA
Solanum tuberosum	2004	USA
Solanum tuberosum	2006	USA
Solanum tuberosum	2006	USA
Solanum tuberosum	2005	USA
Solanum tuberosum	2006	USA
	Solanum tuberosum Solanum tuberosum	Solanum tuberosum2007Solanum tuberosum2005Solanum tuberosum2007Solanum tuberosum2005Solanum tuberosum2005Solanum tuberosum2005Solanum tuberosum2005Solanum tuberosum2005Solanum tuberosum2005Solanum tuberosum2005Solanum tuberosum2006Solanum tuberosum2006Solanum tuberosum2006Solanum tuberosum2004Solanum tuberosum2004Solanum tuberosum2006Solanum tuberosum2005Solanum tuberosum2006Solanum tuberosum2006Solanum tuberosum2006Solanum tubero

HQ912890	Solanum tuberosum	2004	USA
HQ912891	Solanum tuberosum	2004	USA
HQ912892	Solanum tuberosum	2005	USA
HQ912893	Solanum tuberosum	2005	USA
HQ912894	Solanum tuberosum	2006	USA
HQ912895	Solanum tuberosum	2006	USA
HQ912896	Solanum tuberosum	2008	USA
HQ912897	Solanum tuberosum	2006	USA
HQ912898	Solanum tuberosum	2006	USA
HQ912899	Solanum tuberosum	2006	USA
HQ912900	Solanum tuberosum	2006	USA
HQ912901	Solanum tuberosum	2006	USA
HQ912902	Solanum tuberosum	2006	USA
HQ912903	Solanum tuberosum	2006	USA
HQ912904	Solanum tuberosum	2005	USA
HQ912905	Solanum tuberosum	2005	USA
HQ912906	Solanum tuberosum	2006	USA
HQ912907	Solanum tuberosum	2006	USA
HQ912908	Solanum tuberosum	2006	USA
HQ912909	Solanum tuberosum	2005	USA
HQ912910	Solanum tuberosum	2006	USA
HQ912911	Solanum tuberosum	2005	USA
HQ912912	Solanum tuberosum	2006	USA
HQ912913	Solanum tuberosum	2006	USA
HQ912914	Solanum tuberosum	2006	USA
HQ912915	Solanum tuberosum	2006	USA

JF928458	Solanum tuberosum	2008	Brazil
JF928459	Solanum tuberosum	2008	Brazil
JF928460	Solanum tuberosum	2008	Brazil
JQ924285	Solanum tuberosum	1985	Brazil
JQ924286	Solanum tuberosum	2009	Brazil
JQ924287	Solanum tuberosum	2007	Brazil
JQ924288	Solanum tuberosum	2007	Brazil
JQ969033	Solanum tuberosum	2010	Belgium
JQ969034	Solanum tuberosum	2010	Belgium
JQ969035	Solanum tuberosum	2010	Belgium
JQ969037	Solanum tuberosum	2010	Belgium
JQ969039	Solanum tuberosum	2010	Belgium
JQ969041	Solanum tuberosum	2010	Belgium
JX424837	Solanum tuberosum	2008	Finland
KC614702	Solanum tuberosum	2009	United Kingdom
KC634004	Solanum tuberosum	2009	United Kingdom
KC634005	Solanum tuberosum	2011	United Kingdom
KC634006	Solanum tuberosum	2009	United Kingdom
KC634007	Solanum tuberosum	2009	United Kingdom
KC634008	Solanum tuberosum	2009	United Kingdom
KF770835	Capsicum annuum	2010-10	South Africa
KF850513	Solanum tuberosum	2009-08-20	Mexico
KJ634023	Solanum tuberosum	2011-08-13	China
KJ634024	Solanum tuberosum	2011-08-13	China
KJ801915	Solanum tuberosum	2011-08-13	China
KJ946936	Solanum tuberosum	2013	Serbia

KM396648	Solanum tuberosum	2007	Slovenia
KP691317	Solanum tuberosum	2012	Australia
KP691318	Solanum tuberosum	2003	Australia
KP691319	Solanum tuberosum	1984	United Kingdom
KP691320	Solanum lycopersicum	2006	Australia
KP691321	Solanum tuberosum	1982	United Kingdom
KP691322	Solanum tuberosum	1982	United Kingdom
KP691323	Solanum tuberosum	1982	United Kingdom
KP691324	Solanum tuberosum	1980	United Kingdom
KP691325	Solanum tuberosum	2011	Australia
KP691326	Solanum tuberosum	1984	United Kingdom
KP691327	Solanum tuberosum	1943	United Kingdom
KP691328	Solanum tuberosum	2008	Australia
KP691329	Solanum tuberosum	2008	Australia
KP691330	Solanum tuberosum	1981	United Kingdom
KP793715	Solanum tuberosum	2014-08	Saudi Arabia
KP793716	Solanum tuberosum	2014-08	Saudi Arabia
KR149260	Solanum tuberosum	2015-02-02	Colombia
KR528584	Vitis vinifera	2013-05-21	Uruguay
KT290511	Solanum lycopersicum	2015-03-01	Colombia
KT290512	Solanum lycopersicum	2015-03-01	Colombia
KT336551	Solanum tuberosum	2015-03-01	Colombia
KT336552	Solanum tuberosum	2015-03-01	Colombia
KT599906	Solanum tuberosum	2014-08-27	Indonesia
KT599907	Solanum tuberosum	2014-08-27	Indonesia
KT599908	Solanum tuberosum	2014-08-27	Indonesia

Solanum lycopersicum	2012	China
Solanum lycopersicum	2012	China
Solanum tuberosum	2011-07-20	China
Nicotiana tabacum	2015-04-25	China
Nicotiana tabacum	2014-05-05	China
Solanum tuberosum	2014-06-25	China
Solanum tuberosum	2011-05-17	Israel
Solanum tuberosum	2014-05-04	Israel
Solanum tuberosum	2014-05-04	Israel
Solanum tuberosum	2011-05-04	Israel
Solanum tuberosum	1994	Poland
Solanum tuberosum	2015-03-01	Colombia
Solanum lycopersicum	2014-08-01	USA
Nicotiana tabacum	2015-03-19	China
Solanum tuberosum	2012-07-07	China
Solanum tuberosum	2012-07-25	China
Solanum tuberosum	2012-08-05	China
Solanum tuberosum	2011-08-23	China
Solanum tuberosum	2012-07-21	China
Solanum tuberosum	2013-07-04	China
Solanum tuberosum	2013	South Africa
Solanum tuberosum	2013	South Africa
	Solanum lycopersicum Solanum lycopersicum Solanum tuberosum Nicotiana tabacum Nicotiana tabacum Solanum tuberosum Solanum tuberosum Solanum tuberosum Solanum tuberosum Solanum tuberosum Solanum lycopersicum Nicotiana tabacum Nicotiana tabacum Nicotiana tabacum Nicotiana tabacum Nicotiana tabacum Solanum tuberosum Solanum tuberosum Solanum tuberosum Solanum tuberosum Solanum tuberosum Solanum tuberosum Solanum tuberosum	Solanum lycopersicum2012Solanum lycopersicum2012Solanum tuberosum2011-07-20Nicotiana tabacum2015-04-25Nicotiana tabacum2014-05-05Solanum tuberosum2014-06-25Solanum tuberosum2014-05-04Solanum tuberosum2014-05-04Solanum tuberosum2014-05-04Solanum tuberosum2014-05-04Solanum tuberosum2014-05-04Solanum tuberosum2011-05-04Solanum tuberosum2015-03-01Solanum tuberosum2015-03-01Solanum lycopersicum2015-03-19Nicotiana tabacum2015-03-19Nicotiana tabacum2015-03-19Nicotiana tabacum2015-03-19Nicotiana tabacum2015-03-19Nicotiana tabacum2012-07-07Solanum tuberosum2012-07-25Solanum tuberosum2012-07-25Solanum tuberosum2012-07-21Solanum tuberosum2013Solanum tuberosum2013

Solanum lycopersicum	2015-07-01	Slovakia
Solanum tuberosum	2015-03-01	Colombia
Solanum tuberosum	2015-01-19	Slovenia
Physalis peruviana	2016-08-01	Colombia
Nicotiana tabacum	2016	China
Solanum tuberosum	2016	United Kingdom
Solanum tuberosum	2005	USA
Solanum tuberosum	2004	USA
Solanum tuberosum	2006	USA
Solanum tuberosum	2004	USA
Solanum tuberosum	2012	USA
Solanum tuberosum	2005	USA
Solanum tuberosum	2006	USA
Solanum tuberosum	2013	USA
Solanum tuberosum	2013	USA
Solanum tuberosum	2005	USA
Solanum tuberosum	2005	USA
	Solanum lycopersicum Solanum tuberosum Solanum tuberosum Physalis peruviana Nicotiana tabacum Solanum tuberosum Solanum tuberosum	Solanum lycopersicum2015-07-01Solanum tuberosum2015-03-01Solanum tuberosum2015-01-19Physalis peruviana2016Nicotiana tabacum2016Solanum tuberosum2016Solanum tuberosum2005Solanum tuberosum2004Solanum tuberosum2012Solanum tuberosum2012Solanum tuberosum2012Solanum tuberosum2012Solanum tuberosum2012Solanum tuberosum2012Solanum tuberosum2013Solanum tuberosum2013 <tr <td="">Solanum tuberosum2</tr>

KY847957	Solanum tuberosum	2005	USA
KY847958	Solanum tuberosum	2006	USA
KY847959	Solanum tuberosum	2005	USA
KY847960	Solanum tuberosum	2005	USA
KY847961	Solanum tuberosum	2013	Germany
KY847962	Solanum tuberosum	2004	USA
KY847963	Solanum tuberosum	2005	USA
KY847964	Solanum tuberosum	2009	USA
KY847965	Solanum tuberosum	2010	USA
KY847966	Solanum tuberosum	2010	USA
KY847967	Solanum tuberosum	2010	USA
KY847968	Solanum tuberosum	2010	USA
KY847969	Solanum tuberosum	2010	USA
KY847970	Solanum tuberosum	2011	USA
KY847971	Solanum tuberosum	2011	USA
KY847972	Solanum tuberosum	2006	USA
KY847973	Solanum tuberosum	2006	USA
KY847974	Solanum tuberosum	2006	USA
KY847975	Solanum tuberosum	2009	USA
KY847976	Solanum tuberosum	2009	USA
KY847977	Solanum tuberosum	2011	USA
KY847978	Solanum tuberosum	2005	USA
KY847979	Solanum tuberosum	2006	USA
KY847980	Solanum tuberosum	2004	USA
KY847981	Solanum tuberosum	2004	USA
KY847982	Solanum tuberosum	2005	USA

Solanum tuberosum Solanum tuberosum	2006 2012	USA
Solanum tuberosum	2012	TIC A
	2012	USA
Solanum tuberosum	2012	USA
Solanum tuberosum	2010	USA
Solanum tuberosum	2010	USA
Solanum tuberosum	2010	USA
Solanum tuberosum	2004	USA
Solanum tuberosum	2005	USA
Solanum tuberosum	2004	USA
Solanum tuberosum	2010	USA
Solanum tuberosum	2011	USA
Solanum tuberosum	2004	USA
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Solanum tuberosum	2009	USA
Solanum tuberosum	2009	USA
	Solanum tuberosum Solanum tuberosum	Solanum tuberosum2012Solanum tuberosum2010Solanum tuberosum2010Solanum tuberosum2004Solanum tuberosum2004Solanum tuberosum2004Solanum tuberosum2004Solanum tuberosum2010Solanum tuberosum2010Solanum tuberosum2010Solanum tuberosum2011Solanum tuberosum2011Solanum tuberosum2004Solanum tuberosum2005Solanum tuberosum2004Solanum tuberosum2004Solanum tuberosum2005Solanum tuberosum2005Solanum tuberosum2005Solanum tuberosum2005Solanum tuberosum2004Solanum tuberosum2004Solanum tubero

17110 40000	G 1 1	2000	TIC A
KY848009	Solanum tuberosum	2009	USA
KY848010	Solanum tuberosum	2010	USA
KY848011	Solanum tuberosum	2010	USA
KY848012	Solanum tuberosum	2010	USA
KY848013	Solanum tuberosum	2010	USA
KY848015	Solanum tuberosum	2012	USA
KY848016	Solanum tuberosum	2012	USA
KY848017	Solanum tuberosum	2006	USA
KY848018	Solanum tuberosum	2005	USA
KY848019	Solanum tuberosum	2006	USA
KY848020	Solanum tuberosum	2004	USA
KY848021	Solanum tuberosum	2005	USA
KY848022	Solanum tuberosum	2005	USA
KY848023	Solanum tuberosum	2013	Germany
KY848024	Solanum tuberosum	2013	USA
KY848025	Solanum tuberosum	2009	USA
KY848026	Solanum tuberosum	2005	USA
KY848027	Solanum tuberosum	2005	USA
KY848028	Solanum tuberosum	2012	USA
KY848029	Solanum tuberosum	2012	USA
KY848030	Solanum tuberosum	2012	USA
KY848031	Solanum tuberosum	2004	USA
KY848032	Solanum tuberosum	2006	USA
KY848033	Solanum tuberosum	2004	USA
KY848034	Solanum tuberosum	2005	USA
KY848035	Solanum tuberosum	2005	USA

KY848036	Solanum tuberosum	2005	USA
KY848037	Solanum tuberosum	2004	USA
KY848038	Solanum tuberosum	2005	USA
KY848039	Solanum tuberosum	2005	USA
KY848040	Solanum tuberosum	2005	USA
KY848041	Solanum tuberosum	2004	USA
KY848042	Solanum tuberosum	2004	USA
KY848043	Solanum tuberosum	2004	USA
KY848044	Solanum tuberosum	2004	USA
KY848045	Solanum tuberosum	2004	USA
KY848046	Solanum tuberosum	2006	USA
KY848047	Solanum tuberosum	2005	USA
KY848048	Solanum tuberosum	2005	USA
KY848049	Solanum tuberosum	2005	USA
KY848050	Solanum tuberosum	2004	USA
KY848051	Solanum tuberosum	2004	USA
KY848052	Solanum tuberosum	2004	USA
KY848053	Solanum tuberosum	2007	USA
KY851109	Solanum tuberosum	2016	India
KY863548	Solanum tuberosum	2014	Egypt
KY863549	Solanum tuberosum	2014	Egypt
KY863550	Solanum tuberosum	2014	Egypt
KY863551	Solanum tuberosum	2014	Egypt
KY983389	Solanum americanum	2016-03-08	China
MF033142	Solanum tuberosum	2016-03-08	China
MF033143	Solanum americanum	2016-03-08	China

MF134861	Physalis peruviana	2015	USA
MF134862	Physalis peruviana	2015	USA
MF134863	Physalis peruviana	2015	USA
MF134864	Physalis peruviana	2015	USA
MF134865	Physalis peruviana	2015	USA
MF134866	Physalis peruviana	2015	USA
MF176826	Solanum tuberosum	2016-08-01	Colombia
MF176827	Solanum tuberosum	2016-08-01	Colombia
MF176828	Solanum tuberosum	2016-08-01	Colombia
MF405303	Solanum tuberosum	2013-08	Switzerland
MF422609	Solanum tuberosum	2014-07	Switzerland
MF422610	Solanum tuberosum	2014-07	Switzerland
MF624282	Solanum tuberosum	2014	USA
MF624283	Solanum tuberosum	2014	USA
MF624284	Solanum tuberosum	2012	USA
MF624285	Solanum tuberosum	2012	USA
MF624286	Solanum tuberosum	2015	USA
MF624287	Solanum tuberosum	2015	USA
MF624288	Solanum tuberosum	2016	USA
MF624289	Solanum tuberosum	2015	USA
MF624290	Solanum tuberosum	2015	USA
MF624291	Solanum tuberosum	2015	USA
MH006954	Solanum tuberosum	2015	Israel
MH006955	Solanum tuberosum	2015	Israel
MH006956	Solanum tuberosum	2015	Israel
NC_001616	Unknow	Unknow	Unknow

τ	J09509	Solanum tuberosum	Unknow	Canada

	Bean yellow mosaic virus										
Event	Recombinant ¹	Recom break	bination cpoints		Parents	Methods ²	P-value ³				
		Initial	Final	Major	Minor						
1	KF155409.2	1294	2191	Unknown	KF155420.2	<u>R</u> GBMCS3	1.26X10 ⁻¹⁰⁵				
2	HG970869.1	5820	8152*	HG970861.1	HG970859.1	RGBMC <u>S</u> 3	1.44x10 ⁻⁵⁵				
2	HG970865.1			HG970852.1	KF632713.1	 					
3	^KF155414.2	4485	9548*	Unknown	KF155420.2	<u>R</u> GBMCS3	$1.06 X 10^{-50}$				
4	HG970853.1	2974	4412	KF155409.2	Unknown	<u>R</u>GBMCS3	1.08X10 ⁻⁴³				
4	HG970851.1			KF155420.2	Unknown						
5	KF155419.2	1844	2073	HG970851.1	KF155420.2	RG <u>B</u> MCS3	2.72X10 ⁻²⁰				
5	KM114059.2			HG970853.1	KF155414.2						
6	^HG970868.1	50	108	Unknown	HG970867.1	RGBMC <u>S</u> 3	6.91X10 ⁻²²²				
7	HG970852.1	7084	8148	AB079888.1	Unknown		2 013/10-10				
7	HG970860.1				Unknown	RGBMC <u>S</u> 3	$2.01X10^{10}$				
7	HG970861.1				Unknown						
8	HG970860.1	6667	7065*	AB079888.1	Unknown	R <u>G</u> BMCS3	3.50X10 ⁻⁰⁹				

Supplementary Table S2. Recombination events detected by RDP in potyvirus datasets retrieved from the GenBank database.

8	HG970852.1				Unknown		
8	HG970861.1				Unknown		
9	HG970851.1	5210	5305	KF155414.2	Unknown	RG B MCS3	6.62X10 ⁻⁰⁷
9	HG970853.1				Unknown		
10	KF155414.2	2376	3233	KF155419.2	KF155409.2	RGBMC <u>S</u> 3	8.02X10 ⁻¹³
11	KF155414.2	1320*	1843*	Unknown	KF155420.2	RGBMC <u>S</u> 3	5.03X10 ⁻⁰⁸

Plum pox virus

		Recom break	bination points		Parents		
Event	Recombinant ¹	Initial	Final	Major	Minor	Methods ²	<i>P</i> -value ³
1	LC228949.1	1*	2726	AY953261.1	HF674399.1	RG <u>B</u> MCS	9.82X10 ⁻¹⁰⁶
2	^HG964686.1	1623	8688	LT600782.1	Unknown		
2	HG964685.1			LC333553.1	Unknown		
2	AY028309.2			LC333552.1	Unknown		
2	LN614587.1			LC331298.2	Unknown		
2	JX013532.1			LC333268.1	Unknown	RGBMCS3	7 53X10 ⁻⁶⁶
2	JQ794501.1			KU508427.1	Unknown		/.55/110
2	GU474956.1			KP998124.1	Unknown		
2	GU461889.1			LT158756.1	Unknown		
2	KM273015.1			KR028387.1	Unknown		
2	KF472134.1			KR028386.1	Unknown		

2	EU117116.1			KR028385.1	Unknown		
3	^MF370991.1	24*	1440	MF346263.1	D13751.1		
3	LC228949.1			MF346282.1_	LC333553.1_		
3	MF371004.1			MF346281.1	LC333552.1		
3	MF371003.1			MF346280.1	LC331298.2		
3	MF371002.1			MF346279.1	LC333268.1		
3	MF371001.1			MF346278.1	KU508427.1		
3	MF371000.1			MF346277.1	KP998124.1		
3	MF370999.1			MF346276.1	LT158756.1		
3	MF370997.1			MF346275.1	KR006730.1		
3	MF370996.1			MF346274.1	NC_001445.1		
3	MF370995.1			MF346273.1	KJ849228.1		
3	MF370994.1			MF346272.1	AB576080.1		
3	MF370993.1			MF346271.1	AB576079.1		
3	MF370992.1			MF346270.1	AB576078.1		
3	MF370990.1			MF346269.1	AB576077.1		
3	MF370989.1			MF346268.1	AB576076.1		
3	MF370987.1			MF346267.1	AB576075.1		
3	MF370986.1			MF346265.1	AB576074.1	RGBMCS <u>3</u>	0.02
3	MF370985.1			MF346264.1	AB576073.1		
3	MF370984.1			MF346262.1	AB576072.1		
3	HF585103.1			MF346261.1	AB576071.1		
3	HF585102.1			MF346260.1	AB576070.1		
3	HF585101.1			MF346258.1	AB576069.1		
3	HF585100.1			MF346257.1	AB576068.1		

3	HF585099.1	MF346256.1	AB576067.1	
3	HF585104.1	MF346255.1	AB576066.1	
3	M92280.1	MF346254.1	AB576065.1	
3	AJ243957.1	MF346253.1	AB576064.1	
3	FM955843.1	MF346252.1	AB576063.1	

Potato virus Y

Event	Recombinant ¹	Recomb Break	oination points		Parents	Methods ²	P-value ³
		Initial	Final	Major	Minor		
1	KX009783.1	1*	2450	KU724101.1	HQ912864.1		
1	KY863550.1			AJ585197.1	NC_001616.1	- <u>R</u> GBMCS 3	
1	AJ889868.1			KY847988.1	A08776.1		
1	KJ801915.1			KY847985.1	D00441.1		1.86X10 ⁻²⁰²
1	KU569326.1			KY847984.1	U09509.1		
1	KJ634023.1			KY847986.1	HM367076.1		
1	AJ890349.1			AM268435.1	KY848053.1		
1	KX688600.1			GQ200836.1	KP691317.1]	
2	KT290512.1	2461	5878	Unknown	JF928459.1		
2	KT290511.1			Unknown	KY800342.1	1	
2	KY711363.1			Unknown	KJ634024.1		
2	KX531041.1			Unknown	KX688598.1	<u>R</u> GBMCS3	3 1.53X10 ⁻¹⁷⁹
2	MF176828.1			Unknown	KX688597.1		
2	AB714135.1			Unknown	GQ200836.1		
2	AJ890346.1			Unknown	KY983389.1		

2	AB711144.1			Unknown	HM590405.1		
2	AJ585198.1			Unknown	KX650862.1]	
2	AJ585197.1			Unknown	HG810952.1]	
2	KY847988.1			Unknown	HG810951.1]	
2	KY847985.1			Unknown	KX688601.1]	
2	KY847984.1			Unknown	KX688602.1		
2	KY847986.1			Unknown	KX650860.1		
2	KU724101.1			Unknown	MF033143.1		
2	AM268435.1			Unknown	MF033142.1		
2	KY848025.1			Unknown	KY863549.1		
2	KY847954.1			Unknown	KP793715.1		
2	KY848024.1			Unknown	MH006956.1		
2	MF624288.1			Unknown	JQ969037.2		
2	KY847953.1			Unknown	AB185833.2		
2	KY848028.1			Unknown	AJ890342.1		
2	MF624290.1			Unknown	KX356068.1		
2	MF624291.1			Unknow	AJ889866.1		
2	MF624289.1			Unknown	HM590406.1		
2	KP793716.1			Unknown	AB461453.1		
2	KY847992.1			Unknown	AJ889867.1		
2	KX650859.1			Unknown	AB461451.1		
3	AJ889868.1	540	2369*	JF928459.1	FJ643479.1		
3	KJ801915.1			FJ666337.1	NC_001616.1	<u>R</u> GBMCS3	2.89X10 ⁻¹²⁶
3	AJ890349.1			KJ634024.1_	A08776.1		

3	KX688600.1	KX688598.1	D00441.1
3	KX032614.12	KX688597.1	U09509.1
3	JQ969039.2C	GQ200836.1	HM367076.1
3	HQ912868.1	KY983389.1	KY848053.1
3	KX710153.1	HM590405.1	KP691317.1
3	MF624284.1	JQ969037.2	KP691325.1
3	MF624287.1	JF928458.1	KY847991.1
3	KY847949.1	AB270705.1	KY848047.1
3	KY847952.1	JQ969033.2	KY847989.1
3	HQ912896.1	HQ631374.1	KY848034.1
3	KY847996.1	AJ890344.1	KY847967.1
3	MF624286.1	JQ969035.2	FJ643478.1
3	KY847977.1	KT599906.1	HQ912884.1
3	KY847943.1	HG810949.1	KY847947.1
3	KY847942.1	AB711143.1	HQ912883.1
3	KY848016.1	HG810950.1	KY847939.1
3	KY847945.1	JQ969034.2	HQ912877.1
3	KY847959.1	KT599907.1	HQ912899.1
3	HQ912863.1	AJ585342.1	HQ912900.1
3	KY847982.1	KT599908.1	HQ912904.1
3	KY848002.1	KY863551.1	KY848035.1
3	KY847981.1	AJ890345.1	HQ912908.1
3	KY847950.1	KC634008.1	KY847998.1
3	KY848021.1	KC634006.1	KY848045.1
3	AJ890350.1	KX184819.1	HQ912903.1
3	HE608963.1	KY851109.1	HQ912905.1

3	HE608964.1	KC634005.1	HQ912906.1	
3	JQ924288.1	KX184818.1	HQ912879.1	l
3	KX713170.1	JQ924287.1	HQ912907.1	
3	KX650858.1	KX710154.1	HQ912901.1	
3	KY847961.1	KY847948.1	KY848036.1	
3	KX184816.1	KY847946.1	KY848039.1	
3	JQ969041.2	KY847970.1	KY848042.1	
3	MH006955.1	KY847969.1	HQ912902.1	
3	KY800342.1	KY847971.1	KY847956.1	l
3	KX650861.1	MF405303.1	KY848046.1	
3	KX650862.1	JF928460.1	HQ912882.1	
3	HG810952.1	KY848008.1	HQ912881.1	
3	HG810951.1	KY847983.1	HQ912886.1	
3	KX688601.1	KM396648.1	KY847987.1	l
3	KX688602.1	KX856986.1	HQ912909.1	l
3	KX650860.1	KY847964.1	KY848038.1	l
3	MF033143.1	KY847975.1	KY848043.1	l
3	MF033142.1	MF422609.1	KY848044.1	
3	AJ890343.1	KT336552.1	KY848040.1	l
3	AB185833.2	KT336551.1	KY848041.1	
3	AJ890342.1	KX756672.1	HQ912880.1	
3	KX356068.1	KR149260.1	HQ912910.1	l
3	AJ889866.1	MF176827.1	KY848037.1	l
3	HM590406.1	KY847973.1	HQ912878.1	l
3	AB461453.1	KY847968.1	HQ912876.1	l
3	AJ889867.1	EF026075.1	KY847972.1	

3	AB461451.1			KJ946936.2	HQ912875.1		
3	AB461452.1			KX184817.1	KY848050.1		
4	HG810951.1_	5910*	8392	AJ890346.1_	KX713170.1_		
4	KX688601.1			KT290512.1	U09509.1		
4	KX688602.1			KT290511.1	HM367076.1		
4	MF033143.1			KY711363.1	KY848053.1		
4	MF033142.1			KX531041.1	KP691317.1	RGBMCS3	1.04 X 10 ⁻⁴⁶
4	KY863549.1			MF176828.1	KP691325.1	<u>KODM</u> <u>C</u> 55	5 1.042(10
4	KP793715.1			AB711144.1	KY847991.1		
4	AJ889867.1			AJ585198.1	KY848047.1		
4	AB461451.1			AJ585197.1	KY847989.1		
4	AB461452.1			KY847988.1	KY848034.1		
5	AB185833.2	5806	9290*	KC614702.1	JQ924286.1		
5	AJ890342.1			JQ969033.2	NC_001616.1		
5	KX356068.1			HQ631374.1	A08776.1	RGBMCS <u>3</u>	3.58X10 ⁻⁹⁵
5	AJ889866.1			AJ890344.1	D00441.1		
5	HM590406.1			JQ969035.2	U09509.1		
6	AJ889868.1	6759	9906*	JQ924286.1	AJ890346.1	RGMCS <u>3</u>	9.92X10 ⁻⁹²
7	KU569326.1	79*	737	KR528584.1	AB270705.1	RGBMCS	1.96×10^{-89}
7	JQ924286.1			AF237963.2	AB714135.1	<u>N</u> ODMCS	1.90x10

8	KJ634024.1	748*	8498	JF928459.1	KY848052.1		
8	KX688598.1			KT290512.1	NC_001616.1		
8	KX688597.1			KT290511.1	A08776.1		
8	GQ200836.1			KY711363.1	D00441.1		2 22X10 ⁻⁹²
8	KY983389.1			KX531041.1	U09509.1	RGBMCS3	
8	HM590405.1			AJ890346.1	HM367076.1	KGDMCS <u>5</u>	2.22/110
8	KY863549.1			AB711144.1	KY848053.1		
8	KP793715.1			AJ585198.1	KP691317.1		
8	AB270705.1			AJ585197.1	KP691325.1		
9	KY863550.1	2543	5959	KY848052.1	AF463399.1	PGPMCS3	1.04¥10-74
9	AJ889868.1			NC_001616.1	DQ309028.1	KODMCS <u>5</u>	1.94/10
10	KJ634024.1	128*	2438*	JF928459.1	KY847936.1		
10	KX688598.1			AJ890346.1	U09509.1		
10	KX688597.1			AB711144.1	HM367076.1		
10	GQ200836.1			AJ585198.1	KY848053.1	RGBMCS <u>3</u>	2.95X10 ⁻⁸³
10	KY983389.1			AJ585197.1	KP691317.1		
10	HM590405.1			KY847988.1	KP691325.1		
10	AB270705.1			KY847985.1	KY847991.1		
11	KY863548.1	683	2438*	FJ666337.1	KY848020.1		
11	KY848023.1			KT290512.1	NC_001616.1	RGBMCS <u>3</u>	3.50X10 ⁻⁷⁹
11	AM113988.1			AB714135.1	A08776.1		

12	KX650861.1	2528*	8622	KT599906.1_	KY848019.1		5.34X10 ⁻⁶⁷
12	KJ634024.1			KT290512.1	KP691317.1		
12	KX688598.1			AB714135.1	KP691325.1	RGBMCS <u>3</u>	
12	KX688597.1			AJ890346.1	FJ643478.1		
12	GQ200836.1			AJ585198.1	HQ912884.1		
12	KY983389.1			AJ585197.1	HQ912883.1		
12	HM590405.1			KY847988.1	KY847939.1		
12	HG810951.1			KY847986.1	HQ912877.1		
12	KX688602.1			KU724101.1	HQ912899.1		
12	KX650860.1			AM268435.1	HQ912900.1		
12	MF033143.1			KY848025.1	KY848035.1		
12	MF033142.1			KY847951.1	HQ912907.1		
12	AJ889867.1			KY847954.1	KY848036.1		
12	AB461451.1			KY848024.1	KY848039.1		
12	AB461452.1			MF624288.1	HQ912881.1		
12	AB270705.1			KY847953.1	HQ912873.1		
	AJ890347.1						
13	HG810952.1	9240	9869*	KY848031.1	KU724101.1	R <u>G</u> BM	3.42X10 ⁻⁵¹
13	JQ969037.2			U09509.1	KT290512.1		
13	JQ969033.2			HM367076.1	KT290511.1		
13	HQ631374.1			KY848053.1	KY711363.1		
13	AJ890344.1			KP691317.1	KX531041.1		
13	JQ969035.2			KP691325.1	MF176828.1		
13	KT599906.1			KY847991.1	AJ890346.1		
13	HG810949.1			KY848047.1	AB711144.1		

13	AB711143.1			KY847989.1	AJ585198.1		
13	HG810950.1			KY848034.1	AJ585197.1	1	
13	JQ969034.2			KY847967.1	KY847988.1	7	
13	KT599907.1			FJ643478.1	KY847985.1		
13	AJ585342.1			HQ912884.1	KY847984.1		
13	KT599908.1			KY847947.1	KY847986.1		
13	KY863551.1			HQ912883.1	AM268435.1		
13	AJ890345.1			KY847939.1	KY847954.1		
13	KC634008.1			HQ912877.1	KY848024.1		
13	KC634006.1			HQ912899.1	MF624288.1		
13	KX184819.1			HQ912900.1	KY847953.1		
13	KC634005.1			HQ912904.1	KY848028.1		
13	KX184818.1			KY848035.1	MF624290.1		
13	JQ924287.1			HQ912908.1	MF624291.1		
13	KX710154.1			KY847998.1	MF624289.1		
13	KY847948.1			KY848045.1	KY847992.1		
13	KY847946.1			HQ912903.1	JF928458.1		
13	KY847970.1			HQ912905.1	JF928459.1		
14	A08776.1	7970	8376	FJ204166.1	KY848012.1	R <u>G</u> BM	8.06X10 ⁻⁴²
14	D00441.1			KT290512.1	KX009783.1		
15	AB714135.1	90*	2442*	JF928459.1	JQ969033.2	RM <u>S</u> 3	5.28X10- ¹⁹
15	AB711144.1			AJ585197.1	KY847988.1		
15	AJ585198.1			KY847988.1	KY847985.1		
16	A08776.1	9629	9906*	KU724101.1	KY848012.1_	B GBM	1.65×10^{-11}
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16	D00441.1			KT290512.1	U09509.1		1.05/10
17	MF134862.1	2450	8462*	KY863549.1	HQ912914.1		
17	MF134865.1				NC_001616.1		
17	MF134864.1				A08776.1	RGB <u>S</u>	6.78x10 ⁻¹⁵
17	MF134866.1				D00441.1		
17	MF134861.1				U09509.1		

Soybean mosaic virus

		Recomb break	bination points				
Event	Recombinant ¹	Inital	Final	Major	Minor	Methods ²	<i>P</i> -value ³
1	FJ640977.1	24*	1825	AY216010.1	KP710865.1		
1	FJ640978.1			S42280.1	KT285170.1		
1	KC845321.1			NC_002634.1	KP710868.1	R <u>G</u> BMCS	6.43X10 ⁻⁹⁶
1	HQ845735.1			FJ640982.1	KP710866.1		
1	KM979229.1			FJ640981.1	KP710864.1		
2	FJ640970.1	7*	2954	FJ640967.1	KP710865.1		
2	FJ640976.1			S42280.1	FJ640978.1		
2	FJ640975.1			NC_002634.1	FJ640977.1	PCPMCS	1 51 1 10-80
2	FJ640974.1			FJ640979.1	KT285170.1	<u>R</u> <u>G</u> DMCS	S 1.51X10 ⁻⁰⁰
2	FJ640972.1			FJ640976.1	Unknown]	
2	FJ640971.1			FJ640975.1	Unknown		

2	FJ640966.1			FJ640974.1	Unknown		
2	FJ548849.1			FJ640972.1	Unknown		
3	^KF297335.1	63*	9060	FJ640955.1	KF135488.1		
3	KP710876.1			S42280.1	KP710871.1	RGMS <u>3</u>	3.41X10 ⁻⁵⁷
3	KP710874.1			NC_002634.1	KP710870.1		
4	KR065437.1	16*	1024	JF833013.1	JF833014.1	DCDMCS	2 21 V 10-56
4	KT285170.1			GU015011	JF833015.1	- <u>R</u> GDMCS	5.51710
5	FJ640981.1	5428	10349*	FJ640980.1	FJ548849.1		
5	AJ312439.1			FJ640970.1	S42280.1	RGBMS <u>3</u>	1.13X10 ⁻³⁷
5	AJ310200.1			FJ640957.1	NC_002634.1		
6	^HM590054.1	821	3579	AJ619757.1	JF833014.1	<u>R</u> GBMCS	6.39X10 ⁻³²
6	^HM590054.1	821	3579	AJ619757.1	JF833014.1	<u>R</u> GBMCS	6.39X10 ⁻³²
6	^HM590054.1 ^AJ619757.1	821 215*	3579 3423	AJ619757.1 JF833015.1	JF833014.1 KP710873.1	<u>R</u> GBMCS	6.39X10 ⁻³²
6 7 7	^HM590054.1 ^AJ619757.1 FJ640981.1	821 215*	3579 3423	AJ619757.1 JF833015.1 S42280.1	JF833014.1 KP710873.1 KP710875.1	<u>R</u> GBMCS <u>R</u> GBMCS	6.39X10 ⁻³² 4.30X10 ⁻³⁰
6 7 7 7 7	^HM590054.1 ^AJ619757.1 FJ640981.1 FJ640980.1	821	3579 3423	AJ619757.1 JF833015.1 S42280.1 NC_002634.1	JF833014.1 KP710873.1 KP710875.1 KP710872.1	<u>R</u> GBMCS <u>R</u> GBMCS	6.39X10 ⁻³² 4.30X10 ⁻³⁰
6 7 7 7 7	^HM590054.1 ^AJ619757.1 FJ640981.1 FJ640980.1	821 215*	3579 3423	AJ619757.1 JF833015.1 S42280.1 NC_002634.1	JF833014.1 KP710873.1 KP710875.1 KP710872.1	<u>R</u> GBMCS <u>R</u> GBMCS	6.39X10 ⁻³² 4.30X10 ⁻³⁰
6 7 7 7 7 8	^HM590054.1 ^AJ619757.1 FJ640981.1 FJ640980.1 ^FJ376388.1	821 215* 3458*	3579 3423 5234	AJ619757.1 JF833015.1 S42280.1 NC_002634.1 JF833014.1	JF833014.1 KP710873.1 KP710875.1 KP710872.1 FJ640982.1	<u>R</u> GBMCS <u>R</u> GBMCS	6.39X10 ⁻³² 4.30X10 ⁻³⁰
6 7 7 7 7 8 8 8	^HM590054.1 ^AJ619757.1 FJ640981.1 FJ640980.1 ^FJ376388.1 S42280.1	821 215* 3458*	3579 3423 5234	AJ619757.1 JF833015.1 S42280.1 NC_002634.1 JF833014.1 HM590054.1	JF833014.1 KP710873.1 KP710875.1 KP710872.1 FJ640982.1 FJ640971.1	<u>R</u> GBMCS <u>R</u> GBMCS <u>R</u> GBMCS <u>R</u> GBMCS <u>R</u> GBMCS	6.39X10 ⁻³² 4.30X10 ⁻³⁰ 1.62X10 ⁻¹⁶
6 7 7 7 7 8 8 8 8	^HM590054.1 ^AJ619757.1 FJ640981.1 FJ640980.1 ^FJ376388.1 S42280.1 NC_002634.1	821 215* 3458*	3579 3423 5234	AJ619757.1 JF833015.1 S42280.1 NC_002634.1 JF833014.1 HM590054.1 KM979229.1	JF833014.1 KP710873.1 KP710875.1 KP710872.1 FJ640982.1 FJ640971.1 AY216987.1	<u>R</u> GBMCS	6.39X10 ⁻³² 4.30X10 ⁻³⁰ 1.62X10 ⁻¹⁶
6 7 7 7 8 8 8 8	^HM590054.1 ^AJ619757.1 FJ640981.1 FJ640980.1 ^FJ376388.1 S42280.1 NC_002634.1	821 215* 3458*	3579 3423 5234	AJ619757.1 JF833015.1 S42280.1 NC_002634.1 JF833014.1 HM590054.1 KM979229.1	JF833014.1 KP710873.1 KP710875.1 KP710872.1 FJ640982.1 FJ640971.1 AY216987.1	<u>R</u> GBMCS <u>R</u> GBMCS <u>R</u> GBMCS <u>R</u> GBMCS <u>R</u> GBMCS	6.39X10 ⁻³² 4.30X10 ⁻³⁰ 1.62X10 ⁻¹⁶
6 7 7 7 8 8 8 8 8 9	^HM590054.1 ^AJ619757.1 FJ640981.1 FJ640980.1 ^FJ376388.1 S42280.1 NC_002634.1 ^JF833015.1	821 215* 3458* 1025*	3579 3423 5234 3457*	AJ619757.1 JF833015.1 S42280.1 NC_002634.1 JF833014.1 HM590054.1 KM979229.1 AY294045.1	JF833014.1 KP710873.1 KP710875.1 KP710872.1 FJ640982.1 FJ640971.1 AY216987.1 HM590055.1	<u>R</u> GBMCS <u>R</u> GBMCS <u>R</u> GBMCS <u>R</u> GBMCS <u>R</u> GBCS3	6.39X10 ⁻³² 4.30X10 ⁻³⁰ 1.62X10 ⁻¹⁶

9	JF833014.1			NC_002634.1	KP710868.1		
10	^AJ619757.1 _for_polyprote	3835*	4893*	FJ640964.1	KX834324.1		
10	FJ640980.1			FJ640982.1	FJ640976.1	DCC	4 00371 0-28
10	FJ640970.1			FJ640969.1	FJ640975.1	RGS <u>3</u>	4.22×10^{20}
10	FJ640957.1			FJ640965.1	FJ640974.1		
10	FJ640954.1			FJ640961.1	FJ640972.1		
11	^HM590055.1	1058*	3593	FJ640969.1	KP710876.1		
11	FJ640977.1			Unknown	FJ640967.1	-	
11	KT285170.1			Unknown	FJ640963.1	MCS <u>3</u>	3.85X10 ⁻¹²
11	KP710868.1			Unknown	KY986929.1		
11	KP710866.1			Unknown	KX834324.1		
12	^FJ640971.1	3811	4893*	Unknown	KX834319.1		
12	FJ640982.1			Unknown	FJ640977.1		
12	FJ640969.1			Unknown	FJ640976.1	RGB3	7.96×10^{-12}
12	FJ640965.1			Unknown	FJ640975.1	100 <u>5</u>	7.70/110
12	FJ640964.1			Unknown	FJ640974.1		
13	FJ640978.1	1826*	3457*	Unknown	FJ640979.1		
13	KP710865.1			Unknown	S42280.1	RGB <u>3</u>	3.58X10 ⁻¹¹
13	KX834320.1			Unknown	NC_002634.1		1
14	^HQ845736.1	6346	10337*	FJ640964.1	KP710871.1	RCB3	2 20X10-09
14	GU015011.1			FJ640982.1	KF135488.1		2.27/10

15	FJ376388.1	6598	8128	FJ807700.1	FJ640980.1	PGCS	2 82X10-09
15	FJ807701.1_			S42280.1	FJ640982.1	KOC <u>S</u>	2.03A10
16	FJ376388.1	5262*	6597*	FJ640980.1	FJ807700.1		
16	FJ640982.1			FJ640970.1	FJ640972.1	PGS3	1.06 V 10 ⁻³⁴
16	FJ640981.1			FJ640957.1	FJ548849.1	K05 <u>5</u>	1.00/10
16	FJ640961.1			FJ640954.1	AY294045.1		
17	FJ376388.1	8140*	9235	FJ640954.1	FJ807700.1	PBCS3	1 32X10-16
17	FJ807701.1			FJ640980.1	FJ548849.1	KDC3 <u>5</u>	H. <i>J</i> 2 A 10
18	^FJ640969.1	36*	3457*	FJ640979.1	Unknown		
18	FJ640982.1			S42280.1	Unknown	RBC <u>S</u> 3	1.21X10 ⁻¹²
18	FJ640971.1			NC_002634.1	Unknown		
19	^FJ640982.1	3682*	6215	KR065437.1	Unknown	PGPS	1 17 X 10 ⁻⁰³
19	FJ640980.1			KP710871.1	Unknown	KOD <u>5</u>	1.1/A10

Sugarcane mosaic virus

		Recomb break	bination points				
Events	Recombinant ¹	Initial	Final	Major	Minor	Methods ²	<i>P</i> -value ³
1	^JX047417.1	1*	4452	JX047394.1	JX047426.1		
1	JX047428.1			JX047395.1	JX047431.1	RGRMCS	4 50X10 ⁻²⁰¹
1	JX047427.1				JX047430.1	KO <u>D</u> MCS	4.30/10
1	JX047422.1				JX047429.1		

1	JX047419.1				JX047425.1		
1	JX047418.1				JX047424.1		
2	^JX047419.1	7266	9640*	JX047417.1	JX047394.1		
2	JX047427.1			JX047431.1	JX047395.1	<u>R</u> GBMCS	3.43X10 ⁻¹²⁹
2	JX047422.1			JX047430.1			
3	^EU091075.1	5050	9630*	Unknown	AY149118.1 _		
3	GU474635.1			Unknown	AF494510.1	<u>R</u> GMCS	3.05X10 ⁻⁹³
3	MG932078.1			Unknown			
4	^AY149118.1	5050	9636*	AJ297628.1	AM110759.1	DGMCS	7 21 V 10-90
4	AF494510.1			JX047384.1		<u>R</u> OMCS	7.31A10
4							
5	^KR611114.1	4900	9345*	KR611105.1	Unknown	PGMCS	2 12X10 ⁻⁵²
5	KR611106.1			KR611113.19	Unknown	<u>R</u> OMC3	2.12A10
6	^MG932079.1	2008	5138	AM110759.1	Unknown		
6	AF494510.1					PGMCS	5 11 X 10-45
6	MG932078.1					KOMC <u>5</u>	J.44A10
6	AY149118.1						
7	^KT895080.1	17*	1425	MG932080.1	Unknown	PCPS	8 00×10-23
7	KT895081.1			MG932077.1	Unknown	KOD <u>5</u>	0.07/10

8	^MG932078.1	8127*	8591*	Unknown	AM110759.1	PGP3	3 89×10 ⁻²⁹
8	MG932079.1			Unknown		KOD <u>5</u>	5.69/10
9	^JX047404.1	46*	2092	JX047428.1	Unknown		
9	JX047397.1_			JX047395.1	Unknown	RGBMCS	1 21 X 10 ⁻²⁹
9	JX047393.1			JX047394.1	Unknown	KODMC <u>5</u>	1.21/10
9	AY042184.1				Unknown		
10	^JX047393.1	2286	5189	Unknown)	JX047394.1		
10	JX047404.1			Unknown	JX047395.1	RCS3	4 13X10 ⁻¹¹
10	JX047397.1			Unknown		<u> </u>	4.15/10
10	AY042184.1			Unknown			
11	^JX047418.1	8158*	8919	JX047401.1	Unknown		
11	JX047428.1			JX047430.1			
11	JX047427.1			JX047426.1		PGM3	2 08V10 02
11	JX047422.1			JX047425.1		KOWI <u>5</u>	5.06A10-05
11	JX047419.1			JX047423.1			
11	JX047417.1			JX047416.1			
12	^KT895080.1	7951	8208	Unknown	AJ310103.1		0.03
12	KT895081.1				AJ310102.1		0.05
13	^JX047394.1	9490*	9527	KR108213.1	JX047417.1	RGBS3	4.64X10-35

13	JX047404.1			
13	JX047395.1			

	Zucchini mosaic virus										
		Recomb break	oination points		Methods ²	<i>P</i> -value ³					
Events	Recombinant ¹	Inital	Final	Major	Minor						
1	^AY279000.1	2927	6400	KX884570.1	KX421104.1	DCMCS	2 15V10-18				
1	AY278999.1			KX884565.1	AB369279.1	<u>R</u> OMCS	5.45710				
2	^KF976713.1	170	5746*	KC665635.1	KY225553.1						
2	KU528623.1			L31350.1	KY225555.1						
2	KU198853.1			KY225548.1	KY225554.1		2.95X10 ⁻²⁰				
2	KT778297.1			KY225547.1	KY225552.1						
2	KF976712.1			KJ875865.1	KY225551.1	RGBMS3					
2	JN183062.1			KJ875864.1	KY225550.1	KODM <u>5</u> 5					
2	AY188994.1			KJ923769.1	KY225549.1						
2	EF062583.1			KJ923768.1							
2	EF062582.1			KJ923767.1							
2	DQ124239.1			KC665634.1							
3	^KY225556.1	7426	9791*	KY225542.1	KY225545.1	PGPMS	Q 04X10-13				
3	KY225544.1			KY225546.1			7.04A10				
4	^AJ316229.2	30*	2362	AB188115.1	KX499498.1	<u>R</u> GBS	5.65X10 ⁻⁰⁹				

4	KT598222.1			DD056806.1			
5	^KY225551.1	2746	6058	KY225547.1	Unknown		
5	KY225555.1			L31350.1 _	Unknown		
5	KY225554.1			KY225548.1	Unknown		
5	KY225553.1			KJ875865.1	Unknown	RGC <u>S</u>	7.07X10 ⁻⁰⁶
5	KY225552.1			KJ875864.1	Unknown		
5	KY225550.1			KJ923769.1	Unknown		
5	KY225549.1			KJ923768.1	Unknown		
6	^KY225556.1	318	2520	KY225545.1	L29569.1 _	DDMS	1 05 V 10 ⁻¹⁸
6	KY225544.1			Unknown	Unknown	KDM <u>5</u>	1.93A10
7	^AJ307036.2	328	1354	KC665635.1	KX499498.1		
7	AM422386.1			L31350.1			
7	AJ316228.2			KY225548.1			
7	NC_003224.1			KY225547.1		<u>R</u> GBS	3.42X10 ⁻⁰⁵
7	AF127929.2			KJ875865.1			
7	DJ418432.1			KJ875864.1			
7	DI159774.1			KJ923769.1			
8	^KX421104.1	3866	6620	Unknown	KC665635.1		
8	KX249747.1			Unknown	L31350.1	RGBS	3 76X10 ⁻¹⁰
8	AY278998.1			Unknown	KY225548.1	100 <u>5</u>	5./0/10
8	AJ515911.1				KY225547.1		

8	AB369279.1				KJ875865.1		
9	^AJ316228.2	6406*	9065	AB188115.1	KU528623.1		
9	AM422386.1			L31350.1			
9	NC_003224.1			DD056806.1		RGB3	8 19X10 ⁻¹⁷
9	AF127929.2			KY225547.1		KGD <u>5</u>	0.17/10
9	DJ418432.1			KX664482.1			
9	DI159774.1			KJ875865.1			
10	^AB369279.1	7093	9005	Unknown	AB188116.1		
10	KX421104.1			Unknown	DD056806.1		0.005
10	AY279000.1			Unknown	AB188115.1	DCMC	
10	AY278999.1			Unknown		KO <u>M</u> C	0.003
10	AY278998.1			Unknown			
10	AJ515911.1			Unknown)			
11	^AY279000.1	872	1406	KC665631.1	AB188115.1		
11	KX421104.1			L31350.1	DD056806.1		
11	KX249747.1			KY225548.1	AB188116.1	RGB <u>M</u>	0.026
11	AJ515911.1			KJ875864.1			
11	AB369279.1			KJ923769.1			

¹Numbering starts at the first nucleotide after the cleavage site at the origin of replication and increases clockwise.

² R, RDP; G, GeneConv; B, Bootscan; M, MaxChi; C, CHIMAERA; S, SisScan; 3, 3SEQ.

³The reported *P*-value is for the program in bold, underlined type and is the lowest *P*-value calculated for the event in question.



Supplementary Figure S1. Secondary structure prediction, (a) Complete genome structure of BYMV, (b) Complete genome structure of ZYMV, (c) Complete genome structure of PRSV, (d) Complete genome structure of LMV, (e) Complete genome structure of PPV, (f) Complete genome structure of SMV. Demarcations points in all the figures indicate the type of secondary structure in each genome position with different colors: stem (purple), bulge (green), multiloop (blue), interior loop (brown).



Supplementary Figure S2. Intrinsically disordered protein regions analysis. The output of *Bean yellow mosaic virus* (BYMV) proteins. Those regions in which the red lines of the graph reach the line indicate the presence of disordered protein regions.



Supplementary Figure S3. Intrinsically disordered protein regions analysis. The output of *Lettuce mosaic virus* (LMV) proteins. Those regions in which the red lines of the graph reach the line indicate the presence of disordered protein regions.



Supplementary Figure S4. Intrinsically disordered protein regions analysis. The output of *Plum pox virus* (PPV) proteins. Those regions in which the red lines of the graph reach the line indicate the presence of disordered protein regions.



Supplementary Figure S5. Intrinsically disordered protein regions analysis. The output of *Papaya ringspot virus* (PRSV) proteins. Those regions in which the red lines of the graph reach the line indicate the presence of disordered protein regions.



Supplementary Figure S6. Intrinsically disordered protein regions analysis. The output of *Soybean mosaic virus* (SMV) proteins. Those regions in which the red lines of the graph reach the line indicate the presence of disordered protein regions.



Supplementary Figure S7. Intrinsically disordered protein regions analysis. The output of *Zucchini Yellow mosaic virus* (ZYMV) proteins. Those regions in which the red lines of the graph reach the line indicate the presence of disordered protein regions.



Supplementary Figure S8. Sequence composition analyses of (a) Complete genome structure of BYMV, (b) Complete genome structure of LMV, (c) Complete genome structure of PRSV, (d) Complete genome structure of PPV, (e) Complete genome structure of SMV, (f) Complete genome structure of ZCMV. The lines indicate the full-length genome of GC percentage in each figure.