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BACTERIÓFAGOS RECOMBINANTES E miRNAs COMO
ESTRATÉGIAS PARA O CONTROLE DE *Spodoptera*
frugiperda

PATOS DE MINAS – MG
FEVEREIRO DE 2019

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Dissertação de Mestrado apresentada ao Programa
de Pós-graduação em Biotecnologia como
requisito parcial para obtenção do título de Mestre
em Biotecnologia.

Orientadora: Profa. Dra. Thaise Gonçalves Araújo

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RESUMO

A lagarta *Spodoptera frugiperda* é considerada uma das principais pragas agrícolas no Brasil, responsável por severos danos a culturas importantes. Inseticidas sintéticos e biopesticidas têm enfrentado limitações como o aumento no impacto ambiental e resistência dos insetos às toxinas recombinantes. A biotecnologia se mostra uma alternativa promissora ao permitir o desenvolvimento de estratégias efetoras para o combate da lagarta. O objetivo deste estudo foi gerar novas moléculas para o controle de *S. frugiperda* por meio da ferramenta biotecnológica de *Phage Display* (PhD), ao selecionar peptídeos ligantes a proteínas intestinais, e prever microRNAs por meio de ferramentas de bioinformática. Utilizando a tecnologia de PhD, conseguimos selecionar sete clones cuja capacidade de se ligar a proteínas intestinais de *S. frugiperda* foi validada por ELISA e imuno-histoquímica. O clone SfF3, quando usado em combinação com a toxina de *Bacillus thuringiensis*, promoveu um aumento da mortalidade das lagartas neonatas. Além disso, demonstrou similaridade com as proteínas cassetes de ligação de ATP subfamília C2 e citocromo P450, as quais são importantes para a sobrevivência desta praga. Quanto aos miRNAs, foram preditos 350 precursores de microRNAs presentes no genoma de *S. frugiperda*, sendo 60 específicos para o gênero *Spodoptera*. Além disso, realizamos o refinamento de bibliotecas de RNA-seq disponíveis no banco de dados do NCBI na busca de possíveis moléculas bioinseticidas e 91 destas sequências ainda não foram descritas na literatura, evidenciando as perspectivas de novas sequências enquanto possíveis alternativas para a regulação de transcritos e consequente alteração na biologia da lagarta. Nossos resultados validam essas tecnologias como uma abordagem agrobiotecnológica para o controle de pragas e identifica novas moléculas com atividade bioinseticidas, podendo ser utilizadas combinadas ou isoladamente.

Palavras-chave: Lagarta do cartucho, Phage Display, RNAi, peptídeos, bioinseticidas.

ABSTRACT

The caterpillar *Spodoptera frugiperda* is considered one of the main agricultural pests in Brazil, responsible for severe damage to important crops. Synthetic insecticides and biopesticides have faced limitations such as increased environmental impact and insect resistance to recombinant toxins. Biotechnology is a promising alternative that allow the development of effective strategies to caterpillar combat. The objective of this study was to generate new molecules for the control of *S. frugiperda* using the biotechnological tool of Phage Display (PhD), when selecting ligand peptides to intestinal proteins, and to predict microRNAs through bioinformatics tools. Using the technology of PhD, we were able to select seven clones whose ability to bind intestinal proteins of *S. frugiperda* was validated by ELISA and immunohistochemistry. Clone SfF3, when used in combination with *Bacillus thuringiensis* toxin, promoted an increase in mortality in newborn caterpillars. In addition, it demonstrated similarity to the ATP binding cassette subfamily C2 and cytochrome P450 proteins, which are important for the survival of this pest. As for the miRNAs, 350 precursors of microRNAs present in the genome of *S. frugiperda* were predicted, being 60 specifics for the genus *Spodoptera*. In addition, we performed the refinement of RNA-seq libraries available in the NCBI database in search of possible bioinsecticides molecules and 91 of these sequences have not yet been described in the literature, evidencing the perspectives of new sequences as possible alternatives for the regulation of transcripts and consequent alteration in the biology of the caterpillar. Our results validate these technologies as an agrobiotechnological approach to pest control and identify new molecules with bioinsecticide activity, which can be used in combination or in isolation.

Key words: Fall armyworm, Phage Display, iRNA, peptides, bioinsecticides.

LISTA DE ABREVIATURAS E SÍMBOLOS

AGO-1	Argonauta-1
ABCC2	cassetes de ligação de ATP subfamília C2
AMFE	Energia Livre Mínima Ajustada
Bt	<i>Bacillus thuringiensis</i>
C-	Controle Negativo
C+	Controle Positivo
DAB	3, 3-diaminobenzidina
Div	Diversidade de conjuntos
dsRNA	RNA dupla fita
Freq	Conjunto de Energia Livre Termal
GC content	Conteúdo de Guanina e Citosina
MFE	Energia Livre Mínima
MFEE	Energia Livre Mínima do Conjunto Termodinâmico
MFEI	Índice Mínimo de Energia Livre
miRNA	microRNA
mRNA	RNA mensageiro
NTC	Controle não alvo
PhD	<i>Phage Display</i>
RISC	Complexo silenciador induzido por RNA
RNAi	RNA de interferência
scL	Cistatina da soja L
scN	Cistatina da soja N
siRNA	pequeno RNA de interferência
ssDNA	DNA circular de cadeia simples
ssDNA	DNA circular de cadeia simples
WT	Fago selvagem

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INTRODUÇÃO

A lagarta-do-cartucho (*Spodoptera frugiperda*), da ordem Lepidoptera e família Noctuidae, é considerada uma das principais pragas no Brasil. Os danos provocados por este inseto podem reduzir a produtividade do milho em até 53% da produção média anual, com valor estimado de perdas de 6.187 milhões de dólares (Abrahams et al., 2017). Apesar das tentativas de combate químico, a severidade do ataque da lagarta-do-cartucho aumentou em várias regiões brasileiras, o que tem evidenciado a necessidade de novas estratégias para seu controle. A biotecnologia surge como uma alternativa ao oferecer métodos inovadores de interesse agrônomo.

Dentre as ferramentas disponíveis, o *Phage Display* (PhD) vem sendo amplamente utilizado nas mais diferentes áreas, incluindo saúde e agropecuária. Esta tecnologia é uma plataforma de seleção molecular, em que peptídeos ou fragmentos de anticorpos podem ser bioengenhierados e selecionados contra alvos de interesse. No PhD, peptídeos aleatórios são expressos em fagos e uma biosseleção é realizada com a finalidade de se obter ligantes altamente específicos. Uma das principais vantagens inclui a seleção sem o conhecimento prévio do alvo, o que não compromete sua eficácia. Além disso, no PhD, há uma conexão entre o fenótipo (o peptídeo exibido no fago) e o seu genótipo (o DNA que o codifica), viabilizando a análise dos recombinantes e sua posterior seleção. A técnica é eficiente, robusta e específica, além de se mostrar promissora na obtenção de ligantes, que, por sua vez, podem ser conduzidos ao controle de pragas agrícolas através da seleção de moléculas com ação inseticida.

Em nível transcricional, os microRNAs (miRNAs) se destacam. Estas moléculas estão envolvidas na regulação pós-transcricional silenciando, por RNA de interferência (RNAi), genes específicos. Este mecanismo serve como a principal linha de defesa antiviral, tanto em plantas quanto em insetos (Gan et al., 2008). Portanto, sua manipulação se traduz em perspectivas ao administrar, através da injeção ou do alimento do inseto, esses miRNAs. Sua principal vantagem é a especificidade ao gene-inseto-alvo, permitindo uma abordagem mais ecológica reduzindo os impactos ambientais na agricultura.

Nesse trabalho hipotetizou-se que peptídeos expressos na superfície de bacteriófagos filamentosos e miRNAs possam combater ou auxiliar no controle de *S. frugiperda*. Para isso, foram selecionados fagos recombinantes contra proteínas do intestino da lagarta, posteriormente validados em ensaios *in vivo*. Adicionalmente,

precursores de miRNAs foram preditos a partir do genoma de *S. frugiperda*, permitindo identificar possíveis moléculas com efeito inseticida. Oferecemos assim, ferramentas para uma dupla estratégia na contenção dessa praga que ainda compromete a produção e o desenvolvimento econômico do país. Novos caminhos são, portanto, abertos na busca de alternativas biotecnológicas e inovadoras voltadas para a agricultura.

OBJETIVOS

O objetivo primordial deste trabalho foi desenvolver novas moléculas para o controle de *S. frugiperda* utilizando as ferramentas biotecnológicas de *Phage Display* para selecionar peptídeos ligantes a proteínas intestinais, e ferramentas de bioinformática para identificar possíveis miRNAs com ação inseticida. Secundariamente, objetivamos estabelecer uma estratégia eficiente de biosselação e predizer para a obtenção de moléculas biologicamente ativas.

CAPÍTULO I

FUNDAMENTAÇÃO TEÓRICA

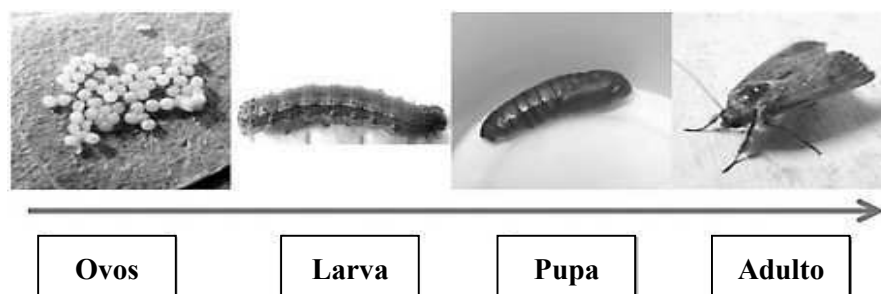
Normas ABNT

1 *Spodoptera frugiperda*

A *S. frugiperda* é considerada uma das principais pragas agrícolas no Brasil, e pode se alimentar de mais de 100 espécies de plantas de uma ampla gama de famílias (CABI, 2018). Este inseto ocorre naturalmente em regiões tropicais e subtropicais. Contudo, a alimentação diversificada e disponível durante todo o ano, além das condições climáticas favoráveis, agrava sua distribuição geográfica em solo brasileiro (Cruz, 1995; Cruz, 1999; Cruz *et al.*, 2009).

O ciclo de vida desta lagarta (Figura 1), sob condições laboratoriais controladas, dura aproximadamente 30 dias e inicia quando os ovos são colocados em massa na parte superior das folhas. Após 3 dias, a 25°C, estes eclodem e originam as lagartas. Inicialmente, estas se alimentam da casca dos próprios ovos e depois raspam as folhas mais novas da planta, ocasionando o sintoma característico denominado “folhas raspadas”. À medida que cresce, a larva se desloca para a região do cartucho, onde ocasiona severos danos. No interior dessa estrutura, o período larval varia de 12 a 30 dias. Quando completamente desenvolvida, a lagarta sai do cartucho e penetra no solo, onde se transforma em pupa, com aproximadamente 15 mm de comprimento. A fase de pupa dura, em média, de 10 a 12 dias. Já a mariposa mede cerca de 35 mm de envergadura e possui coloração das asas anteriores parda-escura e posteriores branca-acinzentada, com pontos claros na região central de cada asa. A longevidade do adulto é de aproximadamente 12 dias. Durante o estágio larval, as lagartas podem passar por até sete instares, dependendo da temperatura e do tipo de alimento. A larva recém-nascida tece um fio de seda que é usado como meio de dispersão e escape de inimigos naturais. No final dessa fase, chega a atingir 50 mm de comprimento (Cruz, 1995).

Figura 1 - Ciclo de vida da *S. frugiperda*. Inicialmente, os ovos são colocados em massa na parte superior das folhas, então eclodem e dão origem as lagartas. Quando completamente desenvolvida, a lagarta sai do cartucho e penetra no solo, onde se transforma em pupa e em seguida origina ao adulto.



Fonte: Adaptado de Schmidt-Duran *et al.* (2015)

As lagartas danificam o limbo foliar das folhas mais novas, produzindo injúrias de “raspagem”. Já as lagartas maiores perfuram as folhas e se desenvolvem no cartucho do milho. Os danos biológicos provocados diminuem a área fotossintética da planta comprometendo sua produtividade. Apesar das lesões não serem devastadoras, abrem orifícios que facilitam a penetração de patógenos. O controle ineficaz desses indivíduos pode reduzir a produtividade do milho em até 53%, valor correspondente a US \$ 6.187 milhões (Abrahams *et al.*, 2017)

O principal controle desta praga é a utilização de inseticidas sintéticos da classe de carbamatos, organofosforados e piretróides. O uso destes produtos promove o surgimento de populações resistentes e gera impactos ao ambiente e inimigos naturais, além de possuírem um elevado custo econômico (Martinelli e Omoto, 2006). A utilização de biopesticidas, como as toxinas Cry produzidas por *B. thuringiensis* (Bt), também é uma importante forma de manejo, proporcionando bons resultados e reduzindo o impacto ambiental (Bobrowski *et al.*, 2003). Evidências sugerem que essas toxinas se ligam a proteínas da membrana do epitélio do intestino médio, levando à lise ou conduzindo à morte celular (Bravo *et al.*, 2007). Apesar das vantagens, uma das dificuldades do uso dos produtos Bt é a susceptibilidade reduzida das larvas do terceiro instar, comparada as mais novas (Ferro e Lyon, 1991; Navon, 2000). Além disso, o número de casos de resistência das plantas geneticamente modificadas para expressar a toxina Bt mais do que triplicou desde 2002 (Tabashnik e Carrière, 2017).

A severidade dos danos causados também aumentou em várias regiões brasileiras. A eliminação dos inimigos naturais, aparecimento de populações resistentes a inseticidas, além da maior disponibilidade de alimento, criam condições favoráveis para a sobrevivência do inseto (Cruz, 1999). Nesse contexto, novas estratégias de controle tornam-se necessárias, envolvendo tecnologias inovadoras que possam gerar moléculas efetoras no controle dessa praga.

2 PHAGE DISPLAY E microRNAs PARA O CONTROLE DE PRAGAS

A biotecnologia emerge como uma promissora alternativa ao oferecer ferramentas para o desenvolvimento de novos ligantes. O *Phage Display* (PhD) se destaca como uma plataforma de seleção molecular, em que peptídeos ou fragmentos de anticorpos podem ser bioengenheirados e selecionados contra alvos específicos. Os miRNAs, por sua vez, são moléculas de pequeno tamanho (20 a 23 nucleotídeos) envolvidas na regulação pós-transcricional. Estes pequenos RNAs permitem o silenciamento gênico mediado por RNA de interferência (RNAi) de alvos específicos e, portanto, com nítida interface econômica. Tratam-se, portanto, de duas técnicas que, isoladamente ou combinadas, podem introduzir novos elementos no controle de pragas, transpondo inúmeras barreiras químicas até então enfrentadas pela economia do país.

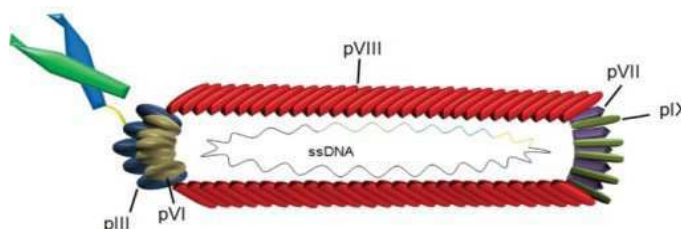
2.1 PHAGE DISPLAY

PhD é uma ferramenta molecular utilizada para apresentar, enriquecer e amadurecer a afinidade de vários peptídeos/proteínas a partir de uma biblioteca com mais de 2×10^9 variantes expressa em vetores virais. A apresentação dos ligantes envolve a utilização de bacteriófagos, os quais são manipulados geneticamente para expressar uma molécula exógena fusionada a uma de suas proteínas, em geral de seu capsídeo (Smith, 1985; Scott e Smith, 1990).

Bacteriófagos filamentosos da família Inoviridae (M13, f1 e fd), parasitas de bactérias gram negativa que contêm o pílus F, são os vetores utilizados nesse processo. A partícula do fago M13, mostrada na Figura 2, é formada por um DNA circular de cadeia simples (ssDNA) cercado por proteínas de revestimento (pIII, pVI, pVII, pVIII e pIX),

formando o capsídeo viral. Existem aproximadamente 2700 cópias da pVIII e, em uma das extremidades do fago, 5 cópias da pIII e pVI, estas envolvidas na ligação à célula hospedeira e no término do processo de montagem da partícula viral, respectivamente. Na outra extremidade, são encontradas 5 cópias de pVII, responsável pelo início da montagem viral e pIX, essencial para a manutenção da estabilidade do fago (Phizicky e Fields, 1995; Webster, 2001).

Figura 2 - Estrutura da partícula de fago M13 exibindo uma proteína exógena fusionada à região N-terminal da proteína de revestimento pIII. As demais proteínas do capsídeo viral pIII, pVI, pVII, pVIII e pIX também são demonstradas, e em seu interior, o DNA fita simples circular.



Fonte: Adaptado de Kierny *et al.* (2012)

O bacteriófago necessita da maquinaria de replicação, transcrição e tradução da bactéria para sua reprodução. A infecção das células de *Escherichia coli* se inicia com a interação entre a proteína pIII e a parte superior do pilus F, um tubo proteico composto por subunidades de pilina, as quais são despolimerizadas retraindo o pilus e internalizando o fago. Após o DNA viral penetrar no citoplasma, é convertido em uma molécula replicativa dupla fita e o material genético do vírus é utilizado como molde para a transcrição e tradução de suas proteínas. Após a síntese da proteína pV, forma-se o complexo DNA-pV, utilizado na montagem do bacteriófago entre a parede e membrana celulares. O fago M13 não provoca a lise da célula hospedeira, mas induz um estado em que a bactéria origina e libera continuamente partículas virais, ocasionando uma queda na sua taxa de reprodução (Maneewannakul *et al.*, 1993; Tikunova e Morozova, 2009).

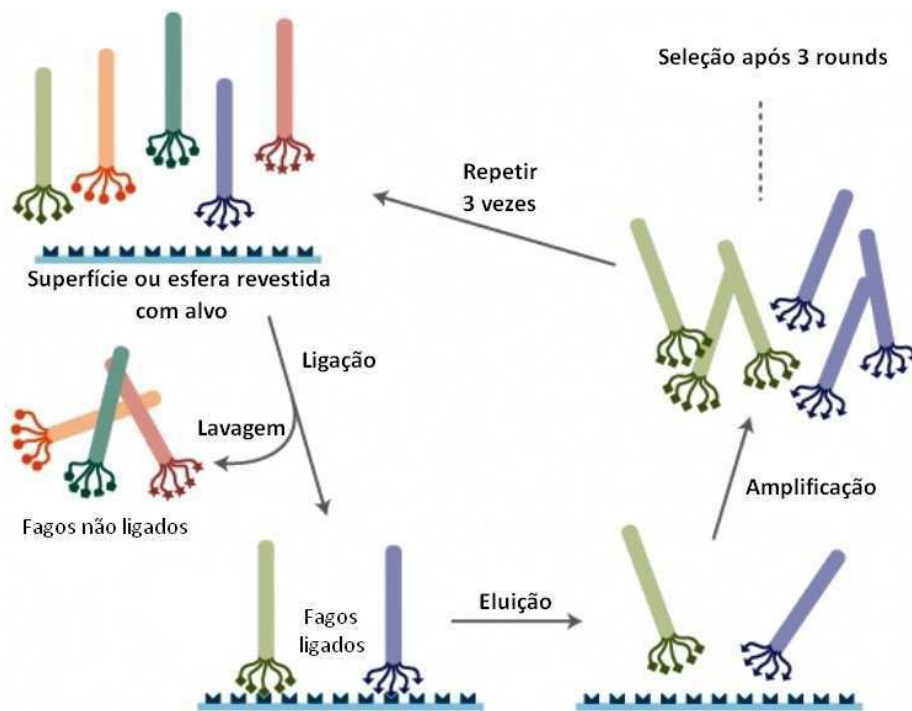
No PhD, o gene exógeno é fusionado, em geral, na região N-terminal das proteínas de revestimento pIII ou pVIII. A escolha da proteína que exibe a biblioteca determina a valência do processo. Utilizando a pIII, as 5 cópias podem expressar peptídeos curtos sem interferir na infectividade do fago. Quando se utiliza a pVIII, aproximadamente 10% destas podem ser alteradas, o que determina uma elevada valência (aproximadamente 200

cópias por vírus) em detrimento da afinidade durante o processo de seleção de recombinantes. Portanto, o sistema baseado na expressão em pIII permite a seleção de ligantes de alta afinidade. O processo de seleção para apresentar, enriquecer e amadurecer a afinidade é denominado de *biopanning* ou biosseleção (Gram *et al.*, 1992; Peltomaa *et al.*, 2016).

O *biopanning* consiste em ciclos ou *rounds* sequenciais de exibição, seleção e enriquecimento (Figura 3). No primeiro toda a biblioteca é exposta ao alvo e uma subpopulação de fagos específicos permanece ligada, enquanto o restante dos vírus é removido por lavagens. As partículas de fagos que ligaram ao alvo são eluídas, e em seguida, infectam bactérias para serem amplificadas em meios seletivos, permitindo aumentar o número de cópias de fagos específicos. A eluição dos ligantes pode ser realizada com pH extremo, clivagem de protease ou com o uso de competidores. O produto do primeiro *round* é utilizado nos ciclos seguintes permitindo enriquecer os recombinantes que se ligam especificamente ao alvo de interesse (Tikunova e Morozova, 2009; Kierny *et al.*, 2012). Para a seleção, o antígeno pode ser imobilizado em suporte sólido ou mantido em solução, utilizando-se partículas que se movem livremente, como esferas paramagnéticas (Kierny *et al.*, 2012).

Figura 3 - Representação do processo de seleção (*biopanning*). A biblioteca de fagos recombinantes é incubada com o antígeno alvo (imobilizado ou em solução). As partículas virais não ligantes são removidas por sucessivas lavagens; as específicas são eluídas (eluição ácida ou competitiva) e posteriormente amplificadas através da infecção

de bactérias. O produto do primeiro *round* é utilizado nos ciclos seguintes permitindo enriquecer os fagos com maior afinidade.



Fonte: Adaptado Biolabs (2017).

O PhD se destaca em relação a outras metodologias por permitir uma ligação direta entre o genótipo encapsulado e o fenótipo expresso em fusão, o que facilita a caracterização da proteína após o processo de seleção. Além disso, são construídas bibliotecas que variam entre 10^6 a 10^{11} variantes expressos, o que amplia a possibilidade de se obter ligantes de interesse (Nixon *et al.*, 2014). Não somente peptídeos, mas também anticorpos, ou até mesmo seus fragmentos, podem ser expressos e selecionados a partir de bacteriófagos recombinantes. A utilização de fragmentos de anticorpos é uma alternativa interessante, pois são menores e apresentam propriedades farmacocinéticas e farmacodinâmicas melhoradas, embora falte a função efetora (Chames *et al.*, 2009; Aghebati-Maleki *et al.*, 2016).

A não utilização de animais de laboratório, a relativa simplicidade dos protocolos e a possibilidade de rastrear uma grande quantidade de moléculas candidatas em um curto período de tempo torna o PhD vantajoso. Peptídeos e anticorpos são selecionados contra diferentes antígenos, incluindo substâncias tóxicas ou vírus altamente perigosos, os quais, por razões éticas, não podem ser utilizados na imunização (Koiwa *et al.*, 1998; Carmen e Jermutus, 2002; Nascimento *et al.*, 2009; Tikunova e Morozova, 2009).

A versatilidade dessa tecnologia é traduzida em diferentes estratégias e moléculas selecionadas com atividades defensivas. Koiwa *et al.* (1998) identificaram o papel inseticida de duas isoformas proteicas de cistatinas da soja, a cistatina da soja N (scN) e a cistatina da soja L (scL) por meio da técnica de PhD. Após o *biopanning*, a afinidade de ligação da scN foi superior a scL, sendo capaz de retardar o crescimento e desenvolvimento de *Callosobruchus maculatus*.

Também expressas na superfície viral, variantes de inibidores de tripsina foram selecionadas quanto à sua capacidade de ligação ao alvo. A Chy8 se mostrou altamente tóxica para as ninfas de *Acyrtosiphon pisum* e moderadamente tóxica para as ninfas de *Aphisgossypii* e *Myzus persicae*, além de cerca de oito vezes mais eficiente que a selvagem. Em outro estudo, variantes da toxina produzida pelo Bt foram exibidas e selecionadas contra o vetor *Aedes aegypti*. Duas moléculas, Cry1Aa13-A8 e Cry1Aa13-A12, foram capazes de induzir uma alta mortalidade (90%) nas populações larvais do mosquito. Outro mutante, Cry1Aa13-A10, não demonstrou efeito no estágio larval, mas induziu a mortalidade tardia das pupas, evidenciando a aplicabilidade do PhD enquanto estratégia para o controle de diferentes pragas (Domínguez-Flores *et al.* (2017).

Considerando o princípio dessa metodologia, o PhD se mostra como uma alternativa biotecnológica interessante no desenvolvimento de inibidores contra pragas agrícolas, gerando novas moléculas inseticidas ou tornando as existentes mais efetivas. Esta tecnologia permite selecionar mutantes contra proteínas ou moléculas receptoras intestinais de insetos, e estas podem mostrar uma nova atividade inseticida, ou podem melhorar a toxicidade contra um alvo específico, ou mesmo demonstrar ser ativa contra insetos resistentes (Bravo *et al.*, 2013).

2.2 microRNAS

Os microRNAs (miRNAs) e os pequenos RNAs de interferência (siRNAs) são moléculas utilizadas em ensaios para o silenciamento. Possuem propriedades físico-químicas e função semelhantes, mas mecanismos distintos (Ahmadzada *et al.*, 2018). Ambos são duplex de RNAs curtos formados por uma cadeia ativa (guia) e uma cadeia complementar inativa (passageiro). Os siRNAs são considerados RNAs exógenos e apresentam complementariedade total ao seu alvo, promovendo, assim, a clivagem do esqueleto fosfodiéster. Os miRNAs são endógenos e reprimem a tradução ao conduzir a um emparelhamento de bases imperfeito. Uma diferença importante no que concerne à

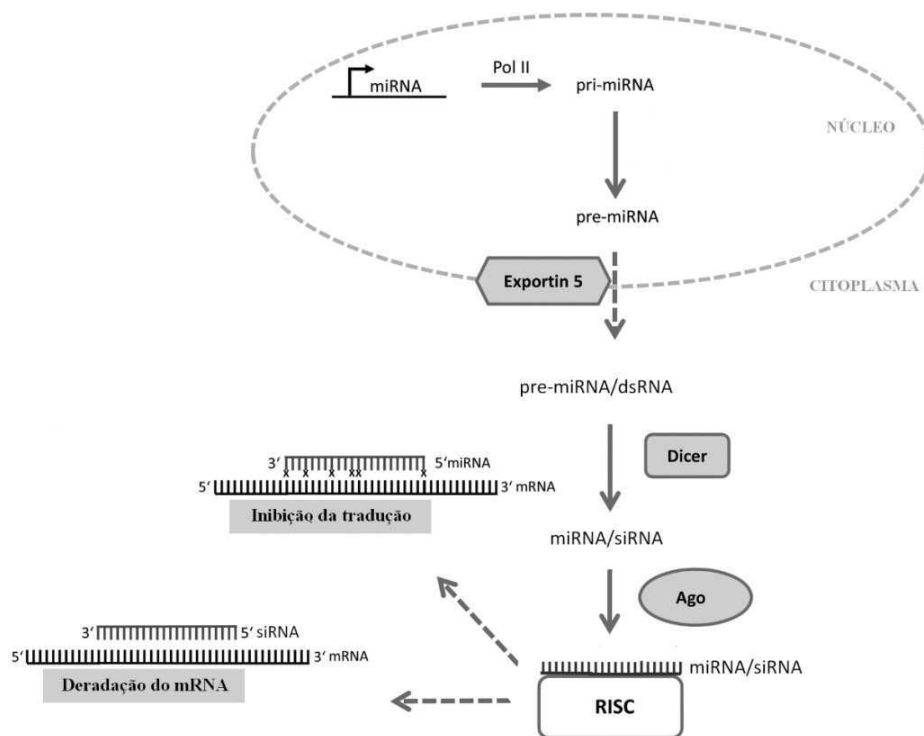
atividade dessas moléculas é a complementariedade do siRNA a uma sequência específica de um único mRNA, inibindo a expressão de um único gene. Já o miRNA pode ter múltiplos alvos e regular diferentes genes em decorrência de seu reconhecimento imperfeito (Lam *et al.*, 2015). Contudo, é possível identificar, por meio de análises *in silico*, miRNAs específicos a um determinado alvo. Os algoritmos utilizados permitem gerenciar a sensibilidade ou o nível de precisão das moléculas preditas (Akhtar *et al.*, 2015).

Para ambos, o silenciamento envolve o mecanismo de RNAi. Primeiramente, um longo RNA dupla fita (dsRNA) expresso ou introduzido na célula é digerido em pequenos RNAs de cadeia dupla pela enzima Dicer. Em seguida, as fitas são separadas e uma destas, conhecida como guia, é acoplada ao complexo RISC. Este é dirigido para a cadeia alvo de seu RNA mensageiro (mRNA) complementar. Após a ligação ocorre o bloqueio da tradução ou degradação do alvo (Winter *et al.*, 2009).

No caso dos miRNAs, estes são primeiramente transcritos em pri-miRNA no núcleo da célula pela RNA polimerase II ou III e possuem sequências complementares em tandem que permitem a dobradura da molécula, gerando um dsRNA (Siomi e Siomi, 2009). Os pri-miRNAs são processados a uma molécula precursora, o pré-miRNA (65 a 70 nucleotídeos). Estes precursores são transportados para o citoplasma pela Exportin 5, onde são clivados em dsRNAs curtos (miRNAs) pela Dicer. A cadeia com menor estabilidade de combinação de bases é carregada no complexo RISC por ligação direta à proteína Argonauta-1 (AGO-1). O complexo RISC/miRNA é guiado até o mRNA para que ocorra a regulação de sua expressão. O grau de complementaridade com o mRNA é crucial para determinar o mecanismo de interferência. Quando ocorre uma complementaridade parcial entre a sequência da extremidade 5' do miRNA (*seed sequence*) com a região do mRNA alvo, é desencadeada a inibição da tradução. O silenciamento por siRNA se inicia com a introdução de uma molécula de dsRNA artificial na célula. Esta é processada pela Dicer em siRNA e também carregada no complexo RISC. Nesse caso, a complementariedade total entre o mRNA e o siRNA conduz à degradação da molécula alvo. Ambos os mecanismos são descritos na Figura 4.

Figura 4 - Mecanismos de silenciamento de genes por miRNA e siRNA. O silenciamento através dos miRNAs inicia-se pela transcrição do pri-miRNA pela RNA polimerase II ou III gerando um dsRNA. Os pri-miRNAs originam ao pré-miRNA que é transportado para o citoplasma pela Exportin 5 e clivado em dsRNA curto (miRNA) pela

Dicer. Uma das cadeias é carregada no complexo RISC por ligação direta à proteína AGO-1 e este complexo é guiado até o mRNA para que ocorra a regulação de sua expressão. A complementaridade parcial entre a *seed sequence* com a região do mRNA alvo desencadeia a inibição da tradução. Já o silenciamento por siRNA se inicia com a introdução de uma molécula de dsRNA artificial na célula. Esta é processada pela Dicer em siRNA e também carregada no complexo RISC. Nesse caso, a complementariedade total entre o mRNA e o siRNA conduz à degradação da molécula alvo



Fonte: Adaptado de Wagner *et al.* (2015)

O RNAi é um mecanismo de regulação pós-transcricional, encontrado em plantas e animais, incluindo insetos (Burand e Hunter, 2013) e visa silenciar a expressão de genes endógenos ou de invasores, parasitas e patógenos, como vírus e elementos genéticos móveis, incluindo os transposons. Como é a principal linha de defesa em plantas, é considerado parte de seu sistema imune inato (Gordon e Waterhouse, 2007). Este mecanismo vem sendo extensivamente explorado para o desenvolvimento de métodos para controle de pragas e vetores de doenças. Várias pesquisas mostraram que as moléculas de dsRNAs administradas por injeção ou ingestão são capazes de penetrar nas células e ativar a via de silenciamento (Matranga e Zamore, 2007; Van Rij e Berezikov, 2009; Berezikov, 2011). Um exemplo é o estudo desenvolvido por Mogilicherla *et al.* (2018), em que foi realizada a administração oral de dsRNA em *Halyomorpha halys* com

a consequente morte do inseto. A administração oral do miR-2703 em ninfas de *Nilaparvata lugens* silenciou o gene A da síntese de quitina, também levando a fenótipos letais (Li *et al.*, 2017). Já o silenciamento dos genes alatostatina C e alototropina 2 de *S. frugiperda*, por RNAi, promoveu uma redução nas taxas de oviposição das fêmeas adultas (Griebler *et al.*, 2008). Em 2018, o grupo de pesquisa de Wang inibiu o gene da calmodulina, relacionada ao desenvolvimento de insetos, utilizando dsRNA em *N. lugens* com consequentes alterações fenotípicas e a mortalidade dos indivíduos tratados (Wang *et al.*, 2018).

A escolha dos miRNAs para serem usados como estratégia inseticida exige minuciosas análises *in silico*, de forma a garantir sua especificidade ao inseto de interesse. miRNAs, ao contrário de siRNAs, são conservados (Grody *et al.*, 2009). Interessantemente, ao definir como alvo essas sequências conservadas, há uma maior chance de se promover a morte do inseto, pois regiões importantes para o funcionamento do organismo seriam silenciadas (Lai, 2005).

De fato, essas moléculas são potencialmente promissoras. A sequência usada pode ser cuidadosamente desenhada para o uso contra uma única praga, ou grupo de espécies relacionadas, sem afetar insetos não-alvo. Assim, a principal vantagem desta tecnologia é sua alta especificidade e explorar esse mecanismo se mostra interessante, especialmente no manejo de pragas agrícolas (Whyard *et al.*, 2009; Parsons *et al.*, 2018).

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CAPÍTULO II

INSECTICIDAL RECOMBINANT PHAGES AGAINST *Spodoptera*
frugiperda: NEW PERSPECTIVES FOR BRAZILIAN AGRICULTURE

Instructions according to **Plos one**

Title: Insecticidal recombinant phages against *Spodoptera frugiperda*: new perspectives for Brazilian agriculture

Short title: New recombinant phages to control *Spodoptera frugiperda*

Autorship

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Abstract

The *Spodoptera frugiperda* is an important pest that promotes large productivity losses of maize. Insecticidal proteins from *Bacillus thuringiensis* has promoted important advances for this pest control. However, the occurrence of resistant insects challenges this technology, and has become important to develop new effective products through innovative techniques in the selection of new binders. The aim of this study was to select recombinant phages expressing gut exogenous peptides binders with insecticidal activity against *S. frugiperda*. Using Phage Display technology, seven clones were successful selected and bound to intestinal proteins. SfF3 clone when combined with the *B. thuringiensis* toxin, has increased mortality of *S. frugiperda* neonates, and also demonstrated similarity to the truncated ABC transporter subfamily C2 and cytochrome P450, which are important proteins to this pest survival. Our innovative study validated the reliability of Phage Display technology as an agribiotechnological approach for pest control, expanding the options to identify new molecules with bioinsecticides activities.

Keywords: Fall armyworm, peptides, phage display, bioinsecticides

Introduction

The fall armyworm (*Spodoptera frugiperda*) is a polyphagous insect that feeds on more than 60 species of plants, being an important pest of corn, rice, sorghum and cotton in Americas [1]. This species can reduce maize production in up to 53% of the annual averaged production, with estimated loss of \$6,187m [2].

The climatic conditions are determinant for the occurrence of lepidopteran pests, interfering in the development, behavior and feeding of the insects [3]. In Brazil, corn

cultivation in two seasons in the central-west region favors the occurrence of high infestations during all phases of plant growth [4]. In the southern, planting is carried out from August to January, thus, the first crops result in low infestations, but in the late months there is an increase in the pest population density [5]. Therefore, frequent applications of insecticides are necessary to reduce the insect population and avoid economic damages [6]. Bioinsecticidal molecules that lead to reduced use of chemical agents have been prominent in the area. In fact, the introduction of transgenic maize lines expressing insecticidal proteins from *B. thuringiensis* (Bt) is considered one of the most successful biotechnology achievements for the pest control. However, reports of insects resistance to transgenic crops arose in 2006 in TC1507 corn in Puerto Rico [6-8].

This event has also been reported in Brazil. Farias et al. [5] detected resistance of *S. frugiperda* to TC1507 corn in Brazil. Besides, low frequency of resistant alleles to *B. thuringiensis* has already been identified in fall armyworm population, demonstrating the potential evolution risk of resistance in Brazil [9]. Farias et al. [10] also demonstrated a significant reduction in susceptibility to the Cry1F toxin in *S. frugiperda* over the years 2010 to 2013, especially in regions with intensive maize production in Brazil. In this context, it becomes necessary to develop new effective products through innovative techniques in the selection of new binders.

Phage Display (PhD) is a high throughput profiling technology based on peptide libraries present on the surface of bacteriophage [11]. This technique has become a tool for finding high affinity bioactive peptides exploring the interaction sites between targets and ligands [12,13]. Therefore, through this technology, it is possible to trace and identify new peptides that bind to the midgut proteins of the caterpillar. Oligopeptides can therefore be identified through an affinity selection strategy, making this tool extremely suitable for the identification of new molecules that act as biopesticides [14].

Although there are no reports of recombinant phage peptides for *S. frugiperda*, different studies have been conducted with this methodology, with particularly promising results. Koiwa et al. [15] used the PhD technology to identify the insecticidal role of two protein isoforms of soybean cystatin, N (scN) and L (scL). After the biopanning, the binding affinity of scN was superior to scL, being able to retard the growth and development of *Callosobruchus maculatus*. The trypsin inhibitor variant Chy8 was selected by PhD by Ceci et al. [16] and was toxic to *Acyrtosiphon pisum*, *Aphisgossypii* and *Myzus persicae* nymphs. In another study, variants of the toxin produced by Bt were displayed on recombinant phages and selected against *Aedes aegypti*, and two molecules, Cry1Aa13-A8 and Cry1Aa13-A12, were able to induce a high mortality (90%) in the larval populations of this mosquito [17].

Therefore, PhD is an innovative agribiotechnological approach for pest control and was explored in this work. The aim was to select recombinant phages expressing gut exogenous peptides binders with insecticidal activity against genotypes of *S. frugiperda* adapted for Brazil.

Materials and Methods

Spodoptera frugiperda and protein extraction

Fifteen larvae of *S. frugiperda* (Lepidoptera: Noctuidae) from the Laboratório de Bio Controle Farroupilha/LALLEMAND (Patos de Minas, MG) were used. They were maintained at $25 \pm 2^\circ\text{C}$, $70 \pm 10\%$ of relative humidity and 12 hours of photophase. Fall armyworm close to the last instar remained fasted for 24 hours. After this period, they were immobilized for 15 minutes at -4°C to be dissected in saline solution (NaCl 0.215 mM). The first and last segments were withdrawn and the intestine removed and stored at -80°C .

The intestines were microdissected and the extraction buffer (125 mM Tris-HCl pH 7.0, 100 mM NaCl, 0.5% Triton X-100, 0.5% Tween-20, e 0.1% Genapol C-100 or SDS 0.1%) was

added (125mg/mL). Then, the sample was homogenized on ice for 1 minute using a homogenizer, followed by 1 minute of rest. This step was repeated five times. After centrifugation at 10,000 x g for 15 minutes at 4°C, the supernatant was discarded, and 100mM phenylmethylsulfonyl fluoride, 1μL 100mM benzamidine and 1μL phosphatase inhibitor cocktail was added to the extracted proteins.

Phage Display and biopanning

The Ph.D.-C7CTM Peptide Library Kit (New England Biolabs) was used for phage selection. Wells of a 96-well high binding plate (Nunc, Denmark), one for each round, were coated with 10μg of midgut proteins in bicarbonate buffer (0.1M NaHCO₃, pH 8.6) overnight at 4°C followed by blocking with NaHCO₃-BSA 5% (0.1M NaHCO₃, pH 8.6; BSA 5mg/mL) for 1h. Wells were washed six times with TBST (Tris-buffered saline containing 0.1% Tween 20), and proteins were incubated with phage library (10¹¹/100uL) diluted in TBST for 1h at 37°C. The supernatant (with non-binding particles) was removed through 10 washes with TBST, and the remained bound phages were eluted in 50μL of 0.2 M glycine buffer, pH 2.0 for 10 min. Neutralization of acid pH was performed with 1 M Tris-base pH 9.0.

The phages were amplified using early-log *Escherichia coli* ER2738 as a host, purified by PEG-8000/NaCl precipitation and titered on LB/IPTG/Xgal plates as described previously [18]. The selection was repeated for two more times (a total of three rounds for enrichment) each one using the previous amplified eluted phages.

Phage-ELISA

For screening of selected clones, 96-well Maxisorp microtiter plates (Nunc, Denmark) were coated with 5μg of midgut protein in 100 mM NaHCO₃, pH 8.6 and overnight at 4°C. A

separate set of wells without gut proteins was coated with blocking buffer TBS-BSA 5% (Tris-buffered saline, BSA 5%) and used as no target control (NTC). Plates were washed twice with TBS 1X and then blocked with TBS-BSA 5%, for 90 min at 37°C. After washing with TBS 1X, culture supernatant (50 µL) from 54 amplified phage particles randomly picked from the second round ($\sim 10^{10}$ pfu/mL) were incubated for 1 h at 37°C. The wells were washed four times with TBS-T 0.1% followed by incubation with the peroxidase-conjugated anti-M13 antibody (GE Healthcare, Chicago, IL) diluted 1:5000 in blocking buffer at 37 °C for 1 hour.

Plates were further washed four times in TBS-T 0.1%, revealed with OPD SigmaFast™ (Sigma-Aldrich) and read at 492 nm. The ELISA assay was performed in triplicate for each of the 54 clones. The same protocol was performed to evaluate the rate of recovered phage clones using the supernatant of *E. coli* infected with the pool of phages from each round.

Phage-clone binding

A new format-ELISA assay was conducted for the seven selected clones (SfF3, SfC4, SfD5, SfB6, SfH6, SfF7 and SfC8). The immunoassay was conducted as above, and the culture supernatant was replaced by 1×10^9 , 1×10^{10} and 1×10^{11} phages/well. The wild type M13 phage vector (no peptide) was also purified and used as control. Phages were purified as previously published [18].

DNA extraction and *in silico* analysis

For DNA extraction and sequencing, isolated colonies from the seven purified phages that were submitted to the above ELISA were transferred to DeepWell plates. DNA extraction and sequencing were performed according to Ferreira et al. [19].

The amino acid sequences were deduced according to the nucleotide sequences using the tool available in ExPASy Bioinformatics Resources Portal (<http://web.expasy.org/translate/>). Peptide sequences were also analyzed with SAROTUP program to identify redundant sequences. Similarity of the peptides to proteins was performed using BLASTp (<http://www.ncbi.nlm.nih.gov/BLAST>) against the proteins of *S. frugiperda*.

The peptides were modeled by the de novo method using the PEPFOLD3 - RPBS software (<http://mobyli.rpbs.univ-paris-diderot.fr/cgi-bin/portal.py#forms::PEP-FOLD3>). Clusters and standard parameters of the program were used (Label: PEPFOLD; Type of simulation: Short) without structural reference. As input, it was submitted the FASTA amino acid sequences of each peptide flanked by cysteines. The program ranks the five best hits based on the free energy and melting temperature and provides the structural odds chart according to the amino acid sequence (output). For this work, it was selected only the first best result of each peptide.

Immunohistochemistry

Caterpillars close to the last instar remained fasted for 24 hours. Subsequently, they were immobilized for 15 minutes at -4°C to be dissected in saline solution (NaCl 0.215 mM). The first and last segments were removed to extraction of the gut, which were fixed in modified Zamboni solution (10% formaldehyde, 15% picric acid in 0.1 μM sodium phosphate buffer) for 24 h at 4°C [20]. Then, they were washed in 10% neutral buffered formalin (1M of formaldehyde, 33mM of sodium phosphate monobasic, 46mM of dibasic sodium phosphate) at 4°C until complete removal of the picric acid. Dehydration of slides was carried out in baths of increasing concentrations of ethanol (70%, 85%, 100%) to include in paraffin.

For immunohistochemical staining, the sections were submitted to antigenic recovery with 10mM citrate buffer pH 6 in microwaves for 15 minutes after xylol and alcohol baths

(100%, 85%, 70%). Endogenous peroxidase was blocked with 3% H₂O₂ (vol. 10). Non-specific sites were blocked with 10% BSA in PBS 1X for 40 min. 1×10^{11} phages/slide diluted in buffer (Novocastra IHC Diluent) were incubated for 30 minutes at 37°C and then overnight at 4°C in a humid chamber. After washing with PBS, the slides were incubated for 1h at 37°C with HRP-labeled anti-M13 (1: 650) (GE Healthcare, Chicago, IL). The slides were revealed with NovoLink polymer (Leica Biosystems) according to the manufacturer's recommendations, 0.5 mg/mL 3, 3-diaminobenzidine (DAB) (Sigma-Aldrich), and counterstaining with Harris Hematoxylin (Dinamica). Histological analysis was performed using a microscope (Aperio-Leica Biosystems) and the images were captured (20x).

***In vivo* assay**

The bioassays were conducted adapting the droplet feeding method [21,22]. Briefly, the solution for the negative control (C-) was prepared using 1 g/L of blue food powder dye (Iceberg), 25 mM sucrose and PBS 1X. This solution was used for the preparation of all other treatment. For the positive control (C +), was used then insecticide Agree® at 1×10^6 CFU/μL. For the treatment with the strain of *B. thuringiensis* GF 07 from Laboratório de Bio Controle Farroupilha/LALLEMAND we used 3×10^3 CFU/μL to visualize the increment of mortality promoted by the peptides. We used 1×10^{11} phages/μL for the seven selected clones (SfF3, SfC4, SfD5, SfB6, SfH6, SfF7 and SfC8). Wild-type phage were used as a control to check whether the exogenous non-peptide phage is capable of killing larvae. Twenty-five newborn hatchlings were used for each of the treatments in trials with five replicates using completely randomized design. The larvae were isolated and stored within 2mL tubes and 0.5μL of each treatment was deposited for ingestion. After five hours, the larvae were transferred to vessels with artificial diet without formaldehyde and incubated at room temperature. After 3 days, the insects were

transferred to a new container containing artificial diet with 25% formaldehyde. Mortality assessments were performed after seven days.

Statistical analyzes

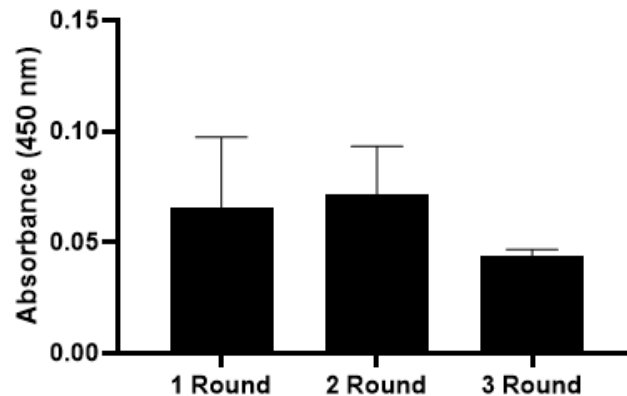
Results were expressed by average \pm standard deviation of the groups. The normality analysis was performed using the D'Agostino & Pearson test. ANOVA test using Dunnett's post-test for multiple comparisons was used to compare the results between groups. Statistical analyses were carried out using GraphPad Prism software (version 6.0 for Windows). Differences were considered significant when $p < 0.05$.

Comparisons for *in vivo* assay were carried out by software R using ANOVA test and Scott Knot test which is a hierarchical clustering algorithm. Differences were considered significant when $p < 0.01$.

Results

In order to identify phage displayed peptides that could bind to midgut proteins of *S. frugiperda* a constrained 7-mer phage library was used for positive panning on immobilized total proteins. The enrichment of recovered phage after each biopanning cycle was determined by ELISA assay (Fig 1). Our data indicated a successful affinity selection of phage clones that recognized the midgut proteins of caterpillar. However, in the last cycle, there was a decrease in absorbance, characterizing a decrease in the enrichment process. Therefore, the phages obtained in the second cycle were used in the subsequent analyses.

Fig 1. Enrichment profile of the phage displayed library during the biopanning against midgut proteins of *S. frugiperda*. Plate was coated with 10µg of midgut proteins and ELISA was conducted to evaluate the enrichment of phages in each of the three rounds. Absorbance of coated wells was subtracted from uncoated wells.

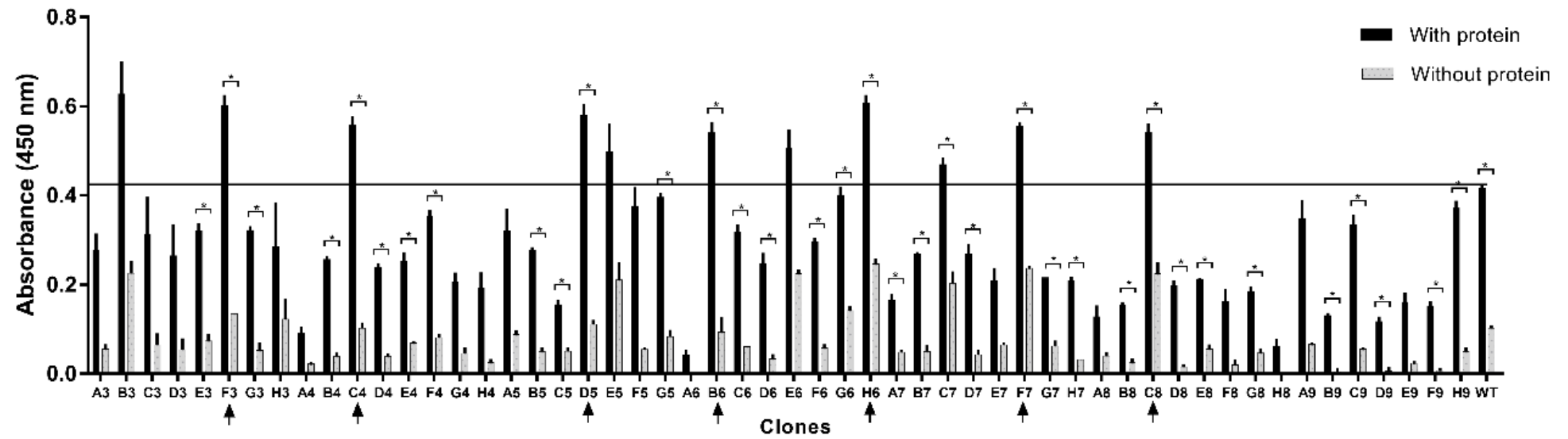


Fifty-four binders were randomly selected after the second cycle of biopanning. Fig 2 shows the reactivity of these clones against caterpillar proteins. The absorbance observed for 35 clones differed significantly from that obtained for uncoated wells ($p < 0.01$). Only the reactivity of the seven clones SfF3, SfC4, SfD5, SfB6, SfH6, SfF7 and SfC8 differed from the wild-type phage (displaying no peptide on its surface). Although SfC7 clone differed significantly from uncoated wells, the absorbance was not significant different from wild-type.

Fig 2. Phage-ELISA for the clones eluted after the second selection cycle. The threshold was based on the absorbance of the Wild-type phage (WT). Selected phages with absorbance greater than WT and significant different from uncoated wells were selected. Of the 54 clones, 35 reacted significantly ($*p < 0.01$) to midgut proteins when compared to uncoated wells. Of these, seven (SfF3, SfC4, SfD5, SfB6, SfH6, SfF7 and SfC8 – indicated by arrows) showed higher absorbance than WT ($p < 0.05$). Data are presented as mean \pm SD D'Agostino & Pearson test was accomplished to compare coated and uncoated wells. ANOVA test using

212 Dunnett's post-test for multiple comparisons was used to compare the results between the 54
213 clones and WT.

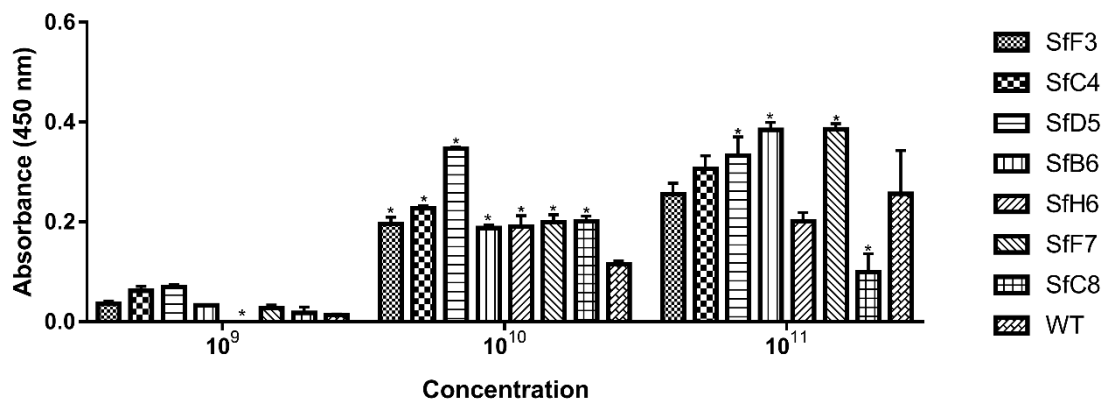
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Clones highlighted in the previous assay were purified and a new format-ELISA was conducted to evaluate the differential reactivity of these promising phages to the midgut proteins of the cartridge caterpillar. Phage clones were using in different titration. As shown in Fig 3, the promising clones only when titrated at 10^{10} and 10^{11} differentially reacted to *S. frugiperda* gut proteins comparing to the wild-type. At 10^{10} , all seven phages differed from the wild-type phage. At 10^{11} , only the SfD5, SfB6 and SfF7 clones showed higher absorbance than the wild-type. Although three clones demonstrated better reactivity, the all seven were evaluated in the further experiments because the selection was performed using the midgut proteins in their non-native conformation (coated in 96-well Maxisorp microtiter plates), and immunohistochemistry and *in vivo* assays included native proteins. Moreover, the conformational library used in this work may allow the selection of molecules with different structures, that may interact with midgut proteins in their native conformation.

Fig 3. Reactivity of purified phage clones against gut proteins from *S. frugiperda*.






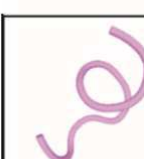

Only seven clones SfF3, SfC4, SfD5, SfB6, SfH6, SfF7 and SfC8 were used based on previous screening assay. Three different phage titers were tested and statistical differences (through ANOVA test using Dunnett's post-test for multiple comparisons) between the selected clones and the wild-type M13 phage (WT) control were observed (* $p < 0.05$).



The sequencing results allowed the characterization of the seven peptides. Alignment analyses were performed using the peptide sequences and no consensus motifs were determined. Result already expected since we use a conformational library. The potential ligands to gut proteins from *S. frugiperda* were predicted through Blastp tool and Fig 4 represents the peptides sequences of the five best-ranked proteins. Using a conformational library is allow selected peptides with different structure, thus were predicted the 3D structure of each peptide predict by PEPFOLD3 – RPBS software.

Fig 4. Exogenous peptides expressed by phage particles. Peptides sequences of the seven phage clones (SfF3, SfC4, SfD5, SfB6, SfH6, SfF7 and SfC8) and their 3D structures were obtained. The potential ligands to midgut proteins of *S. frugiperda* were predicted through BLASTp alignment and the five best-ranked proteins are represented. Conformational library

for biopanning was chosen to allow interaction between the 3D-structure of the peptides and the midgut proteins.

A	Peptide SfF3 - NQSGSSV <table border="1"> <thead> <tr> <th>Aligned proteins</th><th>Cover</th><th>Identity</th></tr> </thead> <tbody> <tr> <td>cytochrome oxidase subunit I [Spodoptera frugiperda]</td><td>85%</td><td>89%</td></tr> <tr> <td>cytochrome oxidase subunit I [Spodoptera frugiperda]</td><td>85%</td><td>89%</td></tr> <tr> <td>cytochrome P450 CYP32B1 [Spodoptera frugiperda]</td><td>100%</td><td>77%</td></tr> <tr> <td>truncated ABC transporter subfamily C2 [Spodoptera frugiperda]</td><td>57%</td><td>100%</td></tr> <tr> <td>hem1 RBR2 ABC2 transporter [Spodoptera frugiperda]</td><td>57%</td><td>100%</td></tr> </tbody> </table>	Aligned proteins	Cover	Identity	cytochrome oxidase subunit I [Spodoptera frugiperda]	85%	89%	cytochrome oxidase subunit I [Spodoptera frugiperda]	85%	89%	cytochrome P450 CYP32B1 [Spodoptera frugiperda]	100%	77%	truncated ABC transporter subfamily C2 [Spodoptera frugiperda]	57%	100%	hem1 RBR2 ABC2 transporter [Spodoptera frugiperda]	57%	100%	
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G	Peptide SfC8 - PSAFHTG <table border="1"> <thead> <tr> <th>Aligned proteins</th><th>Cover</th><th>Identity</th></tr> </thead> <tbody> <tr> <td>Reckless: Full-40S ribosomal protein S8</td><td>57%</td><td>100%</td></tr> <tr> <td>chymotrypsin-like serine protease precursor [Spodoptera frugiperda]</td><td>57%</td><td>100%</td></tr> <tr> <td>trypsin-like serine protease precursor [Spodoptera frugiperda]</td><td>57%</td><td>100%</td></tr> <tr> <td>scavenger receptor SR-C-like protein [Spodoptera frugiperda]</td><td>85%</td><td>35%</td></tr> <tr> <td>scavenger receptor SR-C-like protein [Spodoptera frugiperda]</td><td>85%</td><td>35%</td></tr> </tbody> </table>	Aligned proteins	Cover	Identity	Reckless: Full-40S ribosomal protein S8	57%	100%	chymotrypsin-like serine protease precursor [Spodoptera frugiperda]	57%	100%	trypsin-like serine protease precursor [Spodoptera frugiperda]	57%	100%	scavenger receptor SR-C-like protein [Spodoptera frugiperda]	85%	35%	scavenger receptor SR-C-like protein [Spodoptera frugiperda]	85%	35%	
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Considering the protein that aligned with the selected peptides, all presented similarity to *S. frugiperda*. A literature search was performed (Table 1) to identify possible molecules that interact with each BLASTp-aligned protein. Only the SfF7 peptide did not present a target described for the genus *Spodoptera*.

The results demonstrated that both, SfF3 and SfF5, presented similarity to cytochrome oxidase. Interestingly, SfF3, SfC4 and SfD5 were similar to cytochrome P450, a detoxification enzyme regulated by endogenous miRNAs [23]. SfF3 also aligned with ABCC2 transporter, a receptor of Cry1A toxins [24], and SfC4 and SfF7 also targeted miRNAs, which are also involved in resistance mechanisms [23].

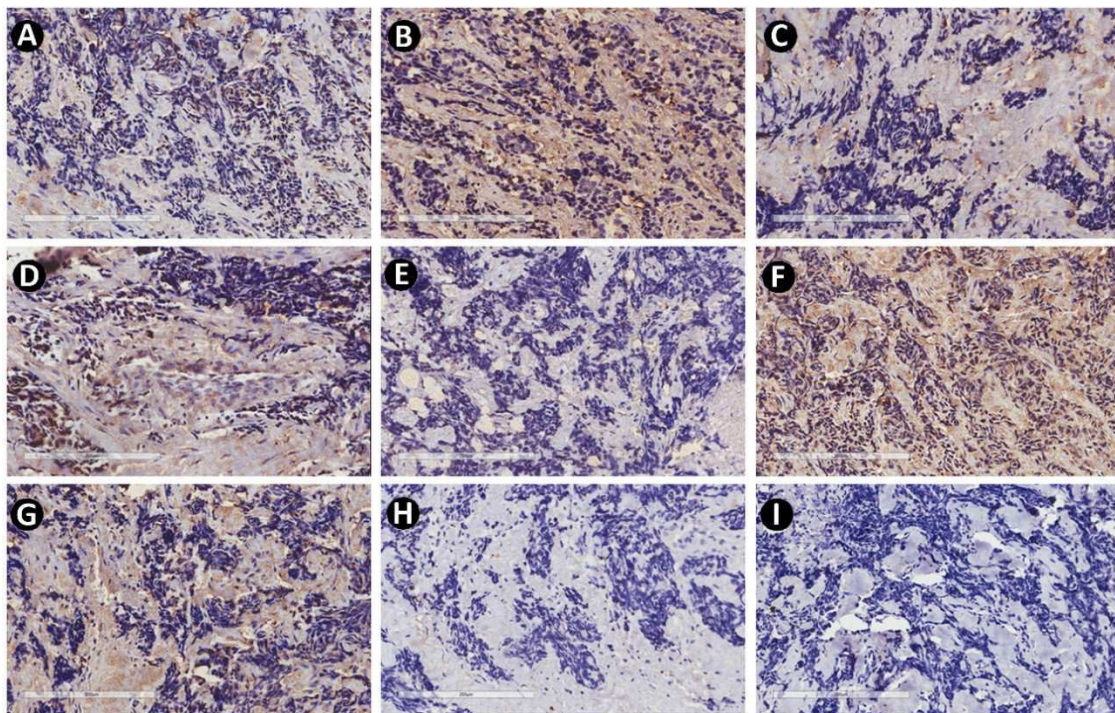
Table 1. Similarities between the peptide sequences and *S. frugiperda* proteins registered in the GenBank database. The target in the same genus of this caterpillar was recorded, and studies in different insects were reviewed.

Peptides	Potential ligands to gut proteins	Described targets	Organisms	References
SfF3	cytochrome oxidase subunit I [Spodoptera frugiperda]	Other subunits	<i>Omphisa fuscidentalis</i>	[25]
	cytochrome oxidase subunit I [Spodoptera frugiperda]	Insecticides	<i>Spodoptera litura</i>	[26]
	cytochrome P450 CYP321B1 [Spodoptera frugiperda]	Cry protein receptor	<i>Spodoptera frugiperda</i>	[27] and [28]
	truncated ABC transporter subfamily C2 [Spodoptera frugiperda]			
	fam1 R2R2 ABCC2 transporter [Spodoptera frugiperda]			
SfC4	PRO [Spodoptera frugiperda]	-	-	-
	prophenoloxidase subunit 2 [Spodoptera frugiperda]	Phenolic substances	<i>Bombyx mori</i>	[29]
	chitin synthase [Spodoptera frugiperda]	Sugar	Insect	[30]
	cytochrome P450 CY321A8 [Spodoptera frugiperda]	Insecticides	<i>Spodoptera litura</i>	[26]
	Dicer-1-PA [Spodoptera frugiperda]	miRNAs	<i>Drosophila</i>	[31]
SfD5	cytochrome P450 CYP6AN4 [Spodoptera frugiperda]	Insecticides	<i>Spodoptera litura</i>	[26]
	acid digestive lipase [Spodoptera frugiperda]	Lipids	<i>Drosophila melanogaster</i>	[32]
	cytochrome c oxidase subunit 1 [Spodoptera frugiperda]	Oxygen		
	cytochrome c oxidase subunit I [Spodoptera frugiperda]	Oxygen	<i>Omphisa fuscidentalis</i>	[25]
	cytochrome c oxidase subunit 1 [Spodoptera frugiperda]	Oxygen		
SfB6	RecName: Full=60S ribosomal protein L32	rRNA / Polypeptides	<i>Spodoptera frugiperda</i>	[33]
	ribosomal protein L12 [Spodoptera frugiperda]	rRNA / Polypeptides		
	charged multivesicular body protein 2b [Spodoptera frugiperda]	Polymers	<i>Homo sapiens</i>	[34]
	charged multivesicular body protein 5 [Spodoptera frugiperda]	Polymers		
	actin related protein 2/3 complex subunit 2 [Spodoptera frugiperda]	Actin binding proteins	<i>Drosophila</i>	[35]
SfH6	RecName: Full=40S ribosomal protein S4	rRNA / Polypeptides	<i>Spodoptera frugiperda</i>	[33]
	prophenoloxidase subunit 1 [Spodoptera frugiperda]	Phenolic substances	<i>Bombyx mori</i>	[29]
	ORF 7 [Spodoptera frugiperda]	-	-	-
	fam1 SS ABCC2 transporter [Spodoptera frugiperda]	Cry protein receptor	<i>Spodoptera frugiperda</i>	[27,28]
	ABC transporter subfamily C2 [Spodoptera frugiperda]			
SfF7	putative CAD trifunctional protein [Spodoptera frugiperda]	Nucleotides	<i>Drosophila melanogaster</i>	[36]
	Dicer-1-PA [Spodoptera frugiperda]	miRNAs	<i>Drosophila</i>	[31]
	Chain A, Mechanisms of Crippling Actin-dependent Phagocytosis By Yopo	-	-	-
	polyadenylate binding protein [Spodoptera frugiperda]	Nucleotides	<i>Drosophila</i>	[37]
	CRM1 [Spodoptera frugiperda]	rRNA / Protein	Mammal	[38]
SfC8	RecName: Full=40S ribosomal protein S8	rRNA / Polypeptides	<i>Spodoptera frugiperda</i>	[33]
	chymotrypsin-like serine protease precursor [Spodoptera frugiperda]	Proteins	<i>Anticarsia gemmatilis</i>	[39]
	trypsin-like serine protease precursor [Spodoptera frugiperda]			
	scavenger receptor SR-C-like protein [Spodoptera frugiperda]	Vip protein	<i>Spodoptera frugiperda</i>	[40]
	scavenger receptor SR-C-like protein [Spodoptera frugiperda]			

The targets are described in agreements with published data.

In order to map the binding of the phages in the midgut tissue of the caterpillar, an immunohistochemistry analysis was performed (Fig 5). The reactivity of the seven clones was observed preferentially in the cytoplasm (Fig 5A-G). As negative control, non-recombinant wild-type phage (Fig 5H) were used, showing poor marking to the gut of the caterpillar. Control of the reaction, in which the virus was suppressed, did not show immunostaining (Fig 5I).

Fig 5. Immunohistochemical analysis of the midgut tissue of *S. frugiperda* (20X) demonstrating the labeling of the selected peptides. A: SfF3; B: SfC4; C: SfD5; D: SfB6; E: SfH6; F: SfF7; G: C8; H: wild-phage and I: negative control without phage-clones. Counterstaining with Harris-Hematoxylin.



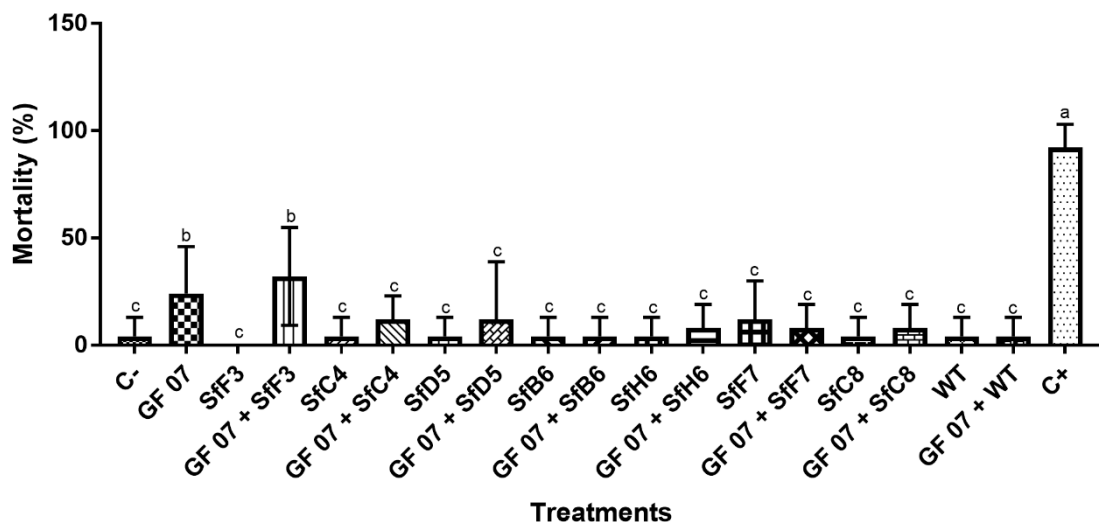
Finally, we carried out the *in vivo* assay to verify the mortality of the newborn fall armyworm fed with the selected seven phages (SfF3, SfC4, SfD5, SfB6, SfH6, SfF7 and SfC8) and the wild type. We also performed the combination of the strain GF 07 with the peptides aiming to observe a synergistic action between both. The results were compared

to the negative control (Fig 6). This consisted of solution without phage or Bt toxins, demonstrating, as expected, low mortality (4%). We used, as a positive control, the Agree® which contains *B. thuringiensis* aizawai transconjugated with *B. thuringiensis* kurstaki. This product has already been tested and was efficient in controlling *S. frugiperda* [41,42], *Plutella xylostella* [43], *Bonagota salubricola* [44] and *Duponchelia fovealis* [45]. In our experiments the mortality of Agree® was 92%.

In this work, we also used *strain* of *B. thuringiensis* GF 07 combined with selected peptides to increase the application spectrum of biopesticides. This combination is interesting due to resistance reports from different pests to bacteria *B. thuringiensis* [46]. In addition, studies indicated that Bt toxins are toxic to caterpillars from 1st to 4th instar, failing to act in the late stages, when damages are more severe [47,48].

Fig 6. *In vivo* assay using the droplet feeding methodology in *S. frugiperda* larvae.

The fall armyworms were fed by the phages SfF3, SfC4, SfD5, SfB6, SfH6, SfF7, SfC8 and wild-type (WT). We also combined these phages with the *B. thuringiensis* strain GF 07 to observe the synergistic effect. As a positive control (C+) we used the bioinsecticide Agree®. All treatments were compared with the negative control (C-) using Scott-Knott test and statistical difference was considered when $p < 0.01$. Treatments followed by the same letter are not statistically different at the 0.01 level as determined by Scott-Knott test.



The analysis of variance using the Scott-Knott's test allowed treatments grouping and consists of partitioning the original set of treatments. This partitioning aims at a maximum differentiation between groups and is interrupted when the groups obtained are no longer significantly different [49]. This process is quite interesting when the number of treatments is large, such as the assay performed in this work.

The treatment with GF 07, GF 07+SfF3 and the positive control were statistically different from the negative control. The treatments lead to a low mortality, except for the positive control (92%). Strain GF 07 resulted in a significantly higher mortality (24%)

comparing to the negative control. Although no phage was capable of killing *S. frugiperda* larvae alone, when associated to GF 07, the GF 07+SfF3 complex caused significantly mortality (32%) compared to the negative control. Thus, by combining this peptide and the GF 07 we achieved an 8% increase in mortality of *S. frugiperda* to the strain GF 07.

Discussion

This is the first study that aimed to use the PhD in searching a new strategy for fall armyworm control. This pest control is economic interest and necessary, since the transgenic plants no longer proves to be effective. For this purpose, the midgut proteins of *S. frugiperda* were extracted for the selection of peptides fused to bacteriophages capsids. The use of peptide libraries expressed on the surface of filamentous phage is a robust *in vitro* selection tool and has been very successful in identifying ligands in different systems [50]. Our strategy, therefore, is innovative when exploring a biotechnological tool that allows selecting molecules with insecticidal activity.

During the biopanning process, after three cycles, there was a reduction in enrichment. In fact, phages may evolve during the process due to their higher infectivity, in detriment of their specificity. The biological bias during this selection functions as an important driving force for the isolation of irrelevant peptides, leading some to be enriched or removed during biopanning. Consequently, these peptides show a higher than expected frequency within the library [51]. By performing fewer rounds of selection, the bias can be reduced by decreasing the loss of relevant sequences [52]. Therefore, the second round was selected for the isolation of phage clones, which were subsequently selective against midgut proteins.

Based on the objectives of this work, the ELISA test was crucial for the validation of the selection process, as well as for the identification of promising clones. This

technique demonstrated the affinity to the target and not only the numbers of viral particles. Ngubane et al. [53] used the same validation technique and achieved a successful enrichment, allowing the identification of specific peptides for mycobacteria. This immunoassay was also previously performed to demonstrate the reactive phage particles to the desired target by [54,55]. Considering our results, the process was successful, since we selected seven clones reactive to the target, compared to the wild-type, a filamentous virus that does not express external peptide fused to protein III. At the end of the selection process, using purified phage, we obtained seven promising clones capable of specifically recognizing midgut proteins of *S. frugiperda*. This reactivity was further confirmed by the immunohistochemistry assay, in which the phage clones recognized proteins in the cytoplasm of intestinal cells.

Throughout the bioinformatics analysis, no protein structural motif was identified during the alignment of the sequences of the seven clones. This result was expected because we used a conformational library for biopanning process. This library is particularly interesting because allows the selection of peptides with different conformations, which can interact with different midgut proteins.

In the *in vivo* assays, we used the combination of phages with the *B. thuringiensis*, a gram-positive bacterium that during sporulation produces lethal endotoxins for different larvae species. These toxins are synthesized in the form of inactive pro-toxins that are solubilized and proteolytically activated in the insect gut. After binding to its receptor, the toxin leads to swelling of the midgut cells and eventual lysis [7,8]. Therefore, the combination of the clone with these bacteria can promote a synergism leading to improved mortality.

With exception of SfF3 clone, no phages were able to promote *S. frugiperda* larvae death, even in combination with *B. thuringiensis* GF 07. Even not toxic, they may

be used as delivery vehicles in a new method design based on biotechnology approaches, once they are ligands of intestinal proteins. Besides, these six peptides may still contribute to fall armyworm control in more advanced instars. Some studies suggest that products based on Bt technology only act on larvae of up to 4th instar [47,48], making them advantageous to be tested as insecticides that act in the other phases

The SfF3 clone, although in isolation has not caused the death of *S. frugiperda* larvae, increased the mortality to the strain GF 07. We predicted similarity of the expressed peptide to the truncated ABC transporter subfamily C2 (ABCC2) by performing alignment through BLASTp tool. Recent studies have shown that mutations in ABCC2 protein confer resistance against Bt toxins in *Bombyx mori* [56], *Helicoverpa armigera* [57], *Spodoptera exigua* [58], *Plutella xylostella* [59] and *S. frugiperda* [27,60]. Researchers suggest that the proteins of the Cry1 family interact with ABCC2 to initiate the formation of pores in the intestinal membrane of the insect and when different mutations render the ABCC2 the protein becomes nonfunctional, leading to resistance [61,62]. ABCC2 sequences possess an aberrant splicing and its function is still unknown [28]. We suggest in this work that the peptide expressed by the SfF3 clone can interact with the Bt toxin and this small sequence of amino acids can reverse the resistant phenotypes, since this peptide showed interaction with the Cry protein receptor. However, experiments with different individuals are needed to test this notion.

Sf3F was also similar to cytochrome P450, which is an enzyme involved in the detoxification of chemical insecticides [63,64] and on pest resistance [65,66]. Recent studies have demonstrated that Bt toxin is capable of inducing the expression of cytochrome P450 in *Manduca sexta* larvae [67]. Furthermore, this enzyme is involved in the resistance of *Spodoptera exigua* to beta-cypermethrin [68]. Interestingly, some

researchers have also reported that microRNAs (miRNAs) can regulate the expression of this enzyme, suggesting new pathways associated to insects' resistance [69-71].

When exposing larvae of *Plutella xylostella* to the insecticide chlorantraniliprole, Etebari et al. [72] found that miR-2b-3p was differentially expressed. This miRNA was able to regulate the transcriptional levels of cytochrome P450, thus playing a role in metabolic resistance. Consequently, the synergistic effect observed when combining SfF3 and strain GF 07 might have occurred due to the modulation of cytochrome P450 through miRNA induction, which need be further evaluated

In summary, this is the first work that used the PhD technology to select clones for *S. frugiperda* control. Seven clones were successful selected and bound to intestinal proteins. Among then, the SfF3 when used in combination with the *B. thuringiensis* toxin, increased the mortality of *S. frugiperda* neonates. However, new trials with fall armyworm in the last instars should be performed, and resistant individuals should be included to evaluate the role of these peptides in reversing pesticide resistance. This study also validated the reliability of PhD technology as an agribiotechnological approaches for pest control.

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CAPÍTULO III

IDENTIFICATION OF miRNAS IN *Spodoptera frugiperda*

Instructions according to **Journal of invertebrate pathology**

Title: Identification of miRNAs in *Spodoptera frugiperda***Autorship**

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ABSTRACT

Spodoptera frugiperda is one of the main agricultural pests causing severe productivity losses. A promising alternative to this pest control is miRNA mechanism, which regulates gene expression and protein translation. In the present study we aimed to identify and characterize, by *in silico* analysis, the miRNAs in the genome of *S. frugiperda*. Using robust software with optimized algorithm, we were able to predict 350 miRNAs precursors, and 60 miRNAs were specific for genus *Spodoptera*. Subsequently, we performed the refinement of 10 RNA-seq and we identified 91 miRNAs not yet described, demonstrating the possibilities of this sequences in describing new important regulatory mechanisms in crop protection. Our results expand the knowledge about the miRNAs in the genome of *S. frugiperda* and bring new possibilities to understand their roles in the biology of this important pest.

Keywords: Fall armyworm, miRNA, bioinformatics, bioinsecticide

1 INTRODUCTION

Fall armyworm (*Spodoptera frugiperda*, Lepidoptera: Noctuidae) is one of the main agricultural pests presents in the tropical and subtropical regions of the American continent (Bateman et al., 2018). The larvae are polyphagous but well adapted to feed on grasses; causing severe damage to economically important crops such as corn, rice, sorghum and sugarcane. In addition, losses are also accounted in other crops such as cabbage, beet, peanut, soybean, alfalfa, onion, cotton, grass, millet, tomato, potato and cotton (Luginbill, 1928). This insect can potentially feed on over 100 species of plants from a wide range of families (CABI, 2018). In Brazil, it is considered the most important corn pest, causing severe productivity losses (Cruz and Turpin, 1983; Sarmiento et al., 2002). Recent studies show yield losses in a range of 21% to 53% of annual average maize production, estimated at \$ 2,481 million - \$ 6,187 million (Abrahams et al., 2017).

In corn, young larvae hide in the cartridge during the day, but leave at night to feed on developing leaves. Older larvae remain inside the cartridge, and reduce product quality by compromising plant reproductive structures (Cruz, 1995). This behavior makes the pest control difficult, especially when using chemical products that depend on direct contact with the larva, which is protected (Bateman et al., 2018). In addition, these

pesticides cause severe health and environment damages (FAO and WHO, 2016). In this context, alternative strategies of combat are particularly interesting.

The use of Cry toxins from *Bacillus thuringiensis* expressed in plants brought significant advances in *S. frugiperda* control. However, resistant insects have already emerged, highlighting the importance of new methods in agriculture field (Flagel et al., 2018). An increasingly exciting option is the use of interference RNA (RNAi) technology, in which microRNAs (miRNAs) or small interfering RNAs (siRNAs) trigger a reduction of transcripts that are complementary to their sequences (Kim et al., 2009; Swevers et al., 2018). They are prominent by their specificity (Bachman et al., 2013; Chen et al., 2018a) and disruption of miRNA genes may cause serious physiological consequences, including developmental defects in invertebrates (Bartel, 2018; Li et al., 2018).

Firstly, described in *Caenorhabditis elegans* (Lee et al., 1993) miRNAs compromise protein expression through inhibition of translation, degradation or decay of mRNA using Argonaute (Ago)/RNA-induced silencing complex (RISC) mechanism (Fu et al., 2017). Insects express different miRNAs during development and have evolved strategies to overcome plants defenses against their attack modulating small non-coding RNAs (Li et al., 2018). It is therefore reasonable to assume that changes in endogenous miRNAs of the insect may also affect its biological functions, and survival.

Bioinformatic tools for prediction of miRNAs have been widely and successfully used (Chen et al., 2018b; Yang et al., 2018). Considering *S. frugiperda*, its genome was recently sequenced (Nandakumar et al., 2017). However, a greater understanding and deeper annotation of its sequences, including the identification of miRNAs, is still incipient and essential as a strategy for this pest control.

The objective of this study was, therefore, to identify and characterize, by *in silico* analysis, the miRNAs present in the genome of *S. frugiperda*. The understanding of these sequences will certainly allow their later use as a tool to control the carcass caterpillar when being included in artificial diets or templates for genetically modified plants.

2 MATERIALS AND METHODS

2.1 PREDICTION OF MATURE miRNAS AND THEIR PRECURSORS (pre-miRNAS)

The most recent data and information on the genome of *S. frugiperda* were accessed and downloaded from the National Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/>). The search for probable miRNAs and their precursors was carried out using software developed in several stages based on conserved characteristics of these molecules (de Souza Gomes et al., 2011). First, sequences of the genome of *S. frugiperda* with potential hairpin formation or similar miRNA precursor structures were obtained. The BLASTn and EINVERTED programs (EMBOSS tool) were used. The undesirable sequences were removed, selecting only those corresponding to probable miRNAs. The filters for removal were based on conserved characteristics of miRNA precursors as well as characteristics of other known regions which have no potential for formation of miRNA precursors. The filters used were: GC content (guanine and cytosine), minimum free energy (MFE), homology with mature miRNAs, homology with protein coding regions, homology with repetitive regions and homology with non-coding RNAs.

We also used a miRNA classifier based on previous features of possible real miRNAs, pseudo-miRNAs and non-real miRNAs, as well as information extracted from machine learning technology "Random forest" (Jiang et al., 2007). For the search for non-conserved miRNAs, an extra step was added to the protocol to search for mature sequences of miRNAs in the probable secondary structures of their respective precursors (Gkirtzou et al., 2010). Prediction of the secondary structure of pre-miRNAs was performed using the RNAfold (Vienna RNA Package) with adjusted parameters (Zuker and Stiegler, 1981).

Additional analyses were performed with the objective of comparing the putative miRNAs with the set of miRNAs found in other organisms. These analyzes involved the study of some previously used parameters and other additional parameters such as: Adjusted Minimum Free Energy (AMFE), Minimum Free Energy Index (MFEI), GC content, Minimal Free Energy of the thermodynamic Ensemble (MFEE), Ensemble Diversity and frequency of the MFE structure in the ensemble.

2.2 IDENTIFICATION OF PROTOSTOME-SPECIFIC miRNAS

To identify the protostome-specific miRNAs, the previously identified precursor sequences were aligned using the BLASTn tool and the miRBase database (Altschul et al., 1990; Kozomara and Griffiths-Jones, 2013).

2.3 SEQUENCING ANALYSIS OF THE RNA-SEQ LIBRARIES

We used 17 sequencing (SRR6670043, SRR6670042, SRR6670041, SRR6670040, SRR1536060, SRR1536059, SRR1536058, SRR850249, SRR850248, SRR850247, SRR850246, SRR850245, SRR850244, SRR850243, SRR850242, SRR496499, SRR496500) and the quality of the RNA-seq libraries was evaluated using the FastQC software. The adapters were removed with Trimmomatic, discarding reads with a score below 20 and a length of less than 17 and greater than 30 nucleotides.

The filtered sequences were mapped and quantified using miRDeep. The miRDeep2 and Perl scripts were used in each sequence separately to generate the number of reads for each identified miRNA. The R program was used to merge the counts of the different sequencing.

3 RESULTS

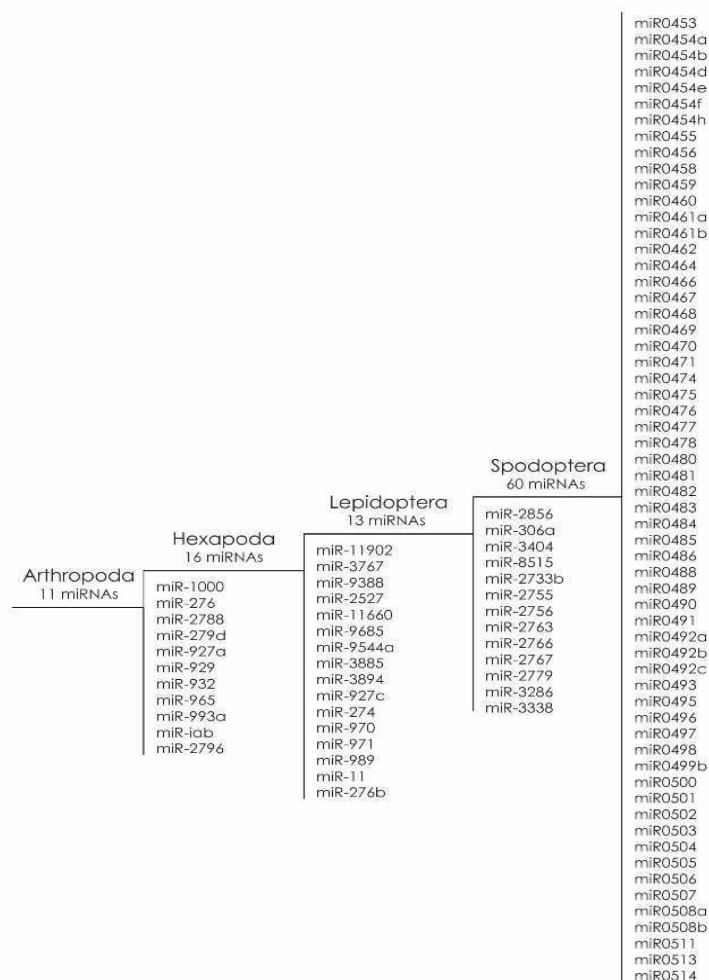
Once miRNAs regulation of gene expression and protein translation is a critical event in insect's physiology, we predicted the miRNAs and their precursors in the genome of *S. frugipeda*. We believe that these sequences may be biotechnologically exploited feeding the larvae with miRNA mimics added to the diet or even expressed by transgenic plants (Peng et al., 2016).

When applying an optimized algorithm, we identified 350 precursor miRNAs and their structural and thermodynamic characteristics are described in Table S1. The MFE of the pre-miRNAs ranged from -18.50 to -86.40 kcal.mol⁻¹, with a mean of -34.97 kcal.mol⁻¹; an average AMFE of -38.78 kcal.mol⁻¹ and MFEI -0.87, similar to the values reported by other studies (Joshi, 2018; Nageshbabu et al., 2012; Sahoo et al., 2018; Sunkar and Jagadeeswaran, 2008; Zhang et al., 2006a; Zhang et al., 2006b; Zhang et al., 2008). The MFE of the precursor miRNAs is generally below -18 kcal.mol⁻¹ and can be distinguished from other classes of RNAs by presenting values of MFEI less than 0.85 (Joshi, 2018; Nageshbabu et al., 2012; Zhang et al., 2006a; Zhang et al., 2006b). In addition, RNA sequences with an AMFE close to -46 kcal.mol⁻¹ are considered likely to

be true miRNAs, compared to the less negative values typical of other classes of RNAs (Zhang et al., 2006a; Zhang et al., 2006b). Our results showed an average AMFE of -38.78 kcal.mol⁻¹, contributing to conclude that the predicted molecules are probable sequences of genuine miRNAs.

A crucial consideration in RNAi technology is specificity for the target specie. No lethally is desired for unrelated insects such as fruit flies (*Drosophila* sp.) or pea aphids (*A. pisum*). To overcome this issue, we performed a phylogenetic distribution of the 350 molecules initially predicted between four different clades defining the protostome-specific sequences (Figure 1). We identified 100 specific molecules to the clade Arthropoda, 89 to *Hexapoda*, 73 to *Lepdoptera* and 60 specifics to genus *Spodoptera*. In this way, the miRNA present in the same genus of the carcass caterpillar were categorized, suggesting its possible application in the specific control of this pest.

Figure 1 Phylogenetic distribution of the miRNAs identified in the genome of *Spodoptera frugiperda*. Of the 100 molecules specific to Arthropoda, 89 were specific to Hexapoda, 73 to Lepidoptera and 60 to genus *Spodoptera*.



The next step was to analyze dataset of publicly available total RNAseq. Seventeen different sequencing of *S. frugiperda* were used (Table S2). Considering the strategy to pest control through the expression of miRNAs mimics, we selected 116 molecules based on the total of counts found in at least 10 sequencing. As such molecules are present in several different sequencing the probability of being real are greater. Of these 116 selected miRNAs, only 25 have already been studied and their biological effects have been described in the literature for other species (Table 1), including associated with mechanisms of resistance to insecticides. The remaining 91 sequences need to be explored and their role must be elucidated in *S. frugiperda*.

Table 1 miRNAs found in the genome of *S. frugiperda* with functions already described for other species. The prediction was performed through an algorithm developed by de Souza Gomes et al. (2011) using 17 RNA-seq libraries.

miRNA	Biological effect	Organism	References
miR-2765	Caste differentiation	<i>Reticulitermes speratus</i>	(Matsunami et al., 2018)
miR-285	Interacts with P450 to regulate resistance to pyrethroids	<i>Culex pipiens pallens</i>	(Tian et al., 2016)
miR-33	Gonadogenesis	<i>Drosophila</i>	(Yang et al., 2016a)
miR-279	Immune pathway regulator	<i>Drosophila melanogaster</i>	(Fullaondo and Lee, 2012)
miR-7	Immunity	<i>Plutella xylostella</i>	(Xu et al., 2017)
miR-932	Mechanism of resistance to insecticides	<i>Culex pipiens pallens</i>	(Liu et al., 2016)
miR-276	Synchronized hatching	<i>Locusta migratoria</i>	(He et al., 2016)
miR-34	Regulates innate immunity and ecdysone signaling	<i>Drosophila</i>	(Xiong et al., 2016)
miR-79	Regulates innate immunity	<i>Rhipicephalus haemaphysaloides</i>	(Wang et al., 2015)
miR-92	Inhibits premature differentiation of neuroblasts	<i>Drosophila</i>	(Yuva-Aydemir et al., 2015)
miR-9	Prevents apoptosis during wing development	<i>Drosophila</i>	(Bejarano et al., 2010)
miR-13	Apoptosis control	<i>Drosophila</i>	(Stark et al., 2003)
miR-1	Muscle development	<i>Drosophila</i>	(Zhu et al., 2017)
miR-308	Regulates immunity	<i>Plutella xylostella</i>	(Shakeel et al., 2018)
miR-745	Immunity	<i>Plutella xylostella</i>	(Xu et al., 2017)
miR-2	Immunity	<i>Plutella xylostella</i>	(Xu et al., 2017)
miR-278	Regulate resistance to pyrethroids	<i>Culex pipiens pallens</i>	(Lei et al., 2015)
miR-14	Regulation of autophagy	<i>Drosophila</i>	(Nelson et al., 2014)
miR-317	Embryogenesis	<i>Drosophila</i>	(Pushpavalli et al., 2014)
miR-10	Development	<i>Drosophila</i>	(Stark et al., 2007)
miR-71	Chitin metabolism	<i>Locusta migratoria</i>	(Yang et al., 2016b)
miR-277	Lipid metabolism	<i>Aedes aegypti</i>	(Ling et al., 2017)
miR-306	Regulates immunity	<i>Plutella xylostella</i>	(Shakeel et al., 2018)
miR-11	Apoptosis control	<i>Drosophila</i>	(Ge et al., 2012)
miR-263	Chitin metabolism	<i>Locusta migratoria</i>	(Yang et al., 2016b)

4 DISCUSSION

miRNAs are important non-coding regulators of physiological events in insects (Gordon and Waterhouse, 2007). Disrupt their expression thorough miRNA mimics may be an excited application in agriculture, controlling pest, decreasing pesticide resistance, and preventing non-target species death (Saini et al., 2018; Swevers et al., 2018). Actually, this is a promising technology that must be explored, especially considering the management of *S. frugiperda*.

S. frugiperda is one of the main agricultural pests and the climatic conditions are determinant for its occurrence (Bavaresco et al., 2002). This species can reduce maize production in up to 53% and frequent applications of insecticides are necessary to reduce the population avoiding economic damages (Abrahams et al., 2017). Despite the genome already described, the notation of its miRNAs is still restricted and describing them is necessary to overcome the challenges faced in plantations. In the present study, the prediction of miRNA precursors from *S. frugiperda* genome allowed the identification of 350 different molecules with determination of their thermodynamic and structural characteristics. The thermodynamic aspects are useful criteria to differentiate miRNAs from other types of RNAs (Zhang et al., 2006b), already the structural characteristics help in these sequences validation as interfering RNAs (de Sousa Cardoso et al., 2016). Considering the thermodynamic parameters obtained in this work we can affirm that the miRNAs identified *in silico* from the genome of *S. frugiperda* are genuine molecules. Thus, it is possible to use them as modulators of the expression of different genes and proteins.

For the design of the molecular interference strategy, special attention should be devoted to off-targets (Birmingham et al., 2006; Jackson et al., 2003; Ljepoja et al., 2018). Considering pest control, the miRNA must be species-specific (Lim et al., 2016) with minimal effect on beneficial insects to the crop. Through the phylogenetic distribution of the miRNAs it was possible to identify 60 molecules specific to genus *Spodoptera*.

Some miRNA molecules are naturally suppressed in healthy organisms. Exogenous administration and, consequently, increased expression may lead to physiological modifications ranging from growth retardation, making the organism more susceptible to control until its death. We initially identified 116 miRNA molecules present in 10 different RNA-seq of *S. frugiperda*. Of these, only 25 have already been studied in other organisms and have their biological effect described. The miR-276, for example, despite the small number of counts, is responsible for the synchrony of progeny hatching in locusts (He et al., 2016) and for the coordination of neural function in *Apis mellifera* (Hori et al., 2011). On the other hand, miR-

932 targets a gene related to resistance to pyrethroids in *Culex pipiens pallens*. Liu et al. (2016) observed that overexpression of this miRNA renders more resistant mosquitoes while inhibition made them more sensitive to deltamethrin. This fact evidences the importance of a careful choice in the design of interfering sequences.

Another approach involving miRNAs may be adopted, including inhibitors of its expression with consequent insect combat. Saini et al. (2018) developed a miRNA capable of silencing the acetylcholinesterase gene of *Helicoverpa armigera*, an essential enzyme for the nervous system. By silencing this enzyme, the researchers observed a delay in larval growth and mortality, as well as the faulty emergence of adults, indicating that this miRNA can be used as an insecticidal molecule. In the present study, we identified 91 miRNAs that lack characterization and, therefore, open new ways in the exploration of these molecules in *S. frugiperda* control.

It is important to notice that, this agrobiotechnology approach for pest control can modify not only the target-gene expression, but also can modulate distinct physiological processes. Therefore, plant-insect interactions must be considered in analysis of the biological effects of the RNAi pathway.

5 CONCLUSION

The identification of miRNAs in *S. frugiperda* genome is an important tool to create alternatives for this pest control. We successfully applied an integrated computational analysis with the prediction of 350 miRNA precursors, being 60 specifics to the study genre. Subsequently, the refinement of 10 RNA-seq showed 91 miRNAs not yet described. Our results demonstrate the possibilities of the sequences described here, which are associated to an attractive regulatory mechanism as biotecnologically feasible in crop protection.

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DECLARATION OF INTEREST

We confirm that all authors fulfill all conditions required for authorship, and have read and approved the manuscript. We also confirm that there is no potential conflict of interest, as described in the Instruction for Authors.

AUTHORS' CONTRIBUTIONS

Experimental design and manuscript writing: TGA e SBRN

Optimization of in silico algorithms: MSG e LRA

In silico analysis: SBRN, TCSC e TCA

Project coordinator and senior author: TGA

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SUPPLEMENTARY TABLES

Table S1 Thermodynamic and structural characterization of *S. frugiperda* pre-miRNAs. The following parameters are described: size, G, A, C and U content, GC and AU content, UA and GC ratio, Minimum Free Energy (MFE), Minimal Free Energy of the thermodynamic Ensemble (MFEE), Free Minimum Thermal set (Freq), Ensemble Diversity (Div), Adjusted Minimum Free Energy (AMFE), Minimum Free Energy Index (MFEI).

Pre-miRNA	Sequence	Size	G	A	C	U	GC	AU	UA-ratio	GC-ratio	MFE	MFEE	Freq	Div	AMFE	MFEI
>sfr-miR-466f-1	CGUUUACAGGAAGUU UGUGCUCaucugUACA CAGACAUACGUGUGU GUGCAUGUGAGCACG AAUACACCGUAAACUA	77,00	23,38	27,27	20,78	27,27	44,16	54,55	1,00	1,13	-35,00	-32,30	0,23	3,19	-45,45	-1,03
>sfr-miR-274	CUUGUUAAGCGAAGU UGGUUUGUGACCGUC ACUAAACGGGCAGUACC UCUAUUUGCUCGUUU UGACGAUCGCAAAAU GAACUCGCUUGACUCU	93,00	22,58	21,51	22,58	32,26	45,16	53,76	1,50	1,00	-41,50	-41,50	0,06	6,09	-44,62	-0,99
>sfr-miR-970	UGUCCGCGAGAGCCU GCGUGUGCUCUUAUU GGUAGUGUGGCUGAU AAUCUAUCAUAAGAC ACACGCGGCUCUCGCU GGCUC	83,00	27,71	16,87	25,30	28,92	53,01	45,78	1,71	1,10	-44,30	-43,40	0,19	4,31	-53,37	-1,01
>sfr-miR-281	UCUGUUAAGAAGAG AGCUAUCCGUCGACAG UAUUGCCAGUAAACAC UGUCAUGGAGUUGCU CUCUUUAUGAACGGU	78,00	23,08	25,64	19,23	30,77	42,31	56,41	1,20	1,20	-34,60	-34,60	0,30	2,44	-44,36	-1,05
>sfr-miR-10486	AGGAUUCAAUAUUU CACUGUCCUGCAGGCG	100,00	22,00	23,00	19,00	35,00	41,00	58,00	1,52	1,16	-20,50	-13,10	0,03	23,9 4	-20,50	-0,50

	CCUUAUCGGCUCUGAU GACAAUUUUGUGAUU AUCUGAUGGCAUGCG GUCUAAUGCAUUUAU GAAUGUC															
>sfr-miR-10459-1	AAUGCUAUACGACAU GAUGCUAUGGAAGUA ACUGAAAUACACUAG UCUUGCCGUGCCUAUG UCGGUUAGCUGU	74,00	22,97	27,03	18,92	29,73	41,89	56,76	1,10	1,21	-23,60	-23,60	0,51	2,11	-31,89	-0,76
>sfr-miR-10459-2	CCCACAGCAAAAUGCC AUAGACAUGAUGCUA UGGAAGUAAACUGAAA UACACUAGUCUUGCCG UGCCUAUGUCGGUUA GCUGUCUG	86,00	22,09	27,91	23,26	25,58	45,35	53,49	0,92	0,95	-20,70	-20,70	0,30	13,9 3	-24,07	-0,53
>sfr-miR-10459-3	CCCACAGCAAAAUGCC AUACGACAUGAUGCU AUGGAAGUAAACUGAA GUAUACUAGUCUUGCC GUGUCUAUGUCAGUU AGCUGUCUG	87,00	21,84	27,59	21,84	27,59	43,68	55,17	1,00	1,00	-20,40	-20,40	0,10	9,44	-23,45	-0,54
>sfr-miR-10459-4	GAAAUACACUAGUCU UGCCGUGCCUAUGUCG GUUAGCUGUCUGAUC UUCCAGACAGCGUAAG CUGCAGAGCUCAAUCG UCCCGCUAGUGAUGUA	95,00	24,21	22,11	25,26	27,37	49,47	49,47	1,24	0,96	-29,20	-25,80	0,07	14,3 4	-30,74	-0,62
>sfr-miR-10514	AGAAGCCAACCUCACC CCUAAACGGCCUGCCA CAGGCCCGAAAUUGCG CGCUUGCCGAUGGCGG GUCGUCAAUUUCGUCU AGGGGUGAGGUUAGC UUCC	100,00	28,00	20,00	31,00	20,00	59,00	40,00	1,00	0,90	-60,10	-57,70	0,23	4,97	-60,10	-1,02

>sfr-miR-10468	CAGUACCUCCGCGUCG CGCGACUUCCCGUCGA AACGUGCAGUGCACUU GGCGAAGUUGCCUGU GACCGCGGACGGGUAG AC	82,00	31,71	17,07	31,71	18,29	63,41	35,37	1,07	1,00	-35,80	-35,80	0,10	6,90	-43,66	-0,69
>sfr-miR-307b	UGCCUCCUCCACGAUC GCUCACUCAAGGAGGU UGUGAUGGGCCGAGC AACUAGUGCACACAUC ACACCCAGGUUGAGUG AGUAACCGGGAUGA GAUC	99,00	27,27	25,25	27,27	19,19	54,55	44,44	0,76	1,00	-36,90	-36,00	0,31	5,48	-37,27	-0,68
>sfr-miR-307	GAUCUCAUCCCCGGUU ACUCACUCAACCUGGG UGUGAUGUGUGCACU AGUUGCUCGGCCCAUC ACAACCUCUUGAGUG AGCGAUCGUGGAGGA GGCA	99,00	27,27	19,19	27,27	25,25	54,55	44,44	1,32	1,00	-42,40	-40,40	0,03	7,66	-42,83	-0,79
>sfr-miR-2a-1	UUAAAUGUGGUUUGG CAUCAAAGUCGGCUUG UCAUAGGUCAUAACG UAGCUAUCACAGCCAG CUUUGAUGAGCAUGA CCUCAUUCU	88,00	21,59	25,00	21,59	30,68	43,18	55,68	1,23	1,00	-29,60	-29,20	0,14	4,68	-33,64	-0,78
>sfr-miR-2b	AGGAAUGAGGUCAUG CUCAUCAAAAGCUGGCU GUGAUAGCUACGUUA UGACCUAUGACAAGCC GACUUUGAUGCCAAAC CACAUUUA	88,00	21,59	30,68	21,59	25,00	43,18	55,68	0,81	1,00	-27,00	-27,00	0,10	5,20	-30,68	-0,71
>sfr-miR-2a-2	CGCGAGUGCAGCGGCG CUCACAAAGUGGCUGU AAUGUGUGUCCCCGUA CAUAUCACAGCCAGCU	87,00	27,59	21,84	26,44	22,99	54,02	44,83	1,05	1,04	-43,90	-43,90	0,21	3,78	-50,46	-0,93

	UUGAUGAGCGCUGAG CAUCAUU															
>sfr-miR-2c	AAGCGCGUCAGGGGG UCAGCAAAGUGGUUG UGUCUUAUGAGAUUC AUGAACAUUACACAGC CAGCUUUGUUGACUUC UUUGUCGCGUAU	90,00	26,67	22,22	18,89	31,11	45,56	53,33	1,40	1,41	-43,20	-43,20	0,06	7,97	-48,00	-1,05
>sfr-miR-11	UCUCGUGGCUAGCACU UUGGCUGUGACCUGU GUGUCGUAGUCACAUC ACAGUCAGAGUUCUA GCUACGCGG	72,00	27,78	16,67	25,00	29,17	52,78	45,83	1,75	1,11	-37,90	-37,90	0,11	5,38	-52,64	-1,00
>sfr-miR-10497	AAUACAACCACAAGCU GCCUUCACACCAAACG GUGUCUGGUCAGGUU CUUAUUCCUGACCAG ACUCGUUUUGGUUGA GAGGCCGCUGUGGUUC GUUCG	100,00	24,00	20,00	27,00	28,00	51,00	48,00	1,40	0,89	-50,80	-46,40	0,02	7,85	-50,80	-1,00
>sfr-miR-10503-1	GGGCAUUGGAUCGUU CGAUUCCCUGCAACAU GGUUGAGUUGGACGG UGCUGGUGUACGUGU CAUGUUGGUGAACGA GCGCUGUCAACUGGGA	93,00	35,48	17,20	18,28	27,96	53,76	45,16	1,63	1,94	-30,80	-29,60	0,04	14,8 8	-33,12	-0,62
>sfr-miR-10503-2	GGGCAUUGGAUCGCUC GAUUCCCUGCAACAUG GUUGAGUUGGGCGGU GCUGGUGUACGUGUC AUGUUCGUGAACGGG CGCUGUCAACUGGGA	93,00	36,56	15,05	20,43	26,88	56,99	41,94	1,79	1,79	-32,70	-32,10	0,07	17,7 8	-35,16	-0,62
>sfr-miR-3192	GACGUACUUUUUUU AAAUGGGACAACCCCA CCACAACCUCUGGGAG GUUGUAGCGGGUUUG	85,00	21,18	27,06	20,00	30,59	41,18	57,65	1,13	1,06	-23,40	-18,60	0,08	13,9 9	-27,53	-0,67

	UUAUUUCAGUAAC AUACUGA															
>sfr-miR-8515-1	UCCCUGACUGUUUCAU UCCAUUACCUACGCGA AAUCGUACGCGGCCGG UAAUGGAAUGAAACA AACAGAAC	72,00	19,44	30,56	26,39	22,22	45,83	52,78	0,73	0,74	-35,30	-35,30	0,92	0,20	-49,03	-1,07
>sfr-miR-8515-2	GCUCGCUUCCCCUUC CUGACUGAUUCAUUC AUUACCUACGCGAAUU CGUACGCGGCCGGUAA UGGAAUGAAUCAAAC AGUCGGGAUGGGAAC GCGAGC	101,00	24,75	22,77	27,72	23,76	52,48	46,53	1,04	0,89	-61,90	-61,90	0,30	2,61	-61,29	-1,17
>sfr-miR-10492b-1	ACAAAACGACGCAAGA GCGCGUUCGAUGCUCU GAUUGGUCGGUGAGA AUUAAUCAACCAAUCA GAGUAUCGAACGCGCU CUGGUUUCGUUUCGC	95,00	25,26	26,32	23,16	24,21	48,42	50,53	0,92	1,09	-59,90	-59,90	0,55	1,49	-63,05	-1,30
>sfr-miR-10492b-2	UUUAGCGCGUUCGGU GCUCUGAUUGGGCGGC UUGAAUAAAUCAACC AAUCAGAGUAUCGAA CGCGCUCUC	71,00	25,35	22,54	23,94	26,76	49,30	49,30	1,19	1,06	-42,20	-41,50	0,23	3,96	-59,44	-1,21
>sfr-miR-10492c-1	UAGAAACGAGAGCGC GUUCGGCGCUGUGAU UGGCCGGCUCUAAUGA ACCAACCAAUCACAAC GCUGAACGCGCUUCGU GUUGAC	85,00	27,06	24,71	27,06	20,00	54,12	44,71	0,81	1,00	-47,70	-47,70	0,23	5,28	-56,12	-1,04
>sfr-miR-10492c-2	UAAAGUAAUCAAAC GAGAGCGCGUUCGGCG CUGUGAUUGGCCGGCU CGAAUAAACCAACCAA UCAGCGCGCCGAACGC	101,00	23,76	26,73	26,73	21,78	50,50	48,51	0,81	0,89	-66,80	-66,80	0,55	1,02	-66,14	-1,31

	GCUCUCGUUUCGAUUA CUUUA															
>sfr-miR-10492a-1	AGUAAUCGAAACGAG AGCGCGUUUGGCACUC UGAUUGGCCGGCUCGA AUUAAACUAACCAAUCA GAGCGCCGAACGUGCU CUCGUUUCGAUUAUA	95,00	24,21	26,32	24,21	24,21	48,42	50,53	0,92	1,00	-63,70	-63,70	0,70	0,91	-67,05	-1,38
>sfr-miR-10492a-2	AGAGCGCGUUCGACGC UUUGAUUGGUCGGCU CGAAUAAACCAACCAA UCAGAGCGCCGAACGU GCGCA	69,00	27,54	26,09	27,54	17,39	55,07	43,48	0,67	1,00	-39,30	-39,10	0,94	0,24	-56,96	-1,03
>sfr-miR-10492c-3	GACGAUAUCGAAACG AAAAUGCGUUCGGCGC UGUGAUUGGCCGGCUC AAAUGAACUAUCCAA UUACAGCGCCGAAUGC ACUUUCGUUUCGAUA UCUUU	99,00	22,22	26,26	23,23	27,27	45,45	53,54	1,04	0,96	-63,10	-63,10	0,69	0,93	-63,74	-1,40
>sfr-miR-10492c-4	AACGAGUUCGAAAUG AAACCGCAUUCGGCGC UGCGAUUGGCCGGCUC CCAUGAAUCAACCAAU CAGAGCGCCGAACGUG CACUCGUUUCGAUCUC UUU	99,00	23,23	24,24	29,29	22,22	52,53	46,46	0,92	0,79	-44,30	-44,30	0,49	2,81	-44,75	-0,85
>sfr-miR-10492c-5	GAGAGUGUGUUCAGC GCUGUGAUUGGCCGG UUCGAAUAAACCAACC AAUCACAACGCUGAAC AACUAAG	70,00	24,29	31,43	22,86	20,00	47,14	51,43	0,64	1,06	-34,30	-34,30	0,57	1,34	-49,00	-1,04
>sfr-miR-10471-1	GGUAAACUAGAAUGC AUUCGGUACCGGUGCU CUGAUUGGUUAGUUC AUUGGAGCCGGCCAAU	97,00	26,80	19,59	22,68	29,90	49,48	49,48	1,53	1,18	-44,40	-44,40	0,24	6,23	-45,77	-0,93

	CAGAGCGCCGAUCGCC GUGUCAUUUCGAUUU CUU															
>sfr-miR-10492c-6	ACUAGAGCGCGUUCGG CGCUGUGAUUGGUUG GUAUAUUGGAGCCGG CCAAUCACAGCGCCGA ACGCACUCUCGA	75,00	30,67	20,00	26,67	21,33	57,33	41,33	1,07	1,15	-56,70	-56,70	0,83	0,35	-75,60	-1,32
>sfr-miR-10492a-3	UCUAAACGAGACCGCG UUCGGCGUUCUGAUU GGUUGGUUUUUAUUCGA ACUGGCCAAUCAGAGC GCCGAACGUGUAAUCG UUUCGA	85,00	27,06	21,18	22,35	28,24	49,41	49,41	1,33	1,21	-54,50	-54,50	0,34	2,29	-64,12	-1,30
>sfr-miR-10492a-4	UGAGAUAAAACGAGU UCGUCGCGUUCGGCGC UGUGAUUGUUGGGUU CAUUGAAGCCGCGCCAA UCACAGCGCCGAACGU GUCUCGUCUCGUUUCG AUCUCG	101,00	28,71	17,82	24,75	27,72	53,47	45,54	1,56	1,16	-61,30	-61,30	0,21	2,90	-60,69	-1,14
>sfr-miR-10492b-3	UUUACGAGAGCGCGU UCGAUGCUCUGAUUG GUUGAUUUUUAUUGAG CUGGCCAAUCAGAGUG CCGAACGCGCUCUCGU UUC	81,00	27,16	17,28	22,22	32,10	49,38	49,38	1,86	1,22	-50,00	-48,80	0,17	3,70	-61,73	-1,25
>sfr-miR-10492b-4	UGAAUUAUAACGAAA AAAGAGCGCGUUCGA UGCUCUGAUUGGUUG UUUUAUUCAAGCUGG CCAAUGAGAGCGCCGA ACGCAAUCUCGUUUCA AUAAUAAAC	101,00	21,78	30,69	18,81	27,72	40,59	58,42	0,90	1,16	-38,70	-38,70	0,20	6,21	-38,32	-0,94
>sfr-miR-10492b-5	AACGAGAACACGUUCG AUGCUCUGAUUGGUU	76,00	26,32	21,05	23,68	27,63	50,00	48,68	1,31	1,11	-36,80	-36,80	0,19	5,53	-48,42	-0,97

	GUUCAUUCGCGCCGGU CAAUCAGAGCGCUGAA CGGAUUCUCGUU																
>sfr-miR-10492b-6	ACGUUGAAACGACACC GCGUUCGAUGCUCUGA UUGGUUAGUUCAUUG AAGUUGACCAAUCACA GUGACAAACACAGUU UCAUUUCAAUAA	92,00	18,48	30,43	20,65	29,35	39,13	59,78	0,96	0,89	-23,50	-13,80	0,04	20,5 3	-25,54	-0,65	
>sfr-miR-10862	AUCGAACCGUUACUGC AUUUCUGUCCUCUGAU UGGCUGCUCCGAGUUA AACGACCAAUCAGAGC GCUAGUAGCGGUAAC GUUUCAAA	88,00	21,59	25,00	25,00	27,27	46,59	52,27	1,09	0,86	-31,70	-31,30	0,17	5,40	-36,02	-0,77	
>sfr-miR-10492a-5	AUGCGAUCAAAACAU UACGGCAUUUUGUCCU CUGAUUGGCUGAUUA ACUUGGAGCUGCCCAA UCAGAGUGCCGAAAG UCAUCAAAAUGUCGA UCGUGU	99,00	22,22	27,27	21,21	28,28	43,43	55,56	1,04	1,05	-29,40	-26,40	0,03	13,4 2	-29,70	-0,68	
>sfr-miR-10492a-6	GCAACAGUUUUUGAG UUUAGCCCUCUGAUUG GUUGGUCGGCCAAUCA GAGCGCCGAACGUCAU AAACCUGUCGA	75,00	25,33	22,67	24,00	26,67	49,33	49,33	1,18	1,06	-30,40	-30,40	0,13	4,01	-40,53	-0,82	
>sfr-miR-10492a-7	UUACCGCGUUUAGCUC UCUGAUUGGUUAGUU AGUCGGAACAGGCCAA UCAGAGCCCGAAGCUG CGGGCU	70,00	28,57	20,00	24,29	25,71	52,86	45,71	1,29	1,18	-28,60	-28,60	0,37	4,97	-40,86	-0,77	
>sfr-miR-124c	AUGAGUCCUCUCUUGG CAUUCACCGCGUGCCU UAUGAUUGCUACAUA ACACAUAAGGCAACG	90,00	25,56	26,67	23,33	23,33	48,89	50,00	0,88	1,10	-38,80	-34,90	0,02	6,83	-43,11	-0,88	

	UAGUGAACGCGAGGA GGAGGACUGCU															
>sfr-miR-124	AGCAGUCCUCCUCCUC GCGUUCACUACGUUGC CUUUAUGUGUUAUGU AGCAAUCAUAAGGCAC GCGGUGAAUGCCAAG AGAGGACUCAU	90,00	23,33	23,33	25,56	26,67	48,89	50,00	1,14	0,91	-37,90	-37,90	0,02	7,27	-42,11	-0,86
>sfr-miR-184b	UAUUGUAUUGGACAU GCCCUUAUCAGUUCUC CGUCCAGUAGUAUGA AAGCACACGGGCAAGA AGAAUGACAAGGGCA CGGCACAAUACAUUC GCAUCUUUUAUGGCG UAUCUAUUUGCACAG UGGUCUUGCAACAAU GCUUAGGCACGUCGUC GGCAAUAAAACGUCA AGACAGAACAA	93,00	22,58	31,18	22,58	22,58	45,16	53,76	0,72	1,00	-29,00	-25,60	0,03	9,61	-31,18	-0,69
>sfr-miR-9544a	AUUAUACACCGCAUUC AGGUACAUAACAUGCAC ACAUAACACCGCAUUC AGGUACCUACUUGCAC AUGUACAC	87,00	21,84	28,74	21,84	26,44	43,68	55,17	0,92	1,00	-21,80	-20,50	0,04	7,51	-25,06	-0,57
>sfr-miR-466o	UGGAUUCGCGUCUUU UCCUGUUUCAGAUCUA CAUCUGGCAAUGGAG AACCCAUUGGACGAUG UAGUUGUGAAACAGG AAAAAACGCUAAUGA U	73,00	12,33	31,51	30,14	24,66	42,47	56,16	0,78	0,41	-19,50	-19,50	0,54	1,33	-26,71	-0,63
>sfr-miR-10506	UGUAUGUGUCUGCCCC CAUUGUAUGUAUGUG UGCAUGUCAUAAAGG AUUUACAUACAUAACA	94,00	23,40	28,72	18,09	28,72	41,49	57,45	1,00	1,29	-44,10	-43,10	0,33	2,28	-46,91	-1,13
>sfr-miR-297a		81,00	20,99	27,16	18,52	32,10	39,51	59,26	1,18	1,13	-25,90	-25,70	0,42	2,85	-31,98	-0,81

	UACAUCGAGCAGCAAU GGU															
>sfr-miR-279d	AAUUUAACUUGAUGA UAUUUGUGGGCGAGU UUGCUUCUGGUGCAU GUUCAUUACUAAUCA UGACUAGAUGCAUACU CGUCUGCAGCCAUCAA CUUAAUCU	101,00	17,82	24,75	19,80	36,63	37,62	61,39	1,48	0,90	-34,10	-34,10	0,27	3,20	-33,76	-0,90
>sfr-miR-279	GGUCAAUUUCUUUCG AUGAGUGGAGGUUUA GUGCAUGUUUCUGUA CAUCAUGACUAGAUGC ACACUCAUCCAUGGAA GUUGCGA	85,00	23,53	23,53	18,82	32,94	42,35	56,47	1,40	1,25	-33,90	-33,90	0,18	2,37	-39,88	-0,94
>sfr-miR-10b	GAUUAACUGACACAA GUACCGACGUAUGGU AGGGCCAAGUACGUCU CUACAGUUCUAAUUG UCUAUCUACAUAAUG UACAUGUGUAGGUAC AC	94,00	20,21	29,79	20,21	28,72	40,43	58,51	0,96	1,00	-21,50	-20,60	0,04	20,5 9	-22,87	-0,57
>sfr-miR-10496	UCUUGGUCUUUGAUU GAAGGCACUGGUUUC UUGAGGGUACUACCU GUACCCUCUUGAUACU GCAGAUUGGCUCAAAC UCAGAUGAGCAGCA	92,00	23,91	22,83	21,74	30,43	45,65	53,26	1,33	1,10	-34,40	-31,10	0,09	10,7 6	-37,39	-0,82
>sfr-miR-10482	GAAUACGUAGUUCGG UAUCGGUGAAUGCCA ACGGAUUUCUAGCU ACUAGCUAUACUCACA GAAAUCCUUGAUAU UCACCGGUAUCGCGCU ACGUAACA	101,00	19,80	28,71	22,77	27,72	42,57	56,44	0,97	0,87	-48,60	-48,60	0,52	1,14	-48,12	-1,13

>sfr-miR-10488	UUUGUGCAAUCGAUG UUUUUUUCCAAUACU UUAGUUUUUAACCACC CUCUAGAAAACAACAU GAAUGCUGUCGGAU AAACAUUGAUUAGGC ACUCC	98,00	14,29	29,59	20,41	34,69	34,69	64,29	1,17	0,70	-20,50	-20,10	0,14	14,8 4	-20,92	-0,60
>sfr-miR-2335	CACAAUCAUACAUUGU CUCCCUUGUAUCUAUG UUCUGGCACACUAUGG GUUAAAUGUACCAUC UGGAUACAUAAGAUAA UGAUGACUAAUUUAU UAUUUA	101,00	13,86	29,70	17,82	37,62	31,68	67,33	1,27	0,78	-19,50	-18,00	0,09	18,2 1	-19,31	-0,61
>sfr-miR-71	GUGUGAAAGACAGGA GUAGUGAGAUGUCCU CGACAUCACAAAUUCU CACUACCUUGUCUUUC AUGC	67,00	20,90	26,87	22,39	28,36	43,28	55,22	1,06	0,93	-33,30	-33,30	0,47	2,73	-49,70	-1,15
>sfr-miR-9807	AACCAUUAACCUGUUG UCUGUCGGAACGCAGA UCUACGCGAGACACAA UGUGUUUUCUAUUGU GUCUUAUAUACCGACA GACAAGGGGUUAAAU GACU	99,00	21,21	28,28	20,20	29,29	41,41	57,58	1,04	1,05	-42,40	-42,40	0,15	3,73	-42,83	-1,03
>sfr-miR-10495-1	CUGCAUCAACCGGGGA UUGUCCUUUGUGUAC UCCAUAAGUGCAUACA GGGAUAAUCGCCGGCU GAUGUCC	71,00	25,35	19,72	25,35	28,17	50,70	47,89	1,43	1,00	-40,00	-40,00	0,71	0,77	-56,34	-1,11
>sfr-miR-8485	GUGGUCCACACACACA CACACACACACACACA CACGUUGUUUGUGCU UGUAUGUGUGUGGUA CAUU	67,00	19,40	25,37	26,87	26,87	46,27	52,24	1,06	0,72	-27,60	-25,30	0,30	6,82	-41,19	-0,89

>sfr-miR-466f-2	UCUCACGCACUUUUAC ACACACGCAUACACAC ACACAUACACAAAGGA GGGUUGAAUUCUUAG UGUUGGUGUUUCAA CAUUGCGGUGGUA	92,00	19,57	29,35	22,83	27,17	42,39	56,52	0,93	0,86	-22,50	-22,50	0,44	2,87	-24,46	-0,58
>sfr-miR-466f-3	CGAACUCACGCACUUU UACACACACGCAUACA CACACACAUACACAAG GGAGGGUUGAAUUCU UAGUGUUGGUGUUUC AAACAUUGCGGUGGU ACC	97,00	20,62	28,87	24,74	24,74	45,36	53,61	0,86	0,83	-22,70	-22,70	0,50	3,17	-23,40	-0,52
>sfr-miR-2733b-1	UUCGUCACUCAGUAGC GAUUGUCACACUCUCA GUAGACAGUGUGUCA UUGUGUCACUGGGUG UGUGAUAUAGAUAC UGAUGACUAC	88,00	23,86	23,86	19,32	31,82	43,18	55,68	1,33	1,24	-40,60	-38,70	0,28	5,33	-46,14	-1,07
>sfr-miR-297b	UAUUAUGUGUGUGUG UGUGUGUCAUAAAAU UGUUGAAAUGGUGUA UGAUUGUAUUGUUGC UAUCCUAUUAUUCUU AUACAUACACACAUAC ACAUACAU	100,00	17,00	25,00	12,00	45,00	29,00	70,00	1,80	1,42	-30,90	-26,18	0,02	37,6 7	-30,90	-1,07
>sfr-miR-2733b-2	UGUGUGUCACUCAGU AGCCGUUGUCAUUCGC CCAGUACAUAUGUGUG UCAUUGUGUCACUGG GUGUGUGAUAUAGA UACUGUUGACUACCGU	93,00	25,81	19,35	19,35	34,41	45,16	53,76	1,78	1,33	-41,80	-40,30	0,35	3,76	-44,95	-1,00
>sfr-miR-12312	AUUAAUUCAGUAUAGC AUACGCCAUACGAUUU UAAAUUUAAAUGACA AGUGAAUUAGUGGGU	88,00	20,45	30,68	11,36	36,36	31,82	67,05	1,19	1,80	-18,50	-11,70	0,05	25,1 6	-21,02	-0,66

	UGGAGCUGUGCUAUA UCUUUAAUGGU															
>sfr-miR-2489	UCGUAGUAGAUUGUU GUUUGUGAUGUUAAG GAAAUGUGUCAUUUU UUAUUGUAUGUUGUA UUUCGUUACUAUCGAC AAUCAAAUAUGUAAU ACAU	96,00	18,75	27,08	8,33	44,79	27,08	71,88	1,65	2,25	-19,40	-19,40	0,19	28,2 6	-20,21	-0,75
>sfr-miR-297	UGUAUGUAUAUUUAU AUGUAUGUAUGUAUG UGUGUGUGUAUGUAU GCAUAUAUGUAUGUA AGUAUGCAUCUA	73,00	21,92	26,03	4,11	46,58	26,03	72,60	1,79	5,33	-26,40	-26,40	0,22	2,51	-36,16	-1,39
>sfr-miR-574	CGAUUUUAUUUAUGGA UCAUACAUAAGACAUG UGUGUGUGUGUGUGU GUGUGUGUGAUGUGU ACAUAUUUAUGUAUU UACGAUAAAGUAUAA AGU	94,00	24,47	27,66	6,38	40,43	30,85	68,09	1,46	3,83	-21,50	-21,50	0,05	13,5 1	-22,87	-0,74
>sfr-miR-11660	UUCAACACAUUCGCGG GCUUUCGUCGCCAUUU UGUAAUUAAAACGUC ACUUUAUUAUUGACA UAUGUAAGAUGGCGA CGUGUCCUCGGAGUGU UCC	98,00	20,41	22,45	21,43	34,69	41,84	57,14	1,55	0,95	-33,30	-33,00	0,10	6,25	-33,98	-0,81
>sfr-miR-1410	AAGGGAAUUUAGAUAU CCGUUUUAUAAUCACAA AGAGUCAUUAGUGAG UGAGACGGAGUCGUG AGUAAUUCCUAC	74,00	24,32	32,43	13,51	28,38	37,84	60,81	0,88	1,80	-24,00	-20,80	0,11	10,5 9	-32,43	-0,86
>sfr-miR-306a	UGACAGUCAGGUACU AGGUGACUCUGAGUG AGUGUGCCCGUGUCAG	70,00	34,29	18,57	20,00	25,71	54,29	44,29	1,38	1,71	-22,10	-20,90	0,39	6,69	-31,57	-0,58

	GUACUAGGUGACUCU GAGUGGCC															
>sfr-miR-466i-1	CCUACAGAGCACAGAU AAUCACAGGAAAUGU UUUCGGCUCAUGAAU AUUUAACGACAUUAG GAUGCAACGUAAUUC UGUGUGUGUGUGUGU GUGUGUG	100,00	25,00	27,00	16,00	31,00	41,00	58,00	1,15	1,56	-27,20	-24,10	0,08	14,5 2	-27,20	-0,66
>sfr-miR-1b	CACGCCCCGCGCAGCUC CAUACUUCUUUACA CCAUACGAUUUAAGC ACUAUGGGAAGUAAG GAAGCACGGAACUCGC GCAAGGCUUU	90,00	21,11	27,78	27,78	22,22	48,89	50,00	0,80	0,76	-33,20	-33,20	0,67	1,87	-36,89	-0,75
>sfr-miR-1770	AUCGCAAGUUACUGA GGUGACGAGGGAAGU GAGGAGGCCUGCGGG GGAGGAGGGAGACCG GUCCAACCUCGUCACU AGGUAUACUCGCAGU	92,00	38,04	23,91	20,65	16,30	58,70	40,22	0,68	1,84	-34,90	-30,60	0,25	27,5 9	-37,93	-0,65
>sfr-miR-466f-4	GUAACACAUGCACAUG CGUGUAUACGUCGUA UGAUAGACGUGAAGA CGUGUGUGUGCAUGU GUGCA	67,00	29,85	25,37	16,42	26,87	46,27	52,24	1,06	1,82	-33,60	-33,60	0,24	4,36	-50,15	-1,08
>sfr-miR-1	AAAGCCUUGCGCGAGU UCCGUGCUUCCUACU UCCCAUAGUGCUUAUA AUCGUAUGGAAUGUA AAGAAGUAUGGAGCU GCGCGGGCGUG	90,00	27,78	22,22	21,11	27,78	48,89	50,00	1,25	1,32	-42,70	-42,70	0,14	9,22	-47,44	-0,97
>sfr-miR-10235	CCCUGUAACGAAACGU CACGUGCAUCCUACUC CCUACACACCAUGUAA AUGUGUGGACGAGUU	93,00	21,51	27,96	25,81	23,66	47,31	51,61	0,85	0,83	-23,20	-23,20	0,10	10,5 2	-24,95	-0,53

	GACUGCGACAUGAGU AAUCGUGAGCAUUA															
>sfr-miR-92a	GGAGGCCCCGUGUGGCU GGGCGGUGACUGGCGC UAUAUUCGGUUUGUG UGCGAUAAUUGCACCAG UCCCGGCCUAUCCAAG CGGGCCCCGU	89,00	35,96	12,36	26,97	23,60	62,92	35,96	1,91	1,33	-47,80	-47,70	0,08	5,88	-53,71	-0,85
>sfr-miR-10475	UGUGUGAAUUGAAGC GUCUCCGUGUGGUCGG CGAACACUGCAACAUG CGACAUGUCGGUACCA UACUCCACCACACGGA GAUGACUCUGAUUCAC UCA	99,00	24,24	24,24	27,27	23,23	51,52	47,47	0,96	0,89	-42,80	-42,20	0,07	9,00	-43,23	-0,84
>sfr-miR-989	GAAGUUCAGGGAUGA AAUGGCCAUUACCUCA CAGUCACGUGGUCAUG ACGAUACAACGUGUG AUGUGACGUAGUGGA AGUCUCGACCUGAACG GU	96,00	29,17	27,08	19,79	22,92	48,96	50,00	0,85	1,47	-37,80	-37,80	0,16	6,47	-39,38	-0,80
>sfr-miR-7398f	UCGAAGUGUAGAAGA GAGUGAUGCCGACGAC CACAUCGUCUGCAGCA CGCUGACUGUCGACAC UUAUU	69,00	26,09	26,09	24,64	21,74	50,72	47,83	0,83	1,06	-21,90	-21,30	0,18	7,11	-31,74	-0,63
>sfr-miR-4756	GCUGCUGAACAAGCGU GAAGCGUGAGUGAGG GAGGCGCUCACUCCUC GCCUCACCGUCACCG UCCUCAGCGUUUCAG CCGU	84,00	28,57	16,67	33,33	20,24	61,90	36,90	1,21	0,86	-30,80	-26,10	0,01	14,9 3	-36,67	-0,59
>sfr-miR-10508a-1	CGUCGCACGCGCCGCA UAGUACGCGCGAUUCC AGGCAGAGGGCGCGA	100,00	35,00	19,00	30,00	15,00	65,00	34,00	0,79	1,17	-34,10	-33,80	0,04	19,6 8	-34,10	-0,52

	UGGGCGCGAAACUCGA GUCGUCAGUAGCAGCG UGACAAUCGCCGCGUC GUGU																
>sfr-miR-10508b-1	UUUCAUGACCAGCUUU CUGCGACAAUCGUAUC GCGCGUAGCAGCGCUG CAAUCGCCGCGUCGUC GUGUGUCGCGGAAUG CUAAGUCCUGGUC	93,00	27,96	16,13	29,03	25,81	56,99	41,94	1,60	0,96	-36,70	-32,30	0,05	11,7 6	-39,46	-0,69	
>sfr-miR-137b-1	UGGAGGUCACGCGUA UUCUUGGGAAUUAAC ACACAUUACGAGAUG UUAUUGCUUGAGAAU ACACGUAGCUGAC	74,00	24,32	28,38	17,57	28,38	41,89	56,76	1,00	1,38	-21,20	-21,10	0,06	3,84	-28,65	-0,68	
>sfr-miR-283	GAAUCGGUUUCCCCGGC UAAAUUUCAGCUGGU AAUUCUGGGGUGCUG UCGCCAGACUACCAG UUGGUAUGAGGCCGG GAAUCGAACG	88,00	30,68	21,59	22,73	23,86	53,41	45,45	1,11	1,35	-50,70	-49,90	0,09	6,05	-57,61	-1,08	
>sfr-miR-137b-2	GUCACGCGUAUUCUUG GGGAAUUAACACACA UUACGAGAUGUUAUU GCUUGAGAAUACACG UAGC	66,00	22,73	28,79	18,18	28,79	40,91	57,58	1,00	1,25	-18,80	-16,40	0,14	5,04	-28,48	-0,70	
>sfr-miR-7086	UACAAUUACGAAACCG GUUUUCCUUGCGACAA AGCAAAGGAAGAGCA AGAGGAGAAAGGUUU CGUAGAUUACC	74,00	24,32	37,84	17,57	18,92	41,89	56,76	0,50	1,38	-21,10	-21,10	0,26	8,33	-28,51	-0,68	
>sfr-miR-3529	UGAGCCGUACUUUGU AGGGAGAUUAGUGAU UUCUUGUUACCCAAAU CAAAACAACAAAUCA	90,00	18,89	31,11	21,11	27,78	40,00	58,89	0,89	0,89	-33,90	-33,90	0,30	5,15	-37,67	-0,94	

	CUAGUCUCCAAACAA CGUGUGGCCUG															
>sfr-miR-7b	CAGGCCACACGUUGUU UGGAAGACUAGUGAU UUUGUUGUUUUGAUU UGGGUAACAAGAAAU CACUAAUCUCCCUACA AAGUACGGCUCA	90,00	21,11	27,78	18,89	31,11	40,00	58,89	1,12	1,12	-27,60	-27,60	0,03	12,3 3	-30,67	-0,77
>sfr-miR-34	CUUCUGUAAGGUAGU CCGCGUUGGCAGUGUG GUUAGCUGGUUGUUU AAAUUGCUACAACAGC CAGUACGACACUGCC UCCGCGUGCUACCUAA AGCAA	100,00	25,00	23,00	24,00	27,00	49,00	50,00	1,17	1,04	-52,50	-52,50	0,12	4,04	-52,50	-1,07
>sfr-miR-33	AAUAACUAUUACGGG AGUGCAUUGUAGUUG CAUUGCACUGGUUUAC AUGAGUUGUGCAAUA UUACUACAAAGCAAA UCUCGCAUUAGUUGCA	93,00	20,43	30,11	16,13	32,26	36,56	62,37	1,07	1,27	-31,50	-30,60	0,24	3,50	-33,87	-0,93
>sfr-miR-10481	GUAAAACGGGGUGAA UAGAAUUCGAGGGGU UAAUAGAUACAGAAU AGGGGUGAAUAAAUG AUGGGAAAAUGUCCU UUGAAUACU AUUCACC CCGUUUCAC	101,00	25,74	34,65	12,87	25,74	38,61	60,40	0,74	2,00	-34,60	-34,04	0,03	8,70	-34,26	-0,89
>sfr-miR-509	GCAGAGUAGUUUCAA AAAGUGACUGAUUGA UACGUCUGUCAAUUCA GUUCUAAUUGCACUA UUCAAA	68,00	17,65	30,88	16,18	33,82	33,82	64,71	1,10	1,09	-20,10	-20,10	0,23	2,58	-29,56	-0,87
>sfr-miR-8327	UGAUUUAAUAAUACG AGUACA UCCCUACCU AGUGAAGCACGUGUA	74,00	20,27	25,68	13,51	39,19	33,78	64,86	1,53	1,50	-21,20	-21,20	0,16	3,10	-28,65	-0,85

	GGUUUUGUAUUUCGU UUUGAUAAGGU															
>sfr-miR-10483	UGAAGUGUGUACUUA CGUCACCUGUAAUUGU UCGUUCCCCUUUGUAA AUAUUAAAGGGGCCG AACAAUCCAGACGAU GUUUGAACAGACUCG U	95,00	21,05	26,32	18,95	32,63	40,00	58,95	1,24	1,11	-34,90	-33,20	0,09	5,68	-36,74	-0,92
>sfr-miR-100	CGGUUAAGCGGAGAC AUUUCCACCCUUUGAA CAAGUUCUUGUUUAU GUUUGCUCAUUAAAC AAAUAAAAGUGGAAA UUCUGUCUUAACUCU	91,00	16,48	29,67	17,58	35,16	34,07	64,84	1,19	0,94	-23,60	-23,60	0,15	10,4 3	-25,93	-0,76
>sfr-miR-8456	CAAUUACAGUUUGCA ACAUUGUUUUAAUUA CUUACUUUGUUUUUAU UUACUUUUUAAACG AGUAGGUAAUUGUUG GAACCAGUGUUUCA AUGAUAAAGAU	101,00	15,84	28,71	10,89	43,56	26,73	72,28	1,52	1,45	-22,00	-22,00	0,05	10,1 5	-21,78	-0,81
>sfr-miR-11902	AUCAUAGCUUGAGCU AAUGACGCGUAAAAG UUCUAAACUUCACGA AGCGUUGAAAAUAUG ACAAGUGAUAAAGCGC AAGCGAUUCA	87,00	20,69	36,78	17,24	24,14	37,93	60,92	0,66	1,20	-20,20	-19,70	0,12	7,17	-23,22	-0,61
>sfr-miR-6399	CUCAGAGACAUUAAU GAUUACUUACACUAA UCUUUUGCAAUGAUG GUAUUAAUCGUAUCG UAUAUCGUAACAAU AAGCAAUCAUUAUU GUCUCUAAAG	100,00	13,00	34,00	16,00	36,00	29,00	70,00	1,06	0,81	-23,00	-23,00	0,03	13,0 1	-23,00	-0,79

>sfr-miR-2767	GGAAAACUUUUACCG UCGACACAAGUAAAUC UCGUGCGGCUUGUGU UUAGAUUAAGCCGUU CGAGAUUUACUUAAG UCAAUGGUAAAUAUU UCUA	96,00	19,79	29,17	16,67	33,33	36,46	62,50	1,14	1,19	-43,80	-43,80	0,23	4,07	-45,63	-1,25
>sfr-miR-2218b	UGAAUAAAUCAAGU GUCUUUAAAACCAAU UAUGAGUACGAUACA UUAGUGGUUAUAAAA UUUGUAGUUUGUAAG GAUAAAUUGAAGUAU GCC	94,00	18,09	37,23	8,51	35,11	26,60	72,34	0,94	2,13	-19,70	-19,70	0,24	7,92	-20,96	-0,79
>sfr-miR-10489-1	AUACCAAGUCGGUUA AUCUAUUUAGUGUAU GAUAUCAUGCCAAUA UUAAAAAUUUUAAC GUUUUAAACAAAUAG AUCGACUUGGACA	89,00	13,48	38,20	13,48	33,71	26,97	71,91	0,88	1,00	-19,60	-18,70	0,04	13,0 9	-22,02	-0,82
>sfr-miR-10489-2	GUAGUGUGAUACAUA CUAAUAAUCAAAUCA AGCGAUGUCCAAGUCG AUCUAUUUGUUUAAA ACGUUAAAUAUUUUU AAUAUUGGCAUGAUA UCAUACACU	101,00	13,86	35,64	13,86	35,64	27,72	71,29	1,00	1,00	-18,90	-17,30	0,05	12,2 1	-18,71	-0,68
>sfr-miR-9518	UGGUCCAGAUCUAGA UUAUCAACCAUUAUU UAUUUUUAUUGAUUU UGUCUGCAAUAUCAU AAGAAUGUAUCAUUU GAGACAAUGAGUCUG GAAUA	96,00	15,63	33,33	12,50	37,50	28,13	70,83	1,13	1,25	-22,60	-20,70	0,06	8,64	-23,54	-0,84
>sfr-miR-10474-1	GAAUAACGCGGAAUU AGAUUGUGAUUUUUU	98,00	17,35	35,71	9,18	36,73	26,53	72,45	1,03	1,89	-25,60	-24,40	0,12	8,06	-26,12	-0,98

	UUAUGUUGAUAGAUU AACGUAUGAAGAGUA UACUAUAUAAUAUC UCAAUCUAAUAUGGC GUUAAAC															
>sfr-miR-10262	UAAGUAGAAGAUGUG AUGUUAUGUUGGUAG CUUUUGGUAGGAUGA UGGUCUACCAACAUUA AUAACAAGUUCGACAC A	79,00	24,05	31,65	11,39	31,65	35,44	63,29	1,00	2,11	-20,10	-20,10	0,06	6,82	-25,44	-0,72
>sfr-miR-10504	UCAGACCUCUCGACCA GUCGUACAUAAGAGACC UGAAGUAUUCACAGC UUCAAAUCUCUGUGU AUGAUUGGCCGCGGGC GUCCCA	85,00	22,35	23,53	28,24	24,71	50,59	48,24	1,05	0,79	-42,30	-42,30	0,90	0,38	-49,76	-0,98
>sfr-miR-10474-2	AUGAACGCGGAAUUA GAUUGUAUUUUUUA UGUUGAUAGAUUAAC GUAUGAAGAGUAUAC UAUAUAAAUAUCUCA AUCUAAUAUGGCGUU AAA	94,00	17,02	36,17	8,51	37,23	25,53	73,40	1,03	2,00	-23,20	-22,00	0,23	6,42	-24,68	-0,97
>sfr-miR-10761	CGGGGAUAAAAGGCG UUACGAUUAUUCUUU UGAAUUUGUUCUUGU AUACUAACACGAAGA AUAUA AAAAGCAUUU CGUCACUGUUGUUCAU CCGUG	97,00	19,59	28,87	15,46	35,05	35,05	63,92	1,21	1,27	-19,20	-15,40	0,02	12,9 7	-19,79	-0,56
>sfr-miR-3924	UAAUAAGGUAUCGUU UUAGUAGUUAAGGAA UACAUUAUUUUGGC ACUUCCCUCAAACUUA UAUAUGUAUAUGUGA	98,00	15,31	31,63	13,27	38,78	28,57	70,41	1,23	1,15	-19,20	-17,80	0,08	20,1 7	-19,59	-0,69

	CUUCAAAAGUUACGU GUACUU															
>sfr-miR-9051	GUGAUGUUUUAGAAU UUAGACUCUUGAUAG AAUAAGGACUUAUUU UGUAGCAGACUUUUU UAAAACAAGUGUCUA UGGACUCCAAAACAUU AU	94,00	17,02	31,91	11,70	38,30	28,72	70,21	1,20	1,45	-19,40	-19,40	0,03	12,4 2	-20,64	-0,72
>sfr-miR-10500-1	GACGCAUAUUCAGUCC AACAGUGACCGUCAUA GAGAUAAAGGAAGCGU GUUCGGGCAUGUAUU CGUCGUAGGGGAGGU CACUUUGAACAAAAU UUGCAA	100,00	27,00	30,00	18,00	24,00	45,00	54,00	0,80	1,50	-28,30	-28,30	0,28	4,38	-28,30	-0,63
>sfr-miR-10500-2	GUCCAAGAGUAACCGU CAUAGAGAUAAAGGAA GUGUGUUCGGGCAUG UACUCGUCGUAGGGA AGGUUACUUUGAAC	76,00	30,26	27,63	15,79	25,00	46,05	52,63	0,90	1,92	-20,00	-20,00	0,20	4,83	-26,32	-0,57
>sfr-miR-10500-3	AAUUCUUCUCUGACGC AUUUUCAGCCCAAGAG UGACCGUCAUAGAGA UAAGGAAGUAUGUUC GGGCAUGUAUUCGUC GUAGGGAAGAUC	91,00	25,27	28,57	18,68	26,37	43,96	54,95	0,92	1,35	-24,10	-24,10	0,07	10,7 1	-26,48	-0,60
>sfr-miR-279b-1	ACUCCUACUUGCUCU AAUGGGUAUAUGUCU AGUGCCACAGUGAAU UAUUUCUGUGACUAG AUCUACACUCAUUGAG AGCUUUUAGGCCU	91,00	18,68	23,08	21,98	35,16	40,66	58,24	1,52	0,85	-32,20	-31,30	0,22	3,15	-35,38	-0,87
>sfr-miR-10461b-1	ACGGCUUGAAACUAG UCGAGUUCUCGUUAU UUAUUAUGUUAUCGG	95,00	20,00	25,26	17,89	35,79	37,89	61,05	1,42	1,12	-44,80	-44,80	0,13	6,97	-47,16	-1,24

	CUUAGUCACGUUAU GUAUGACGAGAAACU CGACUAGUUUCAAGCU AU															
>sfr-miR-279b-2	UGUAAGCAGCCUCCAC GGAAGAUAAAGCGAUA UUCUAGUAUCAAGUU GUUUACGUAUCUUGA CUAGAUUAUCACUUA UCCUCUGAGGUCGCUU UCG	96,00	19,79	25,00	20,83	33,33	40,63	58,33	1,33	0,95	-34,10	-34,10	0,02	5,19	-35,52	-0,87
>sfr-miR-10847	AAUGAGAAUGAUUGA UUGCGUGUCCCGCUCG CACCUGACAGGAACAA UUUUACUACACACGAU CAAUCAUUCUCGAC	78,00	17,95	29,49	25,64	25,64	43,59	55,13	0,87	0,70	-26,40	-22,90	0,16	5,75	-33,85	-0,78
>sfr-miR-10480	GCAAGUUUCCGUGGCG UGUUUAGCCGUCACAG UGACAUUUUUAACU ACAUA AUGUCACUUU GACAGCAAACCACAAC AUGGAAAACU AAC	92,00	17,39	32,61	22,83	26,09	40,22	58,70	0,80	0,76	-33,10	-33,10	0,05	6,62	-35,98	-0,89
>sfr-miR-2779-1	CCCAACCGAGCCAUA UAAAAUGAUCCGGCUC GAAGGACCAUUUUUAU GAGCGCUGCCGCGUUA AG	66,00	22,73	27,27	28,79	19,70	51,52	46,97	0,72	0,79	-20,00	-20,00	0,36	12,8 3	-30,30	-0,59
>sfr-miR-232b	GUCUCACAAUACCGAA CAAAGAGAAAAUGCA CCUUAACUUAGCAAUA UUUAAGGAUGUAUUU UGCUUGUUUGGAAAU GGAAUU	84,00	17,86	35,71	14,29	30,95	32,14	66,67	0,87	1,25	-20,50	-18,30	0,04	7,55	-24,40	-0,76
>sfr-miR-5394	AAUUUGUCGAUUUUC ACCAUCAGGAAAACUA CACUGUUACUUGGACA	93,00	17,20	37,63	17,20	26,88	34,41	64,52	0,71	1,00	-21,30	-21,30	0,41	3,62	-22,90	-0,67

	GAGCUAAAUUGUUAA AGCUAAACUGAUUUAU GAAAAGAGACAACCG															
>sfr-miR-6948	UGGUGGAGUUCAUGC GGAAAUAAACUUGCA GCUGGCCGUGAGCGAU AAUUAGAUAAAGUUCA GACAGGACUGCAAUU AAUUAACCUAUUGAA CAACCAGAU	101,00	23,76	33,66	16,83	24,75	40,59	58,42	0,74	1,41	-23,20	-23,20	0,10	8,32	-22,97	-0,57
>sfr-miR-83	CUCCUCAAGAUAGCAA UUACAAACUUAUUCA AAAGCAAUUCAUCCAA AACUGAAUUUAUGUG UGAAUGCGUUUGCAU CGUGACCUUGAGCGG	93,00	17,20	32,26	20,43	29,03	37,63	61,29	0,90	0,84	-19,40	-19,10	0,05	5,86	-20,86	-0,55
>sfr-miR-10469	GUGUGCAUGCAGUA AAUAAACACAUUUGU CAUCAAACGGGAUCCU GUAAAAACUGGACU GCCGUGUGAAAAGGC UGUAUGUAGGUAACU ACAGCAAAA	101,00	22,77	35,64	16,83	23,76	39,60	59,41	0,67	1,35	-23,80	-21,40	0,15	14,9 4	-23,56	-0,60
>sfr-miR-8468	CCGUAAACUAUCUAAAU AUCCAAUAAUGUAUU AUGCAAUAAGUAAUU UGAACAAGGCAAACU GCUUUGUUGUAUUUAU AAUAAUAGUUUUAGA UAGAUAUAA	101,00	12,87	39,60	10,89	35,64	23,76	75,25	0,90	1,18	-18,90	-14,20	0,01	23,5 2	-18,71	-0,79
>sfr-miR-2763	AAAUCUACAACGUUCU CCAAAGUAGUGAACG UAAUUUAUCGUGAGG UAAAUCAAUAGAUAU UAUGCUCAUUACUUU	95,00	16,84	34,74	13,68	33,68	30,53	68,42	0,97	1,23	-33,70	-33,70	0,17	6,67	-35,47	-1,16

	GGAUAUGUUGUACGA AUA															
>sfr-miR-2527	AUCCAAACACAUCAUC GCAUCCGUGUCAUUA CUCCUAAAAGAAGUGA GUUCUUCAGCGCAGGC AUCAGCAUCUGAUGG AUGUAGGUGUGUGAC U	95,00	22,11	27,37	23,16	26,32	45,26	53,68	0,96	0,95	-25,50	-25,50	0,14	16,6 2	-26,84	-0,59
>sfr-miR-95	UACGUAGUAACAGCA UUCAAUGGGUAUUUA CGAGUGUGAAAUAGC CACAAAAUAUGCUAU UACUACAUI	70,00	17,14	35,71	15,71	30,00	32,86	65,71	0,84	1,09	-20,50	-20,50	0,13	6,65	-29,29	-0,89
>sfr-miR-10498	GCGAGUGUCUCGCGUC UUUGGUCAACGUUCA ACACAGCAGUUCAGUU GCUGUUUUGGACAUI GGUGUAAGGCGCGGA UACUGUA	85,00	29,41	18,82	20,00	30,59	49,41	49,41	1,63	1,47	-38,30	-38,30	0,13	3,71	-45,06	-0,91
>sfr-miR-10460	UGAGCCAAUGUUCGU UAGUGAUGGAACUAI UCCCAAAGUUCAAUI CUUCCAUAAGCAUUGGC AGA	66,00	19,70	28,79	19,70	30,30	39,39	59,09	1,05	1,00	-22,80	-22,80	0,47	1,35	-34,55	-0,88
>sfr-miR-1415	ACCCACUGACAUIUGC AGGCGCUGUAAUUCU GCUUCUGAAACCAUUC AGCAUAUIACAGGUG CCUGCAAAGCCAUIUGU CC	81,00	18,52	24,69	28,40	27,16	46,91	51,85	1,10	0,65	-29,40	-29,40	0,06	7,21	-36,30	-0,77
>sfr-miR-2172	ACUCAUAUIUCUGUI UAUUGAUGCCUACAI GUAUAGUAUIACAAC UAUGUGUUGUACAAC AUAUAUACCUAUCA	93,00	12,90	32,26	15,05	38,71	27,96	70,97	1,20	0,86	-19,10	-17,20	0,04	12,1 8	-20,54	-0,73

	AUGUAUAGGAUAUGU CA															
>sfr-miR-7360	CUUGCCAACAAUAUUG UAUCACAGAUUCAUCC UAGGUCAUGCUAUA UACCUAGGAAGGAAA AAUAUGUACAUAUUU AUGGCCGU	86,00	16,28	33,72	18,60	30,23	34,88	63,95	0,90	0,88	-23,10	-23,10	0,14	6,02	-26,86	-0,77
>sfr-miR-7866	GAUCUGGGAUGAGAC AUUGCAGUUUCUUGU UAAAUUAUGAUAAU UUAAGUAAGUAUGC AUAUUUAAAAUAAU AUCAUUGUGAAUCGC AGGCA	96,00	18,75	35,42	9,38	35,42	28,13	70,83	1,00	2,00	-19,60	-19,30	0,06	9,57	-20,42	-0,73
>sfr-miR-8408	UUACGGGACGUCGGCC UGCACAAGUAUAUUG GUGAGUUAUGAGAUC UACUCCCCAUAAUUU AUUAUACUAGCUAGCC CGGCAAACGUACCGCC U	96,00	20,83	26,04	23,96	28,13	44,79	54,17	1,08	0,87	-25,70	-23,40	0,04	16,1 5	-26,77	-0,60
>sfr-miR-308	GACGACGCUUCCGCGC GCAGUAUUUAUACUUG UGAAUGUGUCCUAC UACAAUCACAGGAUA AUACUGCGAGUGGCG GCGUCCAC	85,00	24,71	23,53	25,88	24,71	50,59	48,24	1,05	0,95	-45,90	-45,90	0,51	1,87	-54,00	-1,07
>sfr-miR-3404	AGUAAGAGAUGGAAU CAUUUCUGCAAUAGA AUUACGAUAUAUUGC AGAACGUGGAGCUUCC AGACUCGUUCAG	74,00	22,97	32,43	16,22	27,03	39,19	59,46	0,83	1,42	-21,20	-21,20	0,56	2,46	-28,65	-0,73
>sfr-miR-12354	CCGCAUGUCUGUAAUA UCCAUUCGAGUACGGU ACUACUUGUAGUUUC	97,00	18,56	25,77	16,49	38,14	35,05	63,92	1,48	1,13	-20,40	-20,20	0,02	10,3 9	-21,03	-0,60

	AUUUAAGUUAUUUGA GUGAUCUCCAUAACUGA AUGAUUCAAAGAUG UGU															
>sfr-miR-10453-1	GGUGCACCGUGCAAU GUCAUUGAUACGCGU AUUCCAUUUUUUUUUA GGAGCGUAAUUUCGC UACUAUAGUUUCAUG GACGAUUGGCCAGUG CGUU	97,00	23,71	21,65	19,59	34,02	43,30	55,67	1,57	1,21	-21,40	-15,70	0,01	21,5 1	-22,06	-0,51
>sfr-miR-10453-2	GGUGCACCGCGUAAU GUCAUUGAUACGCGU AUUCCAUUUUUUUUUA GGAGCGUAAUUUCGC UUCUAUAGUUUCAUG GACGAUUGGCGCAGU GCGUU	97,00	24,74	20,62	18,56	35,05	43,30	55,67	1,70	1,33	-30,20	-29,80	0,07	6,48	-31,13	-0,72
>sfr-miR-3767	CUAAUCGAUACAAGCG AUGUAGAUAAUUGAG UAAAUAAACAUGUAG ACACACAGUCUGCACG CUUUACACAUUAUUU ACUAUCGACUGAUAU ACUAUCAA	101,00	13,86	37,62	18,81	28,71	32,67	66,34	0,76	0,74	-22,40	-20,50	0,04	9,72	-22,18	-0,68
>sfr-miR-3618	UUUCUAGACCACAUCU GUCUACAUUAAUGAA UCAUAAUGUAUCAUU UCAUGCUAUAGUAU GCGGUCUAGGCA	74,00	14,86	29,73	18,92	35,14	33,78	64,86	1,18	0,79	-20,30	-19,90	0,13	5,01	-27,43	-0,81
>sfr-miR-929	CCCAGCUCAUGGAUGU UAAAUUGACUCUAGU AGGGAGUCCGU AUGU ACGCGCGACUCCCUAA UCGAGUCAGGUUGAC UUCUUUGAGGCUAAC	93,00	24,73	22,58	23,66	27,96	48,39	50,54	1,24	1,05	-42,50	-42,50	0,17	3,33	-45,70	-0,94

>sfr-miR-10455	UGGUCUCGGAUGUCU UUCUAAAUCAAUCAA AUCUAUGAUGAGAUC UUGCUACAUAUCGACC UUCUUGGGUAUAGU UAGACGUCCGAGAUG U	93,00	20,43	24,73	19,35	34,41	39,78	59,14	1,39	1,06	-28,70	-27,50	0,13	11,1 1	-30,86	-0,78
>sfr-miR-iab-4	CGCGUCGCCUUCUGUA CGUAUACUGAAUGUA UCCUGAGUGCCUCAU CUUCCGGUAUACCUU CAGUAUACGUAACAG AAGAUGACACU	90,00	18,89	23,33	25,56	31,11	44,44	54,44	1,33	0,74	-38,30	-32,60	0,14	7,89	-42,56	-0,96
>sfr-miR-252a	GAAAAAACUAAGUCU UGUUCCUAAGUACUA GUGCCGCAGGAGUGU UACUAAUUUCUCCUGC UGCUUCAGUGCUUAUC AAUGAGUAUUUAGUG UUACA	98,00	19,39	26,53	18,37	34,69	37,76	61,22	1,31	1,06	-33,70	-33,70	0,28	3,43	-34,39	-0,91
>sfr-miR-3532	CUAAUUUUUGUAGC ACUAAUUAGCUGAUG UUGCACUGCAGCUGCU GAAAAACUUUCAGUG UAACAUACAAUGUAG UUGUACUGCGAAAUG AUAUG	97,00	19,59	30,93	15,46	32,99	35,05	63,92	1,07	1,27	-23,90	-23,00	0,04	10,3 3	-24,64	-0,70
>sfr-miR-2756	GUUGGGUUGGUAGCA UACCCUGUAGCUGCUA AGGGGCGCAGUUUAA UACCUCUUUGCUG CUACAUUGUAUGCUAC GGACCCUUG	88,00	26,14	17,05	25,00	30,68	51,14	47,73	1,80	1,05	-46,90	-45,90	0,27	2,82	-53,30	-1,04
>sfr-miR-10476	ACCGAAAGUUUACAUC GAGCGAUGGUUGGAA UCCAACUGUUGAUACA	89,00	21,35	26,97	17,98	32,58	39,33	59,55	1,21	1,19	-35,10	-35,10	0,35	4,02	-39,44	-1,00

	CAUUGGAUUUUAAAGU UCCAUCGCUAGUUGUG GACUAUCUUA															
>sfr-miR-6844	AGCAAAUAAUAGUU UUCUUUGUUUUUAAU UGGUUUUACGACAAG UUAUAUGCAAUUUGU CGUCUAUGAAAAACA UUCAAAACGUUUUU UUGCU	96,00	13,54	30,21	11,46	43,75	25,00	73,96	1,45	1,18	-23,30	-23,30	0,12	5,95	-24,27	-0,97
>sfr-miR-9789	UUGAAAAAGGCAUUC UAAGUUUAGCUUCCCA UCCAACUAGUUGGU AUUUAGAUUGAAGGC CAGUUAACGACAAU GUUUUUUAUA	87,00	17,24	31,03	14,94	35,63	32,18	66,67	1,15	1,15	-19,00	-18,60	0,20	5,36	-21,84	-0,68
>sfr-miR-133	CGUUGUCCGCUUUAGC UGGUUGACUUCGGGU CAAAUUGUAAUUUGG AUAUCAUUUGGUCCCC UUCAACCAGCUGUAGU UGACAUCU	87,00	21,84	18,39	21,84	36,78	43,68	55,17	2,00	1,00	-33,70	-33,40	0,34	4,40	-38,74	-0,89
>sfr-miR-133c	AGAUGUCAACUACAGC UGGUUGAAGGGGACC AAAUGAUAUCCAAAU UACAAUUUGACCCGAA GUCAACCAGCUAAAGC GGACAACG	87,00	21,84	36,78	21,84	18,39	43,68	55,17	0,50	1,00	-29,80	-29,80	0,32	3,80	-34,25	-0,78
>sfr-miR-12357	CUGAAAUGAAAAUAA UUCGGAUAAUUUACU GGCACCCUAACUUCUG CCAGUUCAUCCGAACA AUUCAUUUUC	74,00	12,16	31,08	24,32	31,08	36,49	62,16	1,00	0,50	-21,40	-21,40	0,28	3,69	-28,92	-0,79
>sfr-miR-10029	AGGUCUAUUUUCGUU UGGCAAAUUUGAAUA UUUGCUAAGAUCUGC	100,00	19,00	30,00	13,00	37,00	32,00	67,00	1,23	1,46	-20,60	-20,30	0,05	15,4 7	-20,60	-0,64

	UCUUAUACCAGUUCGU AGUAGGCCAAAUAAC AUUUUUGAAAGGUUA AGAGAUUU															
>sfr-miR-4331	GUUAGUCAAGAGUA UAGGCUGUGGCUGUG GUGUAGGUUAGUGUU AGUCUAUGGUCGUGU UGUGUGAAUGAUACG GCCUAAUGCUAUUUCA CUGAU	97,00	30,93	20,62	11,34	36,08	42,27	56,70	1,75	2,73	-28,40	-25,80	0,13	26,7 9	-29,28	-0,69
>sfr-miR-745	CAAAACUCUUGUCGGU UGCGGCUCAUCGUAUG GCAGUUUGCUGUGUA GUAUUGCCAGCUGCCU AGCGAAGGGCAACAA AUGACUGAGUUGAU	93,00	27,96	22,58	20,43	27,96	48,39	50,54	1,24	1,37	-37,90	-36,10	0,20	4,12	-40,75	-0,84
>sfr-miR-96	UUUCACUAAUCUUAU UCAUUGGUACACAUA GCUUGGCACUGGCGGA AACGGACUCAGCUAAG CUAUGUUUUUAUUAU GGAAAGAUACGUGCU A	94,00	20,21	27,66	18,09	32,98	38,30	60,64	1,19	1,12	-26,90	-25,80	0,17	4,91	-28,62	-0,75
>sfr-miR-10507-1	GGAGAACCGUCCAAGU UUAACUGUAGUGGU CUUUUGAGGAAAGAA CCACAGCAGUUUGAGC GGACUUGUUCAGA	76,00	27,63	27,63	17,11	26,32	44,74	53,95	0,95	1,62	-27,50	-27,50	0,23	4,28	-36,18	-0,81
>sfr-miR-375	GACAUGACAGCGAGAC AACCCGAGCGGAUUGA GCAAACUAUUUGUCU GUUAGCAAGUUUGUU CGCCCCGGCUCGAGUC GUCUGGCGUCAUACC	94,00	26,60	22,34	26,60	23,40	53,19	45,74	1,05	1,00	-39,30	-39,30	0,17	4,38	-41,81	-0,79

>sfr-miR-10507-2	GUCUCUUUUAAAAGUCC AGACAAAAUCAAUUA AAGAUGUCGGAGAAC CGUGCAAGUUUAUUC UGUAGUGGUCUUUAG AGGAAAG	84,00	22,62	32,14	14,29	29,76	36,90	61,90	0,93	1,58	-20,00	-17,20	0,15	6,98	-23,81	-0,65
>sfr-let7f	GCUUACAUAUUUUGA AAUGGUUUUGGAGCU ACAGUAGGCUUUGGU UGUAGAGCUAUACAG UCUACUGUAGACCUGA AGACAAAAUCAAAU AUUUUUUU	100,00	19,00	34,00	13,00	33,00	32,00	67,00	0,97	1,46	-28,80	-27,30	0,07	10,4 5	-28,80	-0,90
>sfr-miR-2796	AUUAAGUACAGUCAG UAGGGGUUUUCUUUCG GCCUUCAGCUGUUGAU AGACUGUAGGCCGGCG GAAACUACUUGCCGAC UCUACUUCUA	89,00	24,72	21,35	22,47	30,34	47,19	51,69	1,42	1,10	-39,60	-39,60	0,28	2,14	-44,49	-0,94
>sfr-miR-10462-1	GUAAAAUUUAAGCAG UCGGGACCCAUUCUAG AAAGCAAGCCGUAUG UGCCCUCACAAAGUCU UUUAUUUUAGAAUGG GUCCCGACUGCUUAAA UUUUAC	100,00	19,00	30,00	21,00	29,00	40,00	59,00	0,97	0,90	-55,50	-54,50	0,36	2,81	-55,50	-1,39
>sfr-miR-10471-2	AGCUAAAAAAAGUAG ACCGUUCAGUACCUUU AGUACGAGUUUAUUU UACGUUUAAACCGGUGC UCUGAUUGGUUAGUU UUUUUAUAC	87,00	18,39	26,44	14,94	39,08	33,33	65,52	1,48	1,23	-18,90	-15,20	0,02	12,0 3	-21,72	-0,65
>sfr-miR-10462-2	AGAAAAUUUAAGCAG UCGGGACCCAUUCAAG AAAGCCAGCCGUAUGU GCCCUUACUAAGUCUU	100,00	19,00	31,00	21,00	28,00	40,00	59,00	0,90	0,90	-42,50	-37,50	0,06	13,9 4	-42,50	-1,06

	CAUAUUUAGAAUGGG UCCCGACUGCUUAAA UUUUA															
>sfr-miR-10462-3	GUCAUCGUAAAAUUU AAGCAAUCGGGACCCA UUCUAGAAAGCCAGCC AAAUAUGUACCCAUA UUUAGAAUGGGUCCC GACUGCUUAAAUUUU ACGAUGAC	101,00	17,82	32,67	21,78	26,73	39,60	59,41	0,82	0,82	-57,90	-57,60	0,22	4,60	-57,33	-1,45
>sfr-miR-10462-4	AAGUCAUCGUAAAAU UUAAGCAGUCGGGACC CAUUCUAGAAAGCCCU CACUAAGUCUAUAUU UAGAAUGGGUCCCGAC UGC UUAAAUAUUACG AUGACUU	101,00	17,82	31,68	20,79	28,71	38,61	60,40	0,91	0,86	-62,40	-62,10	0,14	3,64	-61,78	-1,60
>sfr-miR-10462-5	AGAAUACAAGAAAAA UUUAAGUAGUCGGGA CCCAUUCUAGAAAGAC AUAUUCAUAAUAUU UAGAAUGGGUCCCGAC UGC UUAAAUUUUUCU UGUAUUCU	101,00	15,84	35,64	14,85	32,67	30,69	68,32	0,92	1,07	-63,20	-63,20	0,46	1,80	-62,57	-2,04
>sfr-miR-10462-6	GUAAAAUUUAAGCAG UCGGGACCCAUUCUAA AUAUAAAGACUGUGU GAGGACACAUACGGCU UGC UUCUAGAAUGG GUCCCGACUGCUAAA UUUUAC	100,00	21,00	30,00	19,00	29,00	40,00	59,00	0,97	1,11	-61,10	-61,10	0,71	1,06	-61,10	-1,53
>sfr-miR-3894	GCGGUUUUAUGGAAU UUGGUUUUUGACAAU CUUUGGAACUAUCUCA UUUUCAGAAUGACCA	90,00	17,78	33,33	15,56	32,22	33,33	65,56	0,97	1,14	-22,90	-17,20	0,08	13,2 3	-25,44	-0,76

	AAGAUAGCCAAAGCCA UUAUAAAACGAA															
>sfr-miR-5838	GAAAUUGUUAUUGGU CUUUGUGAGUAUUUU CGGUUUUGAGUUUCCG AAGCAAAAUAACUUG CAUAUAAUAAUAAUA AUCGU	81,00	18,52	30,86	9,88	39,51	28,40	70,37	1,28	1,88	-22,40	-16,70	0,08	14,2 6	-27,65	-0,97
>sfr-miR-927a	GGGUUUUGGUUUUAGA AUUCCUACGCUUUACC GUUGCCAAGCAGUGU UGGGCAAAGCGUUUG GAUUCUAAAACUAAA UAG	80,00	25,00	25,00	16,25	32,50	41,25	57,50	1,30	1,54	-32,50	-26,00	0,14	10,5 2	-40,63	-0,98
>sfr-miR-193	UAUGUCCACCACGGCC AGGGUCUUGGCGGUC UGGUCGGUGUAUGAU UCUCUUACUGGCCUGC UAAGUCCCAAGCCUUG GUGGACUUU	88,00	28,41	13,64	26,14	30,68	54,55	44,32	2,25	1,09	-45,60	-45,60	0,45	4,03	-51,82	-0,95
>sfr-miR-927c	CUAUUUAGUUUUAGA AUCCAAACGCUUUGCC CAACACUGCUUGGCAA CGGUAAAAGCGUAGGA AUUCUAAAACCAAACC C	80,00	16,25	32,50	25,00	25,00	41,25	57,50	0,77	0,65	-26,20	-25,30	0,07	6,46	-32,75	-0,79
>sfr-miR-138	UAAAUUAGACAUGCG AUAGUUUCCGAUACA AAAUCAUUGCUAGCU GGUGUUGUGAAUGCU UUUUUAAUCAGGAAA UCGCGUGUUGUAAUG AU	93,00	21,51	29,03	12,90	35,48	34,41	64,52	1,22	1,67	-23,80	-22,60	0,04	11,0 7	-25,59	-0,74
>sfr-miR-1651	UCACAUAAGCUUAUU GGGUACUUUAUAUGAA GGUGGUAAAACAGGC	99,00	20,20	33,33	18,18	27,27	38,38	60,61	0,82	1,11	-26,30	-25,40	0,10	5,85	-26,57	-0,69

	GCUAAGACUGAUAAA AUCCACUAUCACUAAAG UACUGGAAUCGGCUU ACUGACU															
>sfr-miR-3885	CCAAGGCGGCGGCGGC GGUAAGUACUCAUUA UAUUGUACUCAAUAA GUACUCAUUAUUUAA GUAGCAGUAUGAAGA UUUACCGGGGCCCCGG CUCAA	98,00	24,49	27,55	21,43	25,51	45,92	53,06	0,93	1,14	-33,30	-30,50	0,09	10,1 4	-33,98	-0,74
>sfr-miR-9-1	GUCGUAGAUGGCACU UAACUCCGGUAACCUA GCUUUAUGAUGUCAG AAUACUCAUACAGCUA GAUAACCAAAGAUAA UUAACCGAUCUACUUU	94,00	15,96	32,98	21,28	28,72	37,23	61,70	0,87	0,75	-23,80	-23,80	0,20	5,48	-25,32	-0,68
>sfr-miR-1798	UGGCUGAAUCGAUUU UUAUGUGGUUUUCAG AAGUGUUUUUGUGUC UAGGAGCAGGUUUUA GUAUAUUAACCGUCA UAAAAAAGAGGUUC AGGAC	96,00	25,00	27,08	10,42	36,46	35,42	63,54	1,35	2,40	-21,10	-19,80	0,05	17,3 2	-21,98	-0,62
>sfr-miR-9c	ACGCCGCGGCUCGUCU CUUUUGGUAUCCUAGC UGUAGGCGUGUCACGC AAACCCUAAAGUUAU GGUACCGAAGUUACG AGUCGCGGUAU	90,00	26,67	20,00	25,56	26,67	52,22	46,67	1,33	1,04	-45,10	-45,10	0,10	6,32	-50,11	-0,96
>sfr-miR-9-2	AAAGUAGAUCGGUUA AUUAUCUUUGGUUAU CUAGCUGUAUGAGUA UUCUGACAUCAUAAA GCUAGGUUACCGGAG	94,00	21,28	28,72	15,96	32,98	37,23	61,70	1,15	1,33	-37,00	-37,00	0,38	2,38	-39,36	-1,06

	UUAAGUGCCAUCUACG AC															
>sfr-miR-79	GACGCGCGGUUCGGCU CUUUGGCGAUUUAGC UCCGUGACGUGAAAA AAGCUCAUAAAGCUA GAUUACCAAAGCACCG UACCGUCGCACG	90,00	25,56	25,56	26,67	21,11	52,22	46,67	0,83	0,96	-38,90	-38,90	0,65	1,09	-43,22	-0,83
>sfr-miR-9b	CGUGCGACGGUACGGU GCUUUGGUAUUCUAG CUUUAUGAGCUUUUU UCACGUCACGGAGCUA AAUCGCCAAAGAGCCG AACCGCGCGUC	90,00	26,67	21,11	25,56	25,56	52,22	46,67	1,21	1,04	-37,60	-37,60	0,16	3,18	-41,78	-0,80
>sfr-miR-2755	UUAUAUGAGCUUGGU GACUCAAGGUGGCCUA GCAGCGUGUUUUCUCU UAAGGUUUGCACCCUG UCAGACCAUACUUGUU UCAACAUGUCUCUAAU C	97,00	20,62	20,62	22,68	35,05	43,30	55,67	1,70	0,91	-26,40	-25,30	0,01	5,12	-27,22	-0,63
>sfr-miR-513d	ACACACACAUUUGUAG UAGUUCACAACGAGG UGUCAUAACUUGUGU UACAACUUUAUUCGU UUCGCACCAUGUAUGU ACAGUACACAUGUGU GGUG	97,00	19,59	26,80	19,59	32,99	39,18	59,79	1,23	1,00	-23,60	-20,40	0,06	22,8 1	-24,33	-0,62
>sfr-miR-7184	AGUCAUGUGAGCCUUC UGCUGGAGCAGGUCG GCUCGCGUGUCAUCGU GCUGGUGAGCUGAUCC AGUAGGGCUAAUGUA A	80,00	33,75	17,50	21,25	26,25	55,00	43,75	1,50	1,59	-33,60	-15,38	0,07	29,0 1	-42,00	-0,76

>sfr-miR-10590	CAAAGUCGCUAGCAAC AGCUAGUAAUACAUA UUAUUUAAAACUUGAA CUAUUUAAUUUAAAAG UCCUCAUGUGUUUACC UGGACAGUCUACUAGC GUCUCUC	101,00	13,86	30,69	20,79	33,66	34,65	64,36	1,10	0,67	-19,00	-14,90	0,07	$\frac{12,8}{9}$	-18,81	-0,54
>sfr-miR-10587	UCAGUUCGUUAGGUA ACUUCUGAACCAAGUU CGGUCGAGUUUCUUUC GAACCUCGUAAUUUG GACCGGAACUCUCAGA AGUAAGCGCCUCA AUG AACAAA	101,00	20,79	27,72	22,77	27,72	43,56	55,45	1,00	0,91	-31,70	-30,30	0,06	$\frac{12,5}{1}$	-31,39	-0,72
>sfr-miR-81c	GCGACAUCUAUCGUGA GAUCAAUUAGACGA AUGUACGUCUGAUGU UGUUUCAUGCAACAU AGUAUGUUGU	72,00	22,22	27,78	15,28	33,33	37,50	61,11	1,20	1,45	-25,50	-25,50	0,39	2,62	-35,42	-0,94
>sfr-miR-10-1	UAGUGCCCUACAUCUA CCCUGUAGAUCCGAAU UUGUUUGAAGUGAGG CGACAAAUUCGGUUCU AGAGAGGUUUGUGUG GUGCAUGC	87,00	27,59	21,84	18,39	31,03	45,98	52,87	1,42	1,50	-37,00	-36,90	0,16	4,16	-42,53	-0,92
>sfr-miR-135a	AUUAUGGCUUUCAUCC UAUGUGGGCUCAUGU UUGAUGGGACAACGU CACACGUUCUUUACGA AUGAAUGACGCACAG AAUGAAACCAUUGA	92,00	21,74	28,26	19,57	29,35	41,30	57,61	1,04	1,11	-23,90	-23,90	0,21	$\frac{10,8}{1}$	-25,98	-0,63
>sfr-miR-993a-1	GCACAUUCGUGAUCUA CCCUGUAGAUCGGGGC UUUUGUGACUUACAC AUCAGAAGCUCGUUUC	86,00	23,26	22,09	24,42	29,07	47,67	51,16	1,32	0,95	-41,30	-41,30	0,15	5,44	-48,02	-1,01

	UACAGGUAGCUCACGA UUGGGA															
>sfr-miR-993a-2	AUUCGUGAUCUACCCU GUAGAUCCGGGCUUU UGUAGUUUAGUUCAC AUCAGAAGCUCGUCUC UACAGGUAUCUCACGG CU	81,00	20,99	19,75	24,69	33,33	45,68	53,09	1,69	0,85	-33,10	-33,10	0,08	3,83	-40,86	-0,89
>sfr-miR-2318	UAAAU AUGAAAUAU UAAUACACGUGCGUA ACGAAUUUCGGAGUA AAUAAUGUCUCGCCUG CUCUAUAUCCGCCGUG UAUGAUGAAUAUUC AUAUCAU	100,00	16,00	32,00	17,00	34,00	33,00	66,00	1,06	0,94	-20,00	-19,40	0,03	11,5 3	-20,00	-0,61
>sfr-miR-10484-1	CCUAAGAAAAGGAGG AUCCUCCGACCAGGCG AGGUGAUCAUGCCUG GCGGGCUUUCUGCCCU AUUUCUUUAA	73,00	26,03	21,92	26,03	24,66	52,05	46,58	1,13	1,00	-27,10	-17,90	0,07	12,7 5	-37,12	-0,71
>sfr-miR-276b	UACCCAAGAGCACGGU AUGAAGUCCUACAG UGGUAGAAUACGUAG GAACUCUAUACCUCGC UAGUGGCAA	72,00	23,61	30,56	22,22	22,22	45,83	52,78	0,73	1,06	-29,50	-29,50	0,70	1,12	-40,97	-0,89
>sfr-miR-10484-2	GUAAGUAAAGUUCCC UAAGAAAAGCAGGAU CCUCCGACGAGGCGAG GUGAUCAUGUCUGGCC GGCUUUCUGCCCUGCC UUUCUGUUGGGAUUU UCUAUC	100,00	26,00	20,00	24,00	29,00	50,00	49,00	1,45	1,08	-30,10	-25,50	0,04	16,3 9	-30,10	-0,60
>sfr-miR-276	UUGCCACUAGCGAGGU AUAGAGUCCUACGU AUUCUACCACUGUAGG	72,00	22,22	22,22	23,61	30,56	45,83	52,78	1,38	0,94	-31,00	-31,00	0,24	5,45	-43,06	-0,94

	AACUUCAUACCGUGCU CUUGGGUA															
>sfr-miR-10-2	UUAGCUCAUACGUGU AACCCUGUAGAUCCGA ACUUUAUCUAUUAAA CUUGAAUAACGUGCG GACCAACAGGAUUACA CUUGUGGGGCACG	90,00	20,00	28,89	22,22	27,78	42,22	56,67	0,96	0,90	-32,10	-31,00	0,15	3,93	-35,67	-0,84
>sfr-miR-7675	CUCUAGGUAUCUACAA UCAAUGCAUGAAGUA GAAUGUAGAUGAAAU GCCACAAUUGUUUCAU UUGUUCGUUCUAUCA GCAUUAUAGCAAUAA GCUAAGA	100,00	17,00	34,00	16,00	32,00	33,00	66,00	0,94	1,06	-21,50	-21,50	0,05	7,95	-21,50	-0,65
>sfr-miR-277	UCCGGAGUUCGUGCCA GGAGUGCGUUUGCAA AGCACGCUACAAGUUU GCGUUAAGGCAAAGU UGACACUGUAAAUGC ACUAUCUGGUACGACA CCCCGGA	101,00	26,73	25,74	23,76	22,77	50,50	48,51	0,88	1,13	-48,30	-48,30	0,20	4,52	-47,82	-0,95
>sfr-miR-316	AAGCAUCAAGCAUAA GCAAGAAGAAGCAGA AGAGAGAAGCAACAG UUCUCCAUAUUUUUG UCUUUUUCCGCUUUGC UGCUGAUAAUUAUGA UC	94,00	19,15	35,11	18,09	26,60	37,23	61,70	0,76	1,06	-20,70	-18,10	0,06	11,6 9	-22,02	-0,59
>sfr-miR-4648	UAAGUGUUAGUGACG CCCAAUGUGGGACUGC AAAUGUUCGCGAGGCC CAGUUAAGGCGCCAAC ACCAU	69,00	28,99	24,64	24,64	20,29	53,62	44,93	0,82	1,18	-24,20	-23,30	0,17	5,58	-35,07	-0,65

>sfr-miR-3338	AGAAGAGUGUUUCAU AGCAAACACAGUAGU GUGCAAUGCUAAAGU CCACCAUGCAUGUACU UACUUUGUUUGUUCU GAAAUACUCUAAAC	90,00	17,78	31,11	18,89	31,11	36,67	62,22	1,00	0,94	-35,90	-33,50	0,31	1,90	-39,89	-1,09
>sfr-miR-965	AUUGCUGUGUAAACUG AACGGGAGAAGUUGU ACCGCUAU AUGUUUU UAAUAAGGUCAUAAG CGUAUAGCUUUUCCCC UUGGUUAUGCAAGCC AG	94,00	23,40	25,53	17,02	32,98	40,43	58,51	1,29	1,38	-37,30	-37,30	0,07	7,41	-39,68	-0,98
>sfr-miR-87	AAGAAAGUACGGUAC ACUGAUGGGCCUGAA UUGUUGCUCGAACCGU UGUUUUGAUC AAGGU GAGCAAACUUCAGG UGUGUUAGUGCCGAC UUUUCA	98,00	26,53	24,49	17,35	30,61	43,88	55,10	1,25	1,53	-40,90	-40,90	0,06	8,43	-41,73	-0,95
>sfr-miR-9388	GAGGUUAACAUUGUA GUAACGGUGCAAGUA CCUGUGUAUGUAUGU AUGUAUGUACAUACG UAGGUUCUCCACACGG CUCACAAGCUUACACA UA	95,00	22,11	28,42	18,95	29,47	41,05	57,89	1,04	1,17	-25,50	-24,50	0,03	11,7 5	-26,84	-0,65
>sfr-miR-10458	UGUGUGUGCUGCGCCG GGAUACGUGUUUACC GCAUGUGUGAUGUGU CUGCCUAAAACAUGUA UCCGGCUCAGCAACCC U	80,00	27,50	17,50	25,00	28,75	52,50	46,25	1,64	1,10	-31,20	-31,20	0,04	9,95	-39,00	-0,74
>sfr-miR-263a-1	AACUCGCACCAUGCAC AGGCAAUGGCACUGG AAGAAUUCACGGGUU	100,00	26,00	23,00	24,00	26,00	50,00	49,00	1,13	1,08	-41,20	-41,20	0,12	7,61	-41,20	-0,82

	ACAUUUUAAUUAGUCCC GUGGUCUCUUAGUGG CAUCACUGGUGCUGGG CGAUAC															
>sfr-miR-263b-1	AUCAGACCAUGUUGA GCCUUGGCACUGGGAG AAUUCACAGGAGUUC UGAAUAUACCGUGAA UUUCCUGAUGCCUAG CUUAAUGCGGUCUUU U	94,00	23,40	23,40	20,21	31,91	43,62	55,32	1,36	1,16	-35,80	-35,80	0,07	5,02	-38,09	-0,87
>sfr-miR-263a-2	AACUCGCACCAUGCAC AGGCAAUGGCACUGG AAGAAUUCACGGGUU ACAUUUUAAUUAGUCC GUGGUCUCUUAGUGG CAUCACUGGUGCUGGG CGAUAC	99,00	26,26	23,23	23,23	26,26	49,49	49,49	1,13	1,13	-39,40	-39,40	0,15	6,60	-39,80	-0,80
>sfr-miR-263b-3	AUCAGACCAUGUUGA GCCUUGGCACUGGGAG AAUUCACAGGAGUUC UGAAUAUACCCGUGA AUUCCUGAUGCCUUA GCCUAAUGCGGUCUUU U	95,00	23,16	23,16	22,11	30,53	45,26	53,68	1,32	1,05	-32,70	-32,70	0,07	7,58	-34,42	-0,76
>sfr-miR-210	UGGUGAAAUGAAGCC CAAAGCAGCUGCUGGC CACUGCACAAGAUUAG UUAGAACUAUACUCU UGUGCGUGUGACAGC GGCUAUUGCGGGCUA UUCAUUG	100,00	27,00	25,00	21,00	26,00	48,00	51,00	1,04	1,29	-44,00	-43,40	0,10	10,0 2	-44,00	-0,92
>sfr-miR-3286	GCUUGAAGCCCCAAAC AACUAUUGGGCACUGC ACAGGACUAGUUACCC GAAUAUACUCUUGUG	97,00	22,68	25,77	22,68	27,84	45,36	53,61	1,08	1,00	-40,60	-40,40	0,07	7,05	-41,86	-0,92

	CGUGUUCUAAUAGUU GUUGUCGGAGCUUCAC AG																
>sfr-miR-317	GGGCUAUCGAACGACC GGGUGCCACGCUGUGC UCUCUCGGGAAUUA UGCAGAGUGAACACA GCUGGUGGUAUCUCA GUUGUUCCCUAGUUC	93,00	29,03	19,35	24,73	25,81	53,76	45,16	1,33	1,17	-39,80	-39,70	0,28	4,30	-42,80	-0,80	
>sfr-miR-210b	GACAAUGAAUAGCCCG CAAUAGCCGCUGUCAC ACGCACAAGAGUAUA GUUCUAAACUAAUCUU GUGCAGUGGCCAGCAG CUGCUUUGGGCUUCAU UUCA	99,00	22,22	26,26	25,25	25,25	47,47	51,52	0,96	0,88	-41,50	-41,50	0,07	7,05	-41,92	-0,88	
>sfr-miR-750	CUAGACCUCACGUCUG AGUUGGACAGGGGAU CUUGACAGUUUGAGC UAUGUGCUGCCAGAUC UAUCUUUCCAGCUCAC GCGUGAAGUCGCU	92,00	26,09	19,57	25,00	28,26	51,09	47,83	1,44	1,04	-39,90	-39,30	0,14	3,30	-43,37	-0,85	
>sfr-miR-10467	GUAUCGAUUGCUCGGC UGUAUUGAUCGAUCA CUAUUCUCGUGAGAU GCACUCGAGCGAUGAA UAGAGCCGAGCGAUCG AAGC	83,00	27,71	24,10	21,69	25,30	49,40	49,40	1,05	1,28	-38,60	-35,00	0,08	8,73	-46,51	-0,94	
>sfr-miR-932	GGAGGGUGUCGAGGC CUCAAUUCCGUAGUGC AUUGCAGUGCAUCCUG CCUCCUGCAAGCAGUG CGGAAGUGAGGCUGA GACACCCCAU	89,00	31,46	20,22	26,97	20,22	58,43	40,45	1,00	1,17	-55,20	-55,20	0,43	1,47	-62,02	-1,06	
>sfr-miR-10505-1	AGCAUCGACUCGCCAG UUCAAAAGUAUAGGG	88,00	23,86	27,27	19,32	28,41	43,18	55,68	1,04	1,24	-21,50	-16,40	0,09	16,0 1	-24,43	-0,57	

	UUAGAAACUUGGCUC UACACGGGAUCUGAU UACUUUUUGACAGUG CGAGCUAU AUG															
>sfr-miR-5700	CGGGUCGCAAGUAUA UAGAUAAUAAGGUGU ACAUA AUGCAUUAAA UUAUGGUUAUACAUG UAUAAUUUAUGUGCU UCUACUAA	84,00	17,86	34,52	10,71	35,71	28,57	70,24	1,03	1,67	-19,40	-18,00	0,18	10,6 6	-23,10	-0,81
>sfr-miR-6830	CGACCUCUCACUGUCG ACUUUUGUAGACUCU GGGCUUCUGCAAAGA UGAAAUGUCGCCAAG GAAGGAGGCUGCUAA AGGAGAGUGAAAGGG AC	94,00	29,79	27,66	20,21	21,28	50,00	48,94	0,77	1,47	-27,60	-27,60	0,16	8,90	-29,36	-0,59
>sfr-miR-16c	ACCGGUCACAAUCCA GACUCCGUGCUACUGC UGAGAAAUUUUCGAA AAACCGAAAAAACUC CAGUAUUACUUUGCU UGGAAUUGAACCCGA	94,00	17,02	32,98	24,47	24,47	41,49	57,45	0,74	0,70	-27,60	-27,00	0,39	3,05	-29,36	-0,71
>sfr-miR-10501-1	CAUCAAAAACCUAGAC CACUCCAAUGGUCGU AUUGACUUGAAAUUU GGCAUGGAGGUAGGU CUUUGGGU	71,00	23,94	26,76	18,31	29,58	42,25	56,34	1,11	1,31	-22,90	-22,90	0,21	5,29	-32,25	-0,76
>sfr-miR-10501-2	ACUCAUAAAAACCUA GACCACUCCAAUGGU CGUAUUGACUUGAAA UUUGGCAUGGAGGUA GGUCUUUAUGUCA	76,00	19,74	28,95	19,74	30,26	39,47	59,21	1,05	1,00	-19,30	-19,30	0,29	7,94	-25,39	-0,64
>sfr-miR-10501-3	UCACUCAUAAAAACC UAGACCACUCCAAUG GUCGU AUUGACUUGA	82,00	21,95	28,05	19,51	29,27	41,46	57,32	1,04	1,13	-21,00	-19,30	0,14	10,5 7	-25,61	-0,62

	AAUUUGGCAUGGAGG UAGGUCUUUAGAUC GGGU															
>sfr-miR-10501-4	CAUCAAAAACCUAGAC CACUCCAAUGGUCGU AUUGACUUGAAAUUU GGCAUGGAGGUAGGU CUUUGGUC	71,00	22,54	26,76	19,72	29,58	42,25	56,34	1,11	1,14	-22,80	-22,80	0,40	3,59	-32,11	-0,76
>sfr-miR-10501-5	AAAACCUAGACCACUU CCAAUGGUCAUAUUG ACUUGAAAUUUGGCA UGGAGGUAGGUCUUU AGGUCAA	69,00	21,74	30,43	17,39	28,99	39,13	59,42	0,95	1,25	-24,10	-24,10	0,21	3,99	-34,93	-0,89
>sfr-miR-10456	GUUAAUUCGCAUGUA AACUUCAUAUGUGCG AUGCAGGAUAAUUUAU GACGGCAUCCGUGGAU UUGCUCGUAGAUAGU CUAUGUGCGACUUCUG	93,00	24,73	22,58	17,20	34,41	41,94	56,99	1,52	1,44	-25,10	-21,40	0,05	20,9 5	-26,99	-0,64
>sfr-miR-10495-2	CUGCAUCAACCGGGGA UUGUCCUUUGUGUAC UCCAUAUGUGCAUACA GGGAUAAUCGCCGGCU GAUGUCU	71,00	25,35	19,72	23,94	29,58	49,30	49,30	1,50	1,06	-40,00	-40,00	0,71	0,77	-56,34	-1,14
>sfr-miR-10495-3	CACAAUAGUUGUGCA UCAACCGGGGAUUGUC CUUUGGUGUAUUUCC AUAGUGCAUACAGGG AUAUUCGCCGGUUGA UGUCCCAUUACAUUUA G	94,00	23,40	23,40	20,21	31,91	43,62	55,32	1,36	1,16	-41,70	-40,70	0,05	8,89	-44,36	-1,02
>sfr-miR-10495-4	CUUACAUCAGCCGGCG AUUGUCCUUUGGUGU ACUCCAUAUGUGCAUA CAGGGAUAAUCGCCGG CUGAUGUCCC	74,00	24,32	18,92	27,03	28,38	51,35	47,30	1,50	0,90	-48,70	-48,70	0,31	2,40	-65,81	-1,28

>sfr-miR-10495-5	CUGCAUCAACCGGGGA UUGUCCUUUGGUGUA CUUCCAUAUGUGCAUAC AGGAUUAUUCGCCGAC UGAUGUCC	72,00	23,61	20,83	25,00	29,17	48,61	50,00	1,40	0,94	-30,20	-30,20	0,24	2,65	-41,94	-0,86
>sfr-miR-305	CAGCCGCCGCACGCCC AUUGUACUUCAUCAG GUGCUCUGGUGAUGA UGGUUCCAGGCGCUUG UUGGAGUACACUUACC GUGUCGGCGGUAG	92,00	30,43	15,22	27,17	26,09	57,61	41,30	1,71	1,12	-49,90	-49,90	0,53	1,83	-54,24	-0,94
>sfr-miR-1000	CUCGCGCGCCACGACC AUUUUGUCCUGUCACA GCAGUACGUGCACUGU UACUGCUGCGUCCGGA CAAACGGGCCGUGGCG UCGCACU	88,00	27,27	17,05	34,09	20,45	61,36	37,50	1,20	0,80	-44,70	-41,70	0,20	3,22	-50,80	-0,83
>sfr-miR-10454a-1	AGUACCCAGGGGCCUU AGUCUGCUAUUACAA UUGUGUCAGAAUGGU CGAUCAUUGAGCACAA UUGUAAUAUCAGACU AAGGCCCCAGGUGGAA	94,00	24,47	28,72	20,21	25,53	44,68	54,26	0,89	1,21	-56,70	-56,70	0,16	4,14	-60,32	-1,35
>sfr-miR-4819	UGAUUAUUUACUUUU AUUAUUACGUCCAUC UGACGCGCUCGCGGCA CUCGCGUGUGAUUUA ACUUGGACAAAAUGG AUAUCAGUAGCAAUA AAAC	97,00	18,56	27,84	20,62	31,96	39,18	59,79	1,15	0,90	-21,70	-20,00	0,02	8,88	-22,37	-0,57
>sfr-miR-10454a-2	GAACAGGGGCCUUAG UCUGCUAUUACAAUU GUGUCAGAAUGAUC GUGAUCGACCAUUCUG ACACAAUUGUAAUAG	96,00	22,92	29,17	20,83	26,04	43,75	55,21	0,89	1,10	-73,30	-73,30	0,82	1,09	-76,35	-1,75

	CAGACUAAGGCCCCUG GUA															
>sfr-miR-10454a-3	GGGGCCUUAUUCUGCU AUUACAAUUGUGUCA GAAUGGUCGAUCACU GAGAUGACGAUGAUC AAACAUUGUGACACA AUUGUAAUAGCAGAC UAAGGCCCC	101,00	22,77	29,70	19,80	26,73	42,57	56,44	0,90	1,15	-63,00	-63,00	0,72	0,88	-62,38	-1,47
>sfr-miR-10454a-4	GGCCUUAGUCUGCUAU UACAAUUGUGUCACA AUGGUCGAUUACUGA GCAGUGCAAGAGGGA UCGACCAUUCUGACAC AAUUGUAAUAGCAGA CUAAGGCC	101,00	23,76	28,71	20,79	25,74	44,55	54,46	0,90	1,14	-69,40	-69,40	0,28	2,57	-68,71	-1,54
>sfr-miR-10454a-5	GGGCCUUAGUCUGCUA UUACAAUUGUGUCAG AAUGGUCAAUCAUUC AACUGAUCAAUGAUC GACCAUUGUGACACAA UUGUAAUAGCAGACU AAGGCC	100,00	20,00	31,00	21,00	27,00	41,00	58,00	0,87	0,95	-71,20	-71,20	0,78	0,79	-71,20	-1,74
>sfr-miR-10454a-6	CCUUAGUCUGCUAUUA CAAUUGUGUCGGAU GGUCGGUCCGUCCCUC UCGCACUGCUCAGUGA UCGACUAUUCUGACAC AAUUGCAAUAGCAGA CUAAAG	101,00	20,79	23,76	25,74	28,71	46,53	52,48	1,21	0,81	-55,10	-54,00	0,11	5,63	-54,55	-1,17
>sfr-miR-10454a-7	UUUUGAUCGCCAUGG UAAAACAAUUGUGCC UUUGAACAGGGGCCU UAGUCUGCUAUUACA AUUGUGUCAGAAUGG UCGAUCACUG	86,00	23,26	24,42	18,60	32,56	41,86	56,98	1,33	1,25	-24,90	-24,90	0,23	5,21	-28,95	-0,69

>sfr-miR-10454a-8	UGUCCAGUCAUCUUGA CUACUACACUGGGGCC UUAGUCUGCUAUUAC AAUUGUGUCAGAAUG GUCGAUCACUGAGCAG UGCAAGAAGGACGGA GC	96,00	26,04	25,00	21,88	26,04	47,92	51,04	1,04	1,19	-25,20	-25,20	0,23	5,73	-26,25	-0,55
>sfr-miR-10454a-9	CAUAUUCUGAACUAG AAUAUUUAUAGUGC CCUUCUAGGGCACAUAU UAUCUGGGGCCUUAAG UCUGCUAUUACAAUU GUGUCAGAAUGGU	90,00	20,00	25,56	17,78	35,56	37,78	61,11	1,39	1,13	-22,20	-12,00	0,04	25,9 1	-24,67	-0,65
>sfr-miR-10454a-10	AGUGACGAUAUGACA CAAGUGUACAACUACU AACCACUGGCCUGUAC UCCAGGGGCCUUAAGUC UGCUAUUACAAUUGU GUAAAAAUGGUCGAU	94,00	21,28	29,79	21,28	26,60	42,55	56,38	0,89	1,00	-22,80	-17,70	0,09	13,5 9	-24,26	-0,57
>sfr-miR-10454a-11	GUUGACACUCCGGGGC CUUAGUCUGCUAUUAC AAUUGUGUCACAAUG GUCGAUCACUGUAAU AGCAGACUAAGGCCCC UGAUGUCGCC	89,00	23,60	22,47	25,84	26,97	49,44	49,44	1,20	0,91	-51,40	-51,40	0,23	4,02	-57,75	-1,17
>sfr-miR-10454a-12	AGUCUGCUAUUACAA UUGUGUCACAAUGGU CGAUCACUGAACAGUG CAAGAGUGCAGUGCUC AGUGAUCGGCCAUUCU GACACAAUUGUAAUA GCAGACU	101,00	22,77	28,71	20,79	26,73	43,56	55,45	0,93	1,10	-73,40	-73,40	0,43	1,19	-72,67	-1,67
>sfr-miR-10454a-13	UCUGCUAUUACAAUU GUAUCAGAAUGGUCG AUCACUGAGCAGUGC AGAGGGACGGAGCUA	100,00	23,00	32,00	20,00	24,00	43,00	56,00	0,75	1,15	-36,80	-31,10	0,03	15,3 6	-36,80	-0,86

	UACAACCUACAUAGCU CUGACACAAUUGUAA UAGCAGA															
>sfr-miR-10454h-1	CUGCUAUUACAAUUA UGUCAGAAUGAUCGA UCAUUGAGCAGUGCA AGAGGGACGGAGCUA UGUAGGUUGUAUAUU CUGACACAAUUGUAA UAGUAG	97,00	24,74	30,93	13,40	29,90	38,14	60,82	0,97	1,85	-37,50	-34,90	0,13	6,01	-38,66	-1,01
>sfr-miR-10454f	UGCUAUUACAAUUGU GUCAGAAUGGUCGAU CACUGAGCAGUGCAAG AGGGACGGAGCUAUG UAGGUUGUAUAGCUC CGUUCUGACACAAUUG UAAUAGCA	101,00	26,73	27,72	16,83	27,72	43,56	55,45	1,00	1,59	-48,80	-47,60	0,03	11,5 5	-48,32	-1,11
>sfr-miR-10454e-1	GCUAUUACAAUUGUG UCACAAUGGUCGAUCA CUGAGCAGUGCGAGA GGGACGGAGCUAUGU AGGUUGUAUAGCCAU UGUGACACAAUUGUA AUUGU	97,00	27,84	26,80	15,46	28,87	43,30	55,67	1,08	1,80	-43,90	-40,30	0,15	9,28	-45,26	-1,05
>sfr-miR-10454b	UAUUACAAUUGUGUC ACAAUGGUCGAUCACU GAGCAGUGCGAGAUG GACGGAGCUAUGUAA GUUGUAUUUAUACG ACCAUUCUGACACAAU UGUAAUA	100,00	22,00	31,00	16,00	30,00	38,00	61,00	0,97	1,38	-37,00	-30,97	0,09	16,4 9	-37,00	-0,97
>sfr-miR-10454h-2	UAUUACAAUUGUGUC AGAAUAGUCGAUCGU UGAGCAAUGCAAGAG GGACGGAGCUAUGUA GGACUGCUCAAUGGUC	101,00	24,75	30,69	15,84	27,72	40,59	58,42	0,90	1,56	-59,00	-59,00	0,33	2,41	-58,42	-1,44

	GACUAUUCUGACACAA UUGUAAUA															
>sfr-miR-10454e-2	UACAAUUGUGUCACA AUGGUCGAUCACUGA GCAGUGCAAGAGGGA CGGAGCUAUGUUGGU UGUAUAGCUCCAUCCC AUGACCAUUGUGACAC GAUUGUA	100,00	26,00	26,00	20,00	27,00	46,00	53,00	1,04	1,30	-53,70	-53,70	0,16	2,90	-53,70	-1,17
>sfr-miR-10454h-3	UAAAAUUGUGUCAGA AUGGUCGAUCAUUGA GCAGUGCAAGAGGGA AGGAGCUAUAACAACCU ACAUAGCUCAGUCCCU CGACCAUUCUGACACA AUUGUC	100,00	22,00	31,00	22,00	24,00	44,00	55,00	0,77	1,00	-48,20	-46,90	0,20	5,78	-48,20	-1,10
>sfr-miR-10454h-4	GUCAGAAUGAUCGAU CAUUGAGAAGUGCAA GAGGGACGGAGCUAU GUAGGUUGUAUAGCU UCUUCUCUCUUGCACU GCUCAAUGGUCGACUA UUCUGAC	100,00	26,00	24,00	19,00	30,00	45,00	54,00	1,25	1,37	-64,00	-64,00	0,29	1,99	-64,00	-1,42
>sfr-miR-10454h-5	UGUCAGAAUGGUCGA UCACUGAGCAGUGCGA GAGGGAUGGAGCUAU GUAGGUUGUAUAGCU CCAUCCCUCUCGCAAU GUCAAUGAUCGACCAU UCUGACA	101,00	26,73	24,75	21,78	25,74	48,51	50,50	1,04	1,23	-76,30	-76,30	0,12	3,29	-75,54	-1,56
>sfr-miR-10454h-6	UGUCAGAAUGGUCGA UCAUUGAGCAGUGCA AAGGGAAGGAGCCAU ACAACCUACAUAGCUU CGUCCCUCUUGCACUG	101,00	21,78	28,71	24,75	23,76	46,53	52,48	0,83	0,88	-64,80	-64,80	0,19	2,34	-64,16	-1,38

	AUCAAUGAUCGACCAU UCUGACA															
>sfr-miR-10454h-7	UGUCAGAAUGGUCGA UCAUUGAGCAGUGCA AGAGGGAUUAUAGCUA UACUACCUACAUAGCU GUCCCUCUCGCACUGC UCAAAGAUCGACCAUU CUGACA	100,00	21,00	28,00	25,00	25,00	46,00	53,00	0,89	0,84	-70,60	-70,60	0,18	2,87	-70,60	-1,53
>sfr-miR-10454e-3	UGUCACAAUGGUCGA UCACUGAGCAGUGCAA GAGGGAUGAAGCUAU ACAACCUACACAGCUC CGUCCCCUCGCACUGC UCAGUGAUCGACCAUU CUGACA	101,00	21,78	26,73	29,70	20,79	51,49	47,52	0,78	0,73	-67,10	-67,10	0,32	2,34	-66,44	-1,29
>sfr-miR-10454d	GUGUCAGAAUGGUCG AUCAUUGGGCAGUGC AAGAGGGAUUGAGCU AACUUACAUAAGCUGCG UCCCUUUGCAGCUGCU CAAUGAUAACCAUUC UGACAC	100,00	24,00	25,00	24,00	26,00	48,00	51,00	1,04	1,00	-76,90	-76,90	0,82	0,46	-76,90	-1,60
>sfr-miR-10454e-4	UUGUGUCACAAUGGU CGAUCACUGAGCAGUG CAAGAGGGAUGGAGC UAUACAACCUACAUAG CUCCGUCACUGCUCAG UGAUCGACCAUUGUG ACACAA	100,00	24,00	28,00	24,00	23,00	48,00	51,00	0,82	1,00	-70,60	-70,60	0,26	1,95	-70,60	-1,47
>sfr-miR-7926	ACUCAGAAGCUCAGAU GAGUUGAUAACUUC AAAGUCUCUAACAUCU CUUUAUUUAUUCGU UUGAUUUUGAAAUCA	97,00	14,43	29,90	17,53	37,11	31,96	67,01	1,24	0,82	-20,40	-19,10	0,25	4,44	-21,03	-0,66

	UGUCAAUGAUUUCU GCAG															
>sfr-miR-285	UAAGUCCACAAUUUG ACAACUGUAUUCGAG UGAGUGGAUAGAGUU GUGAUUCUCUAGCACC AUUCGAAUUCAGUGC UCGAAUUUGGACGAU	92,00	22,83	27,17	17,39	31,52	40,22	58,70	1,16	1,31	-38,80	-38,80	0,48	1,65	-42,17	-1,05
>sfr-miR-971	AAGAAGUCCGGUAUC UCGGCUCACUCUAAGU AUGAACGCCAAGCAUA UUGAAAAGAGUUUGG UGUUCUACCUUACAGU GAACUGAGUACUAGA CAUUCAU	101,00	20,79	30,69	19,80	27,72	40,59	58,42	0,90	1,05	-40,70	-40,70	0,30	3,49	-40,30	-0,99
>sfr-miR-7006	GUAGAAAAUUGAAGU CAAGAAGCUGGUAGA UGAUUUCAAGACCCAC AGGGUUUCUGACCUG GAUCCUGACUUCUCCU UUCGGU	84,00	23,81	26,19	20,24	28,57	44,05	54,76	1,09	1,18	-22,30	-20,80	0,16	6,09	-26,55	-0,60
>sfr-miR-10508a-2	AGCUAUUCUAGAGGC UCAUAUGCUGCAGUA GCAGCGCGACAAUCGG CGCGUCGUGAGUCGCG GAAUGCUCUCAUGAA UAUGACCCUGAAAUA UGA	97,00	26,80	25,77	23,71	22,68	50,52	48,45	0,88	1,13	-30,50	-30,50	0,13	12,8 5	-31,44	-0,62
>sfr-miR-10508a-3	AGCCAUUCAAGAGGCU CAUAUGAUCAGUAGC AGCGCGACAAUCGCCG CGUCAUGUGUCGCGGA AUGCUGCUCAUGAAU AUGAGCCUCUAGCAUG ACU	98,00	25,51	25,51	25,51	22,45	51,02	47,96	0,88	1,00	-42,20	-37,60	0,04	21,0 5	-43,06	-0,84

>sfr-miR-10508b-2	UCAAGCCAUGCUAGAG GCUACUACGAGUAGCA GCGCGGCAAUCGCCGC GUCGUGUGUCGCGGA AUGCUGCUCAUGAAU AUGAGCCUCAAGCAUG GCUUGA	101,00	29,70	22,77	25,74	20,79	55,45	43,56	0,91	1,15	-56,60	-56,60	0,13	7,81	-56,04	-1,01
>sfr-miR-10508a-4	AGCCAUGCAAGAGGCU CAUAUUCAUGAGGAG CAUUUCGCGACACACG ACGCGGCGAUUGUCGC GAAAUUGCUCUCAUGA AUAUGAGCCUCUUGCA UGGCU	101,00	25,74	24,75	25,74	22,77	51,49	47,52	0,92	1,00	-86,40	-86,40	0,66	0,66	-85,54	-1,66
>sfr-miR-10508b-3	AAUUAUUCAUUCCAGC AGCAUUCCGCGACAAU CGCCGCGUCGUGUGUC GCGGAAUGCUGCUCAU GAAUAUGA	73,00	23,29	23,29	26,03	26,03	49,32	49,32	1,12	0,89	-40,20	-40,20	0,41	3,20	-55,07	-1,12
>sfr-miR-10461b-2	CUAGAAGCUCAUAUUC AUGAGCAGCAUUCCGC GACAAUCGCCGCGUCG UGUGUCGCGGAAUGC UACUGAUGAAACUAG UCGAGUCCUC	90,00	24,44	23,33	26,67	24,44	51,11	47,78	1,05	0,92	-38,20	-38,10	0,05	7,99	-42,44	-0,83
>sfr-miR-10508b-4	UGUUUGUAUUAUAAG AAGCAGCGCGACCGGG GACAAUCACCGCGUCG UGUGUCGCGGAAUGC UGCUC AUGAAUAUAA GCC	81,00	28,40	24,69	22,22	23,46	50,62	48,15	0,95	1,28	-26,30	-10,15	0,03	22,2 6	-32,47	-0,64
>sfr-miR-10508a-5	ACAUCACACGAAUACG AAUAGUUCAUAUUC GCGACAAUGUUGUCGC GGAAUGCUGCUCAUG	90,00	20,00	30,00	18,89	30,00	38,89	60,00	1,00	1,06	-25,60	-24,70	0,03	18,0 2	-28,44	-0,73

	AAUAUGAACUUUUAG CAUGUUUGAGAU																
>sfr-miR-10508a-6	UCAAGCCAUGCUAGAG GCUCACACACAUGCGU CGUGUGUCGCGAAAU GCUGCUCUAUGAAU GAGCCUCUAGCAUGGC UAUU	83,00	24,10	24,10	25,30	25,30	49,40	49,40	1,05	0,95	-44,50	-44,50	0,60	8,12	-53,61	-1,09	
>sfr-miR-10461a-1	GUUUGACGAGGAACU CGACUAGUGUCGUGU GUCGCGAAAU CAUGAAUAUAAGCCUC UAUCAUGGCUUGAAA CUAGUCGAGUUUCUCG UCAAAC	100,00	24,00	25,00	21,00	29,00	45,00	54,00	1,16	1,14	-52,00	-52,00	0,28	1,99	-52,00	-1,16	
>sfr-miR-10461a-2	UUGACUAGGAACUCA ACUAGUUUAGUACUA GCGCGACUGUCGCCGC CUCUCUAGUAUGGCUU GAUAUGAAACUAGUC GAGUUUCUCGUCAA	92,00	21,74	23,91	22,83	30,43	44,57	54,35	1,27	0,95	-30,50	-24,57	0,02	14,9 6	-33,15	-0,74	
>sfr-miR-10461b-3	CUGUUUGACGAAGACC UCGACUAGUUUCAAGC CGUACUAGAGACUUA UAAUUUAUACUACUGA UCAUGAAUAUGAAAC UAGUCGAGUUCUCGU CAAACAG	101,00	17,82	31,68	20,79	28,71	38,61	60,40	0,91	0,86	-38,70	-37,70	0,12	8,84	-38,32	-0,99	
>sfr-miR-10461b-4	UUACUCACGUAACUGU UUGUCGAGGAAUGCU GCUCAUGUAUGAGUC UCUAACAUGGCUUGA AACUAGUCGAGUCCU CGUCAAAACAGUUACGU GAGUAA	100,00	22,00	26,00	20,00	31,00	42,00	57,00	1,19	1,10	-49,20	-49,00	0,06	12,8 5	-49,20	-1,17	

>sfr-miR-10508a-7	CUACACUGUUACGUGU GUCGUGGAAUGCUGC UCAUGAAUAUGAGCC UCUAGCAUGGCUUGA AACUAGUCGAGUUUC UCGUCAAAACGUAACG UGAGU	98,00	24,49	24,49	20,41	29,59	44,90	54,08	1,21	1,20	-30,50	-28,70	0,10	7,95	-31,12	-0,69
>sfr-miR-10461b-5	ACUCACGUGACGUUUU GACGAGGAAUUCGAC UGCUCAUAAAUAUGA GCCUCUAGCAUGGCUU GAAACUAGUCGAGUU CCUCGUCAAACAGUUA CGUGAGU	101,00	22,77	26,73	21,78	27,72	44,55	54,46	1,04	1,05	-55,80	-55,80	0,17	10,4 9	-55,25	-1,24
>sfr-miR-10461b-6	AUUAUUUUUCAACG GCUUACUCAUUGGCCU CUAGCAUGGCUUGAA ACUAGUCGAGUCCUC GUCAAACAGUUACGU GAGUAAGCCGAUAAU AUA AUGAU	101,00	18,81	28,71	19,80	31,68	38,61	60,40	1,10	0,95	-30,80	-30,10	0,09	14,5 4	-30,50	-0,79
>sfr-miR-10461a-3	AAUUGUCCGACGAGG AACUUCAGUAGCAGCG CGACGGCGCUCAUAAA UAUGAGCCUUUAGUA UGGCUUGAAACUAGU CGAGUUUCUCGUCAAA CAGUU	99,00	24,24	27,27	21,21	26,26	45,45	53,54	0,96	1,14	-37,50	-36,30	0,20	5,51	-37,88	-0,83
>sfr-miR-92b	ACUGCGGUGGCCACGC UUAGGACGCGAUUCG GUGUAAAACGUUGUGA CUUGUAUCAAAUUGC ACCAAUCCCGGCCUGC CUGUGGUCACCGCGCC	94,00	27,66	18,09	29,79	23,40	57,45	41,49	1,29	0,93	-41,50	-40,20	0,08	6,65	-44,15	-0,77
>sfr-miR-9685	GACGCCGGAGUGAAGC UGCUGCACUCGCCGAC	98,00	28,57	17,35	37,76	15,31	66,33	32,65	0,88	0,76	-33,10	-27,20	0,03	28,9 8	-33,78	-0,51

	CGGGUGCAUACAGUGC GUGUACCUCACCACGC CCAGCAUGCAGCUCGC CUUCAACGCCGUGCCG C															
>sfr-miR-10502-1	GAUAGAAAAUCCCAAC GAAGUUGGGCAGAAA GCCCGCCAGGCUGAUC ACCUCGCCUGGUCGGA GGAUCCUCCUUUUCUU UGGGAACUUUACUCUC	96,00	23,96	22,92	28,13	23,96	52,08	46,88	1,05	0,85	-34,90	-30,50	0,08	9,11	-36,35	-0,70
>sfr-miR-10502-2	AACAACAAAAGGGCA GAAAGCCCGCCAGGCU GAACACCUCGCCUGGU CGGAGGAUCCUCCUUU UCUUAGG	71,00	25,35	26,76	29,58	16,90	54,93	43,66	0,63	0,86	-24,90	-24,90	0,42	5,72	-35,07	-0,64
>sfr-miR-12237	GCGGGGCGCGGGGGG GCUGACCUGGUAGUA GACAGUGUUGGGGUG UCCUUGUCGGGGAUCC AGGUGACGACACCAGG GUUGCCCUCCUCGCAG ACCAGC	100,00	41,00	14,00	26,00	18,00	67,00	32,00	1,29	1,58	-44,60	-44,60	0,06	16,4 2	-44,60	-0,67
>sfr-miR-4705	UAAUUGUUGCUUAUA AGCGGUUGCGUGACCU GCUGGGUGAUUCGGU UACCGUCAAUACAUUG GUAAUGCAACCAAGU G	79,00	26,58	21,52	18,99	31,65	45,57	53,16	1,47	1,40	-25,30	-25,30	0,22	6,23	-32,03	-0,70
>sfr-miR-12389	CCGUCUGAAGUUAAG GUCGAGAAUCGCUGG GAAGGCGGCAGCACUA UGGUCCUCGAUGCGAU UACGUUGCGUAACUU GAGAAUU	85,00	30,59	23,53	20,00	24,71	50,59	48,24	1,05	1,53	-22,10	-15,70	0,05	19,2 0	-26,00	-0,51

>sfr-miR-2064	CCCGAGUCGACCGAAG CGAUACCACGGCCACG GUUAUAGGAAACCGG UGUGAACGGUUGCAU UGUGUUUCGCCGGGU GAGUAGGUUACGGCG ACCUCUUC	101,00	31,68	19,80	25,74	21,78	57,43	41,58	1,10	1,23	-33,50	-33,50	0,13	14,3 6	-33,17	-0,58
>sfr-miR-981	AAGAAUCCGCAUAUA AUUCGGGUUUCGGGA CGACGGAACCGUGAUU AAAUUGGUUCGUUGU CGACGAAACCUGCAAA AUGUGCGGUUGAA	91,00	27,47	28,57	17,58	25,27	45,05	53,85	0,88	1,56	-39,80	-39,80	0,10	5,12	-43,74	-0,97
>sfr-miR-11240-1	AGAACACAAAGAUUG CACGGUUGGUGCGUU AGCUGGGCAACCGGCU GCUGCGCAACGUGUAG CGGGUUCGAUUCCCGC ACGAAACUACUCUUUG UGUUAU	101,00	28,71	21,78	23,76	24,75	52,48	46,53	1,14	1,21	-45,90	-45,90	0,28	3,89	-45,45	-0,87
>sfr-miR-11240-2	UGCACGGUUGGUGCG GUGGCUGGGCAAACG GCUGCCACGUAGCGUG UAGCGGGUUCGAUUCC CGCACGGACCAACUGU UUG	82,00	36,59	14,63	25,61	21,95	62,20	36,59	1,50	1,43	-37,30	-37,30	0,41	4,94	-45,49	-0,73
>sfr-miR-11240-3	AGUUGCACGAAUAAA AUCGUCUGCACGGUUA GCGCAGUGAUUGGGU UCCGCGCAACGUGUAG CGGGUUCGAUUCCCGC ACAG	83,00	30,12	21,69	24,10	22,89	54,22	44,58	1,06	1,25	-31,70	-25,80	0,24	3,74	-38,19	-0,70
>sfr-miR-11240-4	AAUGAUUAUACAGUU GGUGCGUUGGCUGGA CAACCGGCUGGCGCGC AACGUGUAGCGGGUU	96,00	30,21	20,83	26,04	21,88	56,25	42,71	1,05	1,16	-43,90	-43,90	0,36	8,32	-45,73	-0,81

	CGAUUCCCCGCACGGAG CAACCCUGUGUAAUCC AC															
>sfr-miR-2788	GCAAGUCGCCGUAGAG CUGGGGUUCAAAGC GGCAUGUGCCACCUGC UAUGCAAUGCCCUUGG AAAUCCCAAACGUGCU GGCGACUGUC	90,00	28,89	21,11	27,78	21,11	56,67	42,22	1,00	1,04	-39,70	-39,50	0,10	6,42	-44,11	-0,78
>sfr-miR-10490-1	UUUUCUAUAAACAUC AGGGAACCCAAGACCU UCCAACGAAUGCAAG ACCGUGGAAAUCGGU UCAUGCGUUCUGGAG UUAUAGAUAG	88,00	21,59	30,68	20,45	26,14	42,05	56,82	0,85	1,06	-22,50	-17,20	0,02	18,3 5	-25,57	-0,61
>sfr-miR-10490-2	UUUGCUAUAAACCUCA CGGAGCCCGAGACCUU UCCAACGAAUGCAAGA CCGUGGAAAUCGGUUC GUGCGUUCUGGAGUU AUAGCGUC	88,00	25,00	23,86	25,00	25,00	50,00	48,86	1,05	1,00	-29,80	-23,20	0,02	8,63	-33,86	-0,68
>sfr-miR-10490-3	CUGCUAUAAACCUCAC GGAGCCGAGACCUUUC CAACGAAUGCAAGACC GUGGAAAUCGGUCCG UGCGUUUUGUAGUUA	79,00	24,05	25,32	25,32	24,05	49,37	49,37	0,95	0,95	-28,60	-27,10	0,16	4,85	-36,20	-0,73
>sfr-miR-10511-1	CGACAGAUACUGUUCG ACAAACCAAUCGAUCG AUUGCCGGUCGAUGGC AUCGUUCAACGUCGAU CGAUGUUGAUCGACU GGUCUGGUA	89,00	25,84	23,60	23,60	25,84	49,44	49,44	1,10	1,10	-33,60	-33,60	0,23	8,59	-37,75	-0,76
>sfr-miR-10511-2	UCAUGCCAUCGAUCGA CAUUGAUCGAUUGGU GUGGUAGCUUUAUU GAUCGGUUACCGGUCG	99,00	27,27	20,20	21,21	30,30	48,48	50,51	1,50	1,29	-50,60	-50,60	0,19	5,26	-51,11	-1,05

	AUUUACCAAUCGAUCG AUUGCCGGUCGAUGGC AGCG																
>sfr-miR-10464-1	GACUGCAUUUCAAUU GCAAUCCAACUGAACA UUGCAGUUCAAGGGC AGUUUCA AUGCAGUU GACACGACUGCAAUAG AACUGCAAGC	88,00	20,45	31,82	22,73	23,86	43,18	55,68	0,75	0,90	-29,80	-29,10	0,21	6,09	-33,86	-0,78	
>sfr-miR-10464-2	UUGACACGACUGCAAU AGAACUGCAAAUCAA ACGUGCGGUCCGAUUG UAGUUGAAUUGCAGU CGGUGUGCA	72,00	26,39	27,78	19,44	25,00	45,83	52,78	0,90	1,36	-29,30	-29,30	0,06	4,77	-40,69	-0,89	
>sfr-miR-10464-3	AUGCAGUUGACACGAC UGCAAUAGAACCGCAA ACCAAACGUGUAGUCC GAUAGUAGUUCUAUU GCAAUCGUACCAGCUG CAU	83,00	20,48	31,33	24,10	22,89	44,58	54,22	0,73	0,85	-33,30	-33,30	0,33	2,42	-40,12	-0,90	
>sfr-miR-10464-4	UGUGCAGUUCACAUA AUGCUAGGCUGCACGG UUGGCUUGCAGUACCA UUGCAGUCGUGUCAAC UGCAUU	70,00	25,71	20,00	22,86	30,00	48,57	50,00	1,50	1,13	-24,60	-23,40	0,13	14,1 4	-35,14	-0,72	
>sfr-miR-2856	GCCAUAGAAUAAUAA GUACAACUACGGCCUA AUUUUAAAACGAAACU CCACAUUCGAGAACCG UAGUAGGCCCUAGGCU AUCUUAUCUUUUAUU UU	96,00	14,58	33,33	21,88	29,17	36,46	62,50	0,88	0,67	-18,60	-14,00	0,09	12,9 1	-19,38	-0,53	
>sfr-miR-10466	UGCUAUAAUUAUCAG CACCGUCCAAAUGUCA AACACUGAUACUUUU	70,00	18,57	32,86	17,14	30,00	35,71	62,86	0,91	1,08	-23,40	-23,40	0,37	2,66	-33,43	-0,94	

>sfr-miR-7847	ACGAGCACUAUGCGUG CGUGGAGGACGAGGA GAAUAUGUCGGCGGC GCGCCACGCCUCGGCC UCCUCCUCCGCGUCGU ACUGCUGCA	88,00	32,95	15,91	32,95	17,05	65,91	32,95	1,07	1,00	-41,30	-41,30	0,19	13,1 3	-46,93	-0,71
>sfr-miR-2779-2	UUGAUCCGGCUCGAAG GACCAGAAUGUGCGG GUCGGAGGGAUCAGG CUGUGGAUCUAUUCU CGACCCGCACAUUCUG GUCCUUCGAGCCGGAU UGU	98,00	31,63	17,35	25,51	24,49	57,14	41,84	1,41	1,24	-78,50	-78,00	0,13	7,26	-80,10	-1,40
>sfr-miR-10485	AAUCGCUGCGCGGGGG ACAGUCUACCCGGACA GGCCGACUACUGUAUG UCGCACUGGACGGGCA GGCCGCCCUCCUCACA AGCGUCA	88,00	30,68	19,32	34,09	14,77	64,77	34,09	0,76	0,90	-41,00	-38,60	0,32	5,94	-46,59	-0,72
>sfr-miR-10477-1	GUCGGCGACGGCGACC UCUCGCGCCCGGCACU GGUUCAGGCCAUGGU GCGGAGCGAGAGGAU GGGAUGCCGUCUCCUC C	80,00	37,50	12,50	32,50	16,25	70,00	28,75	1,30	1,15	-39,80	-39,80	0,08	10,1 8	-49,75	-0,71
>sfr-miR-10477-2	GUCGGCGACGGCAACC UCUCGCGCUCGGCACU GGUUCAGGCCAUGGU GCGGAGCGAGAGGGA CUGGGAUGCCGUCUCC UCC	82,00	36,59	13,41	31,71	17,07	68,29	30,49	1,27	1,15	-43,40	-42,30	0,19	4,66	-52,93	-0,77
>sfr-miR-10477-3	GCUGAGCACCGCCAAG UCCUCAGGGACCUCUC GCGCCCAGCACUGGUU CAGGCCAUGGUGCGGA	93,00	33,33	16,13	33,33	16,13	66,67	32,26	1,00	1,00	-42,30	-41,50	0,15	11,4 0	-45,48	-0,68

	GCGAGAGGGACUGGG AUGCCGUCUCCUC																
>sfr-miR-10477-4	CGCUAGGUACAUAACC UUGCGGAGCGAGAGG GACUGGGAUGCCGUCU CCUCCUUCUGCGAAGC AGUCAUGCAGCUAA GG	81,00	30,86	20,99	25,93	20,99	56,79	41,98	1,00	1,19	-29,40	-25,50	0,10	14,5 8	-36,30	-0,64	
>sfr-miR-7398p-1	UCUUGUGUACUCGCGC UCGCCGCCACCAGCUC CGUCAACUACUGCGGC GCCAAGAUGUGUGGA GGCAAGGAUACACAU AC	81,00	25,93	20,99	32,10	19,75	58,02	40,74	0,94	0,81	-25,20	-25,20	0,26	6,22	-31,11	-0,54	
>sfr-miR-7398p-2	UCCUGUGUACUCGCGC UCGCCGCCACCAGCUC CGUCAACUACUGCGGC GCCAAGAUGUGUGGA GGCAAGGAUACACAU AC	81,00	25,93	20,99	33,33	18,52	59,26	39,51	0,88	0,78	-25,20	-25,20	0,25	7,09	-31,11	-0,52	
>sfr-miR-10470-1	CCGUUUCUACUUCACC UUUUCGAGCCGGAGCC CCGGUAAACUUGCCAA CCUCAGUAUCUACUGG GCUCCAUCUGAAUACA GGUAAGCUUAGAAAU AU	98,00	18,37	24,49	28,57	27,55	46,94	52,04	1,13	0,64	-28,60	-28,60	0,04	10,3 5	-29,18	-0,62	
>sfr-miR-10470-2	CUCCUGCUUGUCGAGC CGGAGCCCCGGUAAGC CCGCUAGGUAGUCCGC AGCUCCGGAUCACACA GCAGAAA	72,00	27,78	20,83	34,72	15,28	62,50	36,11	0,73	0,80	-32,70	-32,70	0,55	2,56	-45,42	-0,73	
>sfr-miR-10470-3	GUGCCUAAUCCUGCUU UUCGAGCCGGAGCCCC GGUAAACCCGCUAGGU AGUCCGCAGCUCCGGA	83,00	26,51	24,10	30,12	18,07	56,63	42,17	0,75	0,88	-27,40	-19,30	0,08	13,9 1	-33,01	-0,58	

	UCAGAACACAAUAGG GAA															
>sfr-miR-10470-4	GUUUCUACUCCUGCUU UUCGAGCCGGAGCCCC GGUAAGCGGUAGGUA GUUCGCAGCUCCGGAU AGCAGUUGUAGAGCU	79,00	30,38	16,46	25,32	26,58	55,70	43,04	1,62	1,20	-29,80	-23,20	0,05	17,3 7	-37,72	-0,68
>sfr-miR-10470-5	GCGUUCCUACUUCUAC UUUUCGAGCCGGAGCC CCGGUAACCCGUUAGG CAUUUCGCAGCUCCGG GUCAAUCGAAUGACA GUUAAAAUAGGAAUU U	96,00	22,92	22,92	26,04	27,08	48,96	50,00	1,18	0,88	-29,00	-25,10	0,05	10,9 3	-30,21	-0,62
>sfr-miR-10470-6	UCCUGCUUUUCGAGCC GGAGCCCCGGUAAACC CGCUAAGUAGUCCGCA GCUCCGAGCUUCAAG GCAUAC	71,00	23,94	21,13	33,80	19,72	57,75	40,85	0,93	0,71	-28,10	-28,10	0,36	2,75	-39,58	-0,69
>sfr-miR-10493-1	UUCCCUCGUUCUGAUC CGAAGCUGCGGACUAC CUAGCGGGUUUACCGG GGCUCCGGGUCGGAAA GCAGGAGU	73,00	32,88	16,44	27,40	21,92	60,27	38,36	1,33	1,20	-35,20	-34,90	0,28	4,11	-48,22	-0,80
>sfr-miR-10493-2	GAUCCAUAUCUAGUU ACAAAUUUUCUGAUCC GAAGCUGCGGACUACC UAGCGGGUUAACCGG GGCUCCGGUUCGAAAA GCAGGAAUAGGUACG GGU	98,00	28,57	25,51	21,43	23,47	50,00	48,98	0,92	1,33	-33,00	-31,80	0,11	7,07	-33,67	-0,67
>sfr-miR-10493-3	UUUAGAUGUAACUGC UAACUACCUACCGGGU UUACCGGGGCUCCGAC UCGCCAAGCAGGAGUA GGAACGGGGUGGUUU	99,00	29,29	23,23	21,21	25,25	50,51	48,48	1,09	1,38	-32,70	-26,40	0,13	12,3 0	-33,03	-0,65

	UUAGUCAGUAAGAGU CUGAC															
>sfr-miR-10493-4	UUACCGGGUGUCUGGC UCGAAAGGCAGGAGU AGAAACGGGGUGGUU UUUAGUCAGUAAGAA UCUGACACUCUCGCUC GCCUCGCCCCGACGCC GGAGA	99,00	31,31	21,21	25,25	21,21	56,57	42,42	1,00	1,24	-40,70	-34,80	0,14	12,8 9	-41,11	-0,73
>sfr-miR-10493-5	GGGUCUCCCGCUUGAG AAGCAAGAGUAGGAA CGGGGUGGUUUUAG UCAGUAAGAGUCUGA CACUCCCUCUCGCUUC ACUCAAGGCGGGAGA AGU	96,00	31,25	22,92	21,88	22,92	53,13	45,83	1,00	1,43	-38,60	-37,30	0,05	13,8 4	-40,21	-0,76
>sfr-miR-1175	GACGCGUGAAGGAUC AAGUGGAGGUGUGAU CUCUUCACUUUUUAUG UUUAAAGUGAGAUUC AACUCCUCCAACUUA UCCGAAACGCUUU	92,00	20,65	26,09	19,57	32,61	40,22	58,70	1,25	1,06	-32,00	-32,00	0,03	8,62	-34,78	-0,86
>sfr-miR-10499b-1	CUCUGCAUGGGGCGGU ACAAGGGUUUGAAAC UCCAUACAUGUGUAU GAGAAUCUGCAUGGA GUUUCAAACUCUUGA AUUGCUAUAUGCAAU G	93,00	24,73	26,88	17,20	30,11	41,94	56,99	1,12	1,44	-44,50	-44,50	0,48	3,24	-47,85	-1,14
>sfr-miR-10499b-2	CUGCAUAGGGCAGUU AAGGGUUUGAAACUC CAUACAUGUAAAUGC GCUUCUGCAUGGAGU UUCAAAACUCUUGAACU GCUCUGUGUAA	88,00	22,73	26,14	19,32	30,68	42,05	56,82	1,17	1,18	-47,90	-45,70	0,05	8,72	-54,43	-1,29

>sfr-miR-10499b-3	UAUCAAUGCAUGGGA CAGUGCAAGGGUUUA AAACUCCAUAACAUGUA UAGCAUCUGCAUGGA GUUUCAAACUCUUGG AUUGCUCUGUGCAAU GUCA	96,00	21,88	28,13	18,75	30,21	40,63	58,33	1,07	1,17	-39,10	-39,10	0,08	6,99	-40,73	-1,00
>sfr-miR-2766	GAAAACGCCCCGUCUCG UCCAUCCUUCGUCUCG ACUGGCGCUGCUUAAC GCUUCAGUCUUGUCGA AUGGUGGGUGAGAUU UGCGUUCUA	89,00	25,84	15,73	26,97	30,34	52,81	46,07	1,93	0,96	-38,40	-36,60	0,34	3,90	-43,15	-0,82
>sfr-miR-10060	AAGCAUGUAGUAAUA UGAAAUGUGUAAUUC CAUGCCAAGGAGCGUG UGGCGUUGCCGAAAA UGCCAAGUGAUUAACC ACAAUCCAAUUUGCUC AUGCCC	101,00	21,78	30,69	21,78	24,75	43,56	55,45	0,81	1,00	-30,70	-30,70	0,18	5,20	-30,40	-0,70
>sfr-miR-10491-1	CAUCAAAACUUAGUGCU GUGCUCACAUGUCAAC CAGGGGGUGACGCUCC AUGAUAGUGCCCUGA UUAACAUGUCAGUAC ACACUGAGCUUGUAC	94,00	22,34	25,53	25,53	25,53	47,87	51,06	1,00	0,88	-34,00	-32,50	0,20	6,54	-36,17	-0,76
>sfr-miR-10491-2	CUUCAAAACUUGGUGCU GUGCUCACAUGUCAAC CAGGGGGUGACGCUCC AUGAUAGUGCCCUGA UUAACAUGUCAGUAC ACACUGAGCUUGUAC	94,00	23,40	23,40	25,53	26,60	48,94	50,00	1,14	0,92	-32,70	-31,20	0,20	6,49	-34,79	-0,71
>sfr-miR-466i	AGUGAAUGAGCCGGC AUGUCAUCGUGUGUG UGUGUGUGUGUGUGU GGCGCGCGCCACACGC	93,00	34,41	13,98	27,96	23,66	62,37	37,63	1,69	1,23	-49,20	-44,90	0,12	11,5 4	-52,90	-0,85

[illegible]

[illegible]

4h-4-3p sfr-miR b-137b-2-5p	NA	NA	2	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	2,00	1
sfr-miR b-10454h-7-3p	NA	NA	NA	2	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	2,00	1
sfr-miR b-10475-5p	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	2	NA	NA	NA	NA	NA	2,00	1
sfr-miR-10509-5p	1	NA	NA	NA	NA	NA	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	2,00	2
sfr-miR b-10455-5p	1	NA	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	2,00	2
sfr-miR b-10481-5p	1	NA	NA	NA	NA	NA	NA	1	NA	NA	NA	NA	NA	NA	NA	NA	2,00	2
sfr-miR b-	1	NA	NA	NA	NA	NA	NA	NA	NA	1	NA	NA	NA	NA	NA	NA	2,00	2

sfr-miR-1046 3-5p	NA	NA	NA	NA	NA	NA	NA	NA	1	1	NA	NA	NA	1	NA	NA	NA	3,00	3
sfr-miR b-1045 4h-1-5p	1	NA	2	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	4,00	3
sfr-miR b-1045 4h-4-5p	1	NA	2	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	4,00	3
sfr-miR b-9388 -5p	1	NA	NA	NA	NA	NA	NA	NA	2	NA	NA	NA	NA	NA	1	NA	NA	4,00	3
sfr-miR b-33-3p	NA	NA	2	1	NA	NA	NA	NA	NA	NA	NA	1	NA	NA	NA	NA	NA	4,00	3
sfr-miR b-1048 4-2-3p	NA	NA	NA	NA	NA	2	NA	NA	NA	1	1	NA	NA	NA	NA	NA	NA	4,00	3
sfr-miR b-1049	NA	NA	NA	NA	NA	NA	1	NA	NA	NA	1	NA	NA	NA	3	NA	NA	5,00	3

[illegible]

9-2-3p e
sfr-miR
b-1045
9-4-3p
sfr-miR
b-2796
-5p
sfr-miR-13a-5p
sfr-miR
b-3286
-5p
sfr-miR
b-34-3p
sfr-miR
b-2755
-5p
sfr-miR
b-

sfr-miR b-2796-5p	2	4	2	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	9,00	4
sfr-miR-13a-5p	NA	2	4	1	NA	NA	NA	NA	NA	NA	NA	NA	2	NA	NA	NA	NA	9,00	4
sfr-miR b-3286-5p	1	5	3	2	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	11,00	4
sfr-miR b-34-3p	2	NA	3	5	NA	NA	NA	NA	NA	NA	NA	NA	1	NA	NA	NA	NA	11,00	4
sfr-miR b-2755-5p	1	2	6	4	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	13,00	4
sfr-miR b-	1	NA	6	6	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	1	NA	NA	14,00	4

[illegible]

[illegible]

sfr-miR b-263b-3-5p																			
sfr-miR b-193-3p	24	138	332	95	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	590,00	5
sfr-miR b-993a-2-3p	91	248	646	289	NA	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	1275,00	5
sfr-miR b-750-5p	659	2421	3991	1771	2	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	8844,00	5
sfr-miR b-316-5p	1187	2912	9260	2951	NA	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	16311,00	5
sfr-miR b-1049-5-4-5p	NA	2	1	NA	2	3	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	10,00	6
sfr-miR-	1	NA	5	NA	2	2	NA	NA	NA	NA	1	NA	NA	1	NA	NA	NA	12,00	6

iab-4-5p																			
sfr-miR b-305-3p	69	275	560	212	1	NA	NA	NA	NA	NA	NA	NA	1	NA	NA	NA	NA	1118,00	6
sfr-miR b-252a-5p	114	268	553	240	NA	NA	1	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	1177,00	6
sfr-miR b-1175-3p	553	117	1048	992	2	4	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	2716,00	6
sfr-miR b-375-5p	45	2007	898	173	1	NA	NA	NA	NA	1	NA	NA	NA	NA	NA	NA	NA	3125,00	6
sfr-miR b-1175-5p	1175	2441	5396	2683	NA	2	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	1	11698,00	6
sfr-miR b-10461b-5-5p	1	2	NA	1	1	NA	NA	2	NA	1	NA	NA	NA	1	NA	NA	NA	9,00	7
sfr-miR b-	1	NA	NA	3	1	NA	NA	1	2	NA	1	NA	NA	NA	1	NA	NA	10,00	7

1047 1-1- 5p e sfr- miR b- 1047 1-2- 5p sfr- miR b- 1049 2c-6- 3p sfr- miR b- 1046 1a-1- 3p; sfr- miR b- 1046 1a-2- 3p e sfr- miR b- 1046 1a-3- 3p sfr- miR b-	NA	1	NA	2	1	NA	2	1	NA	NA	4	NA	NA	NA	3	NA	NA	14,00	7
	9	3	NA	1	NA	NA	NA	1	NA	1	1	NA	NA	1	NA	NA	NA	17,00	7
	1	1	NA	2	NA	NA	NA	NA	5	NA	2	NA	NA	2	6	NA	NA	19,00	7

1124
0-1-
5p;
sfr-
miR
b-
1124
0-2-
5p;
sfr-
miR
b-
1124
0-3-
5p e
sfr-
miR
b-
1124
0-4-
5p
sfr-
miR
b-
1047
6-5p
sfr-
miR
b-
1049
0-1-
5p;
sfr-
miR
b-
1049

2	4	7	5	5	5	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	2	30,00	7
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2	NA	NA	1	15	9	2	NA	NA	NA	NA	NA	NA	1	2	NA	NA	32,00	7
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[illegible]

263b
-3-
3p
sfr-
miR-
6094
-5p
sfr-
miR
b-
927a
-5p
sfr-
miR
b-
133-
3p
sfr-
miR
b-
133-
5p
sfr-
miR
b-
750-
3p
sfr-
miR
b-
274-
5p
sfr-
miR
b-
1050

74	1039	849	206	7	2	1	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	2178,00	7
289	741	744	540	28	12	9	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	2363,00	7
284	2045	5645	1187	NA	1	1	NA	NA	NA	NA	NA	NA	1	NA	NA	NA	9164,00	7
284	2045	5645	1187	NA	1	1	NA	NA	NA	NA	NA	NA	1	NA	NA	NA	9164,00	7
5356	22979	33723	12984	21	15	4	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	75082,00	7
9856	29946	51900	22394	NA	2	NA	NA	1	1	NA	NA	NA	NA	NA	NA	NA	114100,00	7
1	NA	1	4	NA	1	2	NA	NA	1	NA	NA	NA	1	1	NA	NA	12,00	8

8a-1-5p sfr-miR b-1045	1	4	2	1	3	2	NA	NA	NA	NA	NA	NA	NA	1	3	NA	NA	17,00	8
4a-13-5p sfr-miR b-1047	NA	NA	NA	4	4	NA	NA	2	3	3	2	NA	NA	2	1	NA	NA	21,00	8
0-6-3p sfr-miR b-965	1	4	7	7	2	NA	1	NA	NA	NA	NA	NA	1	NA	1	NA	NA	24,00	8
5p sfr-miR-1050	1	6	1	2	12	3	6	NA	1	NA	NA	NA	NA	NA	NA	NA	NA	32,00	8
9-3p sfr-miR b-1046	2	26	4	1	1	NA	2	NA	NA	NA	NA	NA	NA	1	NA	NA	1	38,00	8
4-4-3p sfr-miR b-466i-1-5p e sfr-	NA	3	NA	3	NA	NA	NA	8	4	5	NA	NA	NA	8	4	NA	3	38,00	8

sfr-miR b-927a-3p	117	224	536	299	9	9	2	NA	1	NA	NA	NA	NA	NA	NA	NA	NA	1197,00	8
sfr-miR-13a-3p	139	343	461	335	38	15	4	NA	NA	NA	NA	NA	NA	NA	NA	NA	7	1342,00	8
sfr-miR-1047-2-3p	442	1398	1567	502	3	3	4	NA	2	NA	NA	NA	NA	NA	NA	NA	NA	3921,00	8
sfr-miR b-305-5p	727	1721	3698	1627	2	NA	NA	NA	NA	1	NA	2	5	NA	NA	NA	NA	7783,00	8
sfr-miR b-281-5p	28161	288483	730454	112824	3	8	NA	NA	NA	NA	NA	NA	NA	NA	2	NA	2	1159937,00	8
sfr-miR b-1049-2b-1-5p	2	1	1	1	NA	1	NA	1	NA	1	1	NA	NA	1	NA	NA	NA	10,00	9
sfr-miR b-1049-2b-3-5p; sfr-	2	4	1	1	NA	2	NA	1	NA	2	1	NA	NA	1	NA	NA	NA	15,00	9

miR
b-
1049
2b-
4-5p;
sfr-
miR
b-
1049
2b-
5-5p
e sfr-
miR
b-
1049
2b-
6-5p
sfr-
miR
b-
307-
5p
sfr-
miR
b-
1047
0-2-
3p e
sfr-
miR
b-
1047
0-3-
3p
sfr-
miR

1	1	6	NA	7	NA	NA	1	1	1	NA	NA	NA	NA	1	NA	4	23,00	9
NA	NA	NA	4	4	1	NA	3	3	3	3	NA	NA	2	1	NA	NA	24,00	9
3	13	3	1	NA	1	NA	NA	1	NA	1	NA	NA	3	1	NA	NA	27,00	9

b-10500-1-3p sfr-miR																			
b-10508a-3-3p; sfr-miR	7	10	1	4	1	2	NA	NA	3	NA	NA	NA	NA	1	3	NA	NA	32,00	9
b-10508a-6-3p e sfr-miR																			
b-10508a-7-3p sfr-miR	6	5	8	4	4	NA	NA	1	NA	NA	2	NA	NA	2	NA	NA	1	33,00	9
b-10507-2-5p sfr-miR	8	NA	1	16	NA	1	NA	3	NA	3	2	NA	NA	2	2	NA	NA	38,00	9

b-
1049
0-2-
3p
sfr-
miR
b-
1047
0-1-
5p;
sfr-
miR
b-
1047
0-2-
5p;
sfr-
miR
b-
1047
0-3-
5p;
sfr-
miR
b-
1047
0-4-
5p;
sfr-
miR
b-
1047
0-5-
5p e
sfr-
miR

NA	1	NA	1	20	11	4	3	NA	1	NA	NA	NA	NA	1	NA	1	43,00	9
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b-1047																			
0-6-5p																			
sfr-miR-1046																			
5-1-5p e	4	8	16	4	3	NA	NA	NA	1	1	NA	NA	NA	3	6	NA	NA	46,00	9
sfr-miR-1046																			
5-2-5p																			
sfr-miR-1045	10	35	2	4	3	1	NA	NA	NA	NA	2	NA	NA	1	1	NA	NA	59,00	9
4c-5p																			
sfr-miR-b-1048	6	13	7	6	NA	1	NA	NA	1	NA	3	NA	NA	14	25	NA	NA	76,00	9
9-2-3p																			
sfr-miR-b-1050	NA	NA	NA	NA	76	83	17	10	19	10	9	NA	NA	4	10	NA	NA	238,00	9
3-1-5p																			
sfr-miR-b-11-5p	32	102	175	64	5	1	NA	NA	NA	1	2	NA	NA	NA	NA	NA	1	383,00	9

sfr-miR b-989-3p	NA	1	1	NA	242	97	30	NA	NA	NA	1	1	NA	1	NA	NA	21	395,00	9
sfr-miR b-2a-1-5p	113	153	259	230	5	4	1	1	NA	NA	NA	NA	NA	NA	NA	NA	1	767,00	9
sfr-miR b-10485-5p	124	408	812	357	3	2	NA	NA	1	NA	NA	NA	NA	2	NA	NA	2	1711,00	9
sfr-miR b-10-2-5p	98	1677	542	233	332	187	129	NA	1	1	NA	NA	NA	NA	NA	NA	NA	3200,00	9
sfr-miR-2765-5p	4	NA	NA	1	2	4	3	1	1	NA	1	NA	NA	1	4	NA	NA	22,00	10
sfr-miR b-10503-1-3p	NA	NA	NA	NA	12	4	1	3	2	1	2	NA	NA	1	NA	1	1	28,00	10
sfr-miR b-10454e-4-3p	1	1	3	NA	12	5	4	NA	1	NA	NA	NA	NA	1	1	NA	1	30,00	10

sfr-miR b-1049 5-1-3p; sfr-miR b-1049 5-2-3p e sfr-miR b-1049 5-4-3p sfr-miR b-1045 6-3p sfr-miR b-1049 2a-1-5p sfr-miR b-33-5p sfr-miR	1	NA	NA	NA	5	6	2	1	4	3	2	NA	NA	2	4	NA	NA	30,00	10
	12	4	2	18	4	NA	NA	NA	1	3	2	NA	NA	7	5	NA	NA	58,00	10
	20	2	15	20	2	1	NA	2	3	1	2	NA	NA	NA	NA	NA	NA	68,00	10
	2	14	18	11	NA	NA	NA	13	7	NA	NA	2	1	1	NA	NA	1	70,00	10
	8	22	35	20	NA	NA	NA	1	NA	2	1	2	1	1	NA	NA	NA	93,00	10

b-
279b
-1-
5p
sfr-
miR
b-
1049
5-1-
5p;
sfr-
miR
b-
1049
5-2-
5p;
sfr-
miR
b-
1049
5-3-
5p e
sfr-
miR
b-
1049
5-5-
5p
sfr-
miR
b-
79-
5p
sfr-
miR
b-

NA	2	1	NA	82	38	24	2	2	1	1	NA	NA	NA	1	NA	NA	154,00	10
91	273	425	215	4	1	NA	NA	NA	NA	1	2	1	NA	NA	NA	2	1015,00	10
2	1	1	1	3	2	1	1	3	NA	1	NA	NA	NA	NA	NA	4	20,00	11

1045 9-1- 5p e sfr- miR b- 1045 9-3- 5p sfr- miR b- 1049 2c-2- 3p sfr- miR b- 285- 3p sfr- miR b- 1047 7-2- 3p; sfr- miR b- 1047 7-3- 3p e sfr- miR b- 1047																			
	3	4	8	3	NA	1	NA	1	1	2	4	NA	NA	2	2	NA	NA	31,00	11
	3	9	9	1	5	5	1	1	1	NA	1	NA	NA	NA	NA	NA	1	37,00	11
	2	NA	NA	4	12	11	NA	6	12	2	2	NA	1	1	1	NA	NA	54,00	11

7-4-3p																			
sfr-miR b-1049	10	6	8	19	2	1	1	1	3	NA	NA	NA	NA	2	3	NA	NA	56,00	11
2b-2-5p																			
sfr-miR b-1045																			
4e-1-5p; sfr-miR b-1045																			
4e-2-5p; sfr-miR b-1045	NA	15	10	10	1	3	NA	4	NA	1	1	NA	NA	7	5	NA	1	58,00	11
4e-3-5p e sfr-miR b-1045																			
4e-4-5p																			
sfr-miR b-1045	7	19	7	7	5	4	NA	1	NA	1	3	NA	NA	4	4	NA	NA	62,00	11

4a- 10- 5p sfr- miR b- 1050 8b- 1-3p; sfr- miR b- 1050 8b- 2-3p; sfr- miR b- 1050 8b- 3-3p e sfr- miR b- 1050 8b- 4-3p																			
	10	5	4	23	NA	2	NA	2	9	3	2	NA	NA	3	3	NA	NA	66,00	11
sfr- miR b- 1045 4a-3- 5p sfr- miR b-	3	14	12	9	12	4	NA	1	NA	2	2	NA	NA	4	5	NA	NA	68,00	11
	9	14	3	13	17	18	4	1	NA	3	1	NA	NA	3	NA	NA	NA	86,00	11

1050
1-1-
3p;
sfr-
miR
b-
1050
1-2-
3p;
sfr-
miR
b-
1050
1-3-
3p;
sfr-
miR
b-
1050
1-4-
3p e
sfr-
miR
b-
1050
1-5-
3p
sfr-
miR
b-
1048
2-3p
sfr-
miR
b-
1045

2	10	12	5	NA	1	NA	NA	NA	4	2	18	31	10	6	NA	NA	101,00	11
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6	38	13	11	12	7	NA	1	NA	2	3	NA	NA	4	6	NA	NA	103,00	11
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4a-12-5p sfr-miR b-1045 4a-1-5p; sfr-miR b-1045 4a-11-5p; sfr-miR b-1045 4a-2-5p; sfr-miR b-1045 4a-4-5p; sfr-miR b-1045 4a-5-5p; sfr-miR	12	38	17	15	12	7	NA	1	NA	2	3	NA	NA	4	7	NA	NA	118,00	11
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b-10454a-6-5p; sfr-miR																			
b-10454a-7-5p; sfr-miR																			
b-10454a-8-5p e sfr-miR																			
b-10454a-9-5p sfr-miR-279a-5p	13	26	43	28	NA	NA	NA	1	NA	2	1	2	1	1	NA	NA	2	120,00	11
sfr-miR b-10496-3p	15	80	26	9	13	15	NA	1	2	1	2	NA	NA	NA	5	NA	NA	169,00	11
sfr-miR b-10500-1-	8	66	38	36	6	7	1	NA	2	2	1	NA	NA	NA	4	NA	NA	171,00	11

5p e
sfr-
miR
b-
1050
0-3-
5p
sfr-
miR
b-
276-
5p
sfr-
miR
b-
1048
3-2-
3p
sfr-
miR
b-
1050
4-3p
sfr-
miR
b-
1050
5-2-
5p
sfr-
miR
b-9c-
3p
sfr-
miR
b-

30	84	124	68	4	21	3	NA	1	NA	NA	1	NA	NA	NA	1	6	343,00	11
56	223	423	211	10	4	10	NA	1	3	2	NA	NA	NA	2	NA	NA	945,00	11
122	167	376	254	NA	1	NA	NA	2	2	NA	9	11	NA	5	NA	26	975,00	11
216	1117	8	2	30	20	5	NA	3	NA	2	NA	NA	1	1	NA	NA	1405,00	11
260	482	723	465	8	4	1	1	NA	2	1	NA	NA	NA	1	NA	NA	1948,00	11
2040	62664	85	15	44	27	10	NA	1	2	2	NA	NA	1	NA	NA	NA	64891,00	11

1050
5-1-
3p
sfr-
miR-
6094
-3p
sfr-
miR-
1045
4c-
3p
sfr-
miR
b-
1050
0-2-
3p
sfr-
miR
b-
1049
3-3-
3p;
sfr-
miR
b-
1049
3-4-
3p e
sfr-
miR
b-
1049
3-5-
3p

31753	170354	436089	100569	96	48	40	NA	1	1	NA	NA	NA	2	1	NA	NA	738954,00	11
1	2	NA	2	6	4	1	2	3	NA	3	NA	1	2	3	NA	NA	30,00	12
1	3	9	8	3	1	NA	2	1	1	2	NA	NA	1	5	NA	NA	37,00	12
8	1	1	5	5	5	1	6	10	NA	3	NA	NA	1	1	NA	NA	47,00	12

sfr-miR b-1048 9-1-5p	8	9	1	2	6	3	2	3	NA	2	2	NA	NA	6	9	NA	NA	53,00	12
sfr-miR b-1049 2c-6-5p	3	14	17	14	4	NA	3	2	3	5	2	NA	NA	1	2	NA	NA	70,00	12
sfr-miR b-1049 3-1-5p e sfr-miR b-1049 3-2-5p	3	1	1	NA	21	23	6	3	6	2	6	NA	NA	5	3	NA	NA	80,00	12
sfr-miR b-1045 4d-5p	6	24	12	28	2	4	NA	2	3	2	NA	NA	NA	2	1	NA	1	87,00	12
sfr-miR b-1045 4h-3-5p;	7	25	12	29	4	5	NA	2	4	2	NA	NA	NA	6	1	NA	1	98,00	12

sfr-miR b-1045 4h-6-5p e sfr-miR b-1045 4h-7-5p																			
sfr-miR b-1051 3-1-5p e sfr-miR b-1051 32-2-5p	4	9	34	10	15	7	1	1	4	8	NA	NA	NA	6	7	NA	NA	106,00	12
sfr-miR b-1045 4f-5p	16	27	14	41	9	5	NA	1	2	4	6	NA	NA	6	10	NA	NA	141,00	12
sfr-miR b-79-3p	102	251	644	257	NA	3	NA	1	3	4	NA	NA	1	NA	1	4	199	1470,00	12

sfr-miR b-1046 0-5p	551	224	711	906	9	23	3	13	11	1	1	NA	NA	NA	1	NA	NA	2454,00	12
sfr-miR b-283- 5p	1624	3023	5934	3659	4	13	3	1	NA	NA	1	NA	1	NA	1	NA	1	14265,00	12
sfr-miR-14- 5p e sfr-miR-14- 5p	1186	4857	6334	4570	79	6	2	2	2	6	3	NA	NA	NA	NA	NA	14	17061,00	12
sfr-miR b-263a-1- 5p e sfr-miR b-263a-2- 5p	39471	631314	180500	80681	31	4	5	NA	NA	1	NA	3	2	NA	NA	-	1	932013,00	12
sfr-miR b-1045 4h-5-3p	1	7	13	11	1	7	1	2	1	1	1	NA	NA	3	2	NA	NA	51,00	13

e sfr- miR b- 1045 4h- 6-3p																			
sfr- miR b- 1046 2-1- 3p; sfr- miR b- 1046 2-2- 3p; sfr- miR b- 1046 2-3- 3p; sfr- miR b- 1046 2-4- 3p; sfr- miR b- 1046 2-5- 3p e	1	3	3	4	5	5	2	4	4	12	3	NA	NA	6	8	NA	NA	60,00	13

sfr-
miR
b-
1046
2-6-
3p
sfr-
miR
b-
1050
1-1-
5p;
sfr-
miR
b-
1050
1-2-
5p;
sfr-
miR
b-
1050
1-3-
5p e
sfr-
miR
b-
1050
1-4-
5p
sfr-
miR
b-
1049
2a-1-
3p;

6	19	3	7	7	9	2	1	3	8	3	NA	NA	11	8	NA	NA	87,00	13
5	11	17	6	9	13	2	4	2	5	5	NA	NA	5	6	NA	NA	90,00	13

sfr- miR b- 1049 2a-2- 3p; sfr- miR b- 1049 2a-3- 3p e																			
sfr- miR b- 1049 2a-6- 3p																			
sfr- miR b- 1049 2c-4- 3p	5	12	20	7	7	14	2	4	1	7	5	NA	NA	6	7	NA	NA	97,00	13
sfr- miR b- 1049 2a-3- 5p	5	17	22	24	8	2	5	7	4	5	2	NA	NA	1	2	NA	NA	104,00	13
sfr- miR b- 1050 2-1- 5p e	3	10	7	9	36	6	4	6	6	7	5	NA	NA	NA	3	NA	2	104,00	13

sfr-miR b-1050 2-2-5p																			
sfr-miR b-1046 2-2-5p	2	27	30	11	36	15	11	3	4	6	2	NA	NA	5	11	NA	NA	163,00	13
sfr-miR b-1046 1b-1-5p	38	42	27	44	4	9	1	3	1	1	1	NA	NA	6	3	NA	NA	180,00	13
sfr-miR b-1046 1b-1-3p; sfr-miR b-1046 1b-2-3p; sfr-miR b-1046 1b-3-3p;	29	31	20	32	17	9	4	NA	8	4	9	NA	1	14	16	NA	NA	194,00	13

sfr-
miR
b-
1046
1b-
4-3p;
sfr-
miR
b-
1046
1b-
5-3p
e sfr-
miR
b-
1046
1b-
6-3p
sfr-
miR
b-
307-
3p
sfr-
miR-
1046
5-1-
3p
sfr-
miR
b-
34-
5p
sfr-
miR-

17	52	94	32	29	58	12	5	6	1	NA	NA	NA	NA	1	15	177	499,00	13
49	63	204	142	38	44	11	2	4	25	16	NA	NA	17	39	NA	NA	654,00	13
113	62	448	297	1	2	NA	4	5	3	3	NA	2	NA	NA	1	3	944,00	13
197	401	791	597	19	26	6	2	6	NA	1	NA	2	NA	NA	4	21	2073,00	13

13b-3p sfr-miR b-1-3p	311	414	1173	594	6	1	3	NA	1	3	6	NA	NA	NA	2	1	1	2516,00	13
sfr-miR b-1045 8-3p	878	4353	9481	2382	67	58	16	2	9	2	NA	NA	NA	NA	4	2	8	17262,00	13
sfr-miR b-1049 2c-1-5p; sfr-miR b-1049 2c-2-5p e sfr-miR b-1049 2c-3-5p	19	9	18	24	10	3	3	1	6	1	1	NA	NA	2	2	NA	1	100,00	14
sfr-miR b-1045 4f-3p	8	25	15	19	17	10	2	2	4	1	5	NA	1	6	7	NA	NA	122,00	14

sfr-miR b-1045 4b-3p	8	28	15	15	17	12	2	2	4	3	5	NA	1	6	8	NA	NA	126,00	14
sfr-miR b-932-5p	13	19	49	32	10	3	1	3	7	3	1	NA	NA	1	1	NA	9	152,00	14
sfr-miR b-1049 9b-1-3p	18	26	45	37	10	31	2	1	3	1	1	NA	1	2	3	NA	NA	181,00	14
e sfr-miR b-1049 9b-2-3p																			
sfr-miR b-1045 4a-13-3p	18	81	49	35	19	39	4	1	3	3	4	NA	1	7	13	NA	NA	277,00	14
sfr-miR b-1045 4e-3-3p	7	86	36	37	23	31	6	7	4	14	10	1	NA	20	19	NA	NA	301,00	14

sfr-
miR
b-
1045
4a-
12-
3p;
sfr-
miR
b-
1045
4a-2-
3p;
sfr-
miR
b-
1045
4a-3-
3p;
sfr-
miR
b-
1045
4a-4-
3p e
sfr-
miR
b-
1045
4a-5-
3p
sfr-
miR
b-
1045

28	86	56	53	23	39	4	2	3	4	5	NA	1	8	13	NA	NA	325,00	14
10	74	50	47	20	28	7	7	9	10	7	NA	NA	33	23	NA	2	327,00	14

4h-5-5p sfr-miR b-1046 2-1-5p; sfr-miR b-1046 2-4-5p e sfr-miR b-1046 2-6-5p sfr-miR b-1048 4-1-3p sfr-miR b-87-3p sfr-miR-1046 5-2-3p	25	185	68	32	144	53	55	11	10	8	3	NA	NA	7	15	NA	2	618,00	14
	1	NA	4	NA	64	113	14	189	225	145	130	6	5	262	257	NA	1	1416,00	14
	175	534	1308	539	16	29	4	1	1	NA	NA	1	1	1	NA	2	100	2712,00	14
	471	1485	3293	1414	28	26	5	3	2	18	13	NA	1	12	21	NA	NA	6792,00	14

sfr-miR b-1049 1-1-3p e sfr-miR b-1049 1-2-3p	2303	6063	17630	7835	803	991	190	212	269	35	24	NA	NA	36	38	NA	27	36456,00	14
sfr-miR b-277-3p	2728	7054	21953	7118	4	4	NA	3	4	5	NA	1	2	1	1	NA	1	38879,00	14
sfr-miR b-7b-5p	22	7	18	14	25	20	5	3	5	5	3	NA	5	2	1	NA	4	139,00	15
sfr-miR-1049 9a-1-3p; sfr-miR-1049 9a-2-3p; sfr-miR-1049 9a-3-	93	144	221	177	9	30	2	1	3	1	1	NA	1	2	3	1	NA	689,00	15

3p; sfr- miR- 1049 9a-4- 3p e sfr- miR- 1049 9a-5- 3p																			
sfr- miR b- 1049 9b- 3-3p	93	144	221	177	9	30	2	1	3	1	1	NA	1	2	3	1	NA	689,00	15
sfr- miR b- 1049 1-1- 5p e sfr- miR b- 1049 1-2- 5p																			
sfr- miR b-2a- 2-3p sfr- miR	9	45	99	45	331	376	91	13	10	22	19	NA	NA	53	59	8	3	1183,0 0	15
sfr- miR b-2a- 2-3p	291	1817	1962	774	1198	567	257	5	5	NA	1	1	1	NA	1	14	109	7003,0 0	15
sfr- miR	435	2283	2975	1137	1210	575	258	5	5	NA	1	1	1	NA	1	14	109	9010,0 0	15

b-2a-1-3p																			
sfr-miR b-993a-1-3p	925	3072	7863	2596	21	14	4	11	8	8	5	NA	NA	7	5	1	16	14556,00	15
sfr-miR b-71-3p	1631	8367	17816	5612	67	67	7	4	4	3	2	3	1	NA	1	NA	1	33586,00	15
sfr-miR b-278-5p	1502	28188	24263	6210	115	16	17	1	NA	4	5	5	5	1	2	1	NA	60335,00	15
sfr-miR b-2755-3p	6097	18247	30346	12625	189	259	63	4	6	20	15	5	5	4	7	NA	NA	67892,00	15
sfr-miR b-92a-3p	44	376	379	166	61	78	14	28	49	50	41	11	17	24	49	NA	6	1393,00	16
sfr-miR b-745-5p	429	1204	2747	1122	127	28	18	3	5	1	2	1	NA	2	1	2	153	5845,00	16
sfr-miR b-	1026	1394	1866	1903	46	80	15	39	44	31	18	5	2	21	38	NA	15	6543,00	16

993a																			
-1-																			
5p e																			
sfr-																			
miR																			
b-																			
993a																			
-2-																			
5p																			
sfr-																			
miR																			
b-																			
2766	414	3279	4051	1006	160	89	37	2	2	12	8	14	19	2	5	NA	73	9173,00	16
-5p																			
sfr-																			
miR																			
b-																			
279-	683	2799	5463	2223	201	222	45	6	15	4	6	6	7	2	3	NA	26	11711,00	16
3p																			
sfr-																			
miR																			
b-																			
278-	611	2619	7329	1918	3	3	1	2	7	5	6	7	15	7	6	NA	3	12542,00	16
3p																			
sfr-																			
miR																			
b-																			
92b-	492	7037	5996	1822	122	290	40	251	278	823	629	165	211	304	470	NA	7	18937,00	16
3p																			
sfr-																			
miR																			
b-																			
970-	834	4851	15827	2816	237	106	30	8	4	8	1	1	3	NA	2	1	20	24749,00	16
3p																			
sfr-																			
miR																			
	3655	4734	12667	9493	52	207	15	11	26	59	25	73	72	37	40	NA	5	31171,00	16

b-1048																			
2-5p																			
sfr-miR																			
b-2c-	3193	17588	37303	12698	571	556	148	17	25	5	1	8	2	1	2	NA	11	72129,00	16
3p																			
sfr-miR																			
b-	12892	29455	85190	37084	266	201	48	15	19	5	3	3	4	8	5	NA	4	165202,00	16
276-																			
3p																			
sfr-miR																			
b-	87	468	630	93	31	13	2	12	26	11	11	4	5	8	15	1	120	1537,00	17
2763																			
-3p																			
sfr-miR																			
b-	251	801	1708	689	117	135	33	1	11	5	3	5	8	7	2	3	158	3937,00	17
2756																			
-5p																			
sfr-miR																			
b-	206	927	1317	502	164	202	48	145	209	11	9	39	59	43	40	24	413	4358,00	17
308-																			
3p																			
sfr-miR																			
b-	146	808	1410	420	10	75	8	31	30	7	3	9	5	4	6	7	1604	4583,00	17
745-																			
3p																			
sfr-miR																			
b-	465	1920	5914	1784	143	480	27	152	180	201	181	1	4	175	212	25	267	12131,00	17

1048																			
3-5p sfr- miR																			
b- 317- 3p	2592	5572	11413	7650	321	396	50	44	69	11	5	3	5	7	8	2	9	28157,00	17
sfr- miR																			
b- 10- 1-3p	3385	4869	12579	7687	46	40	16	14	16	1	7	15	6	3	8	3	6	28701,00	17
sfr- miR- 279a	3953	6782	12172	7813	53	83	13	13	21	6	11	9	11	2	11	5	13	30971,00	17
-3p																			
sfr- miR																			
b- 279b	3953	6782	12172	7813	53	83	13	13	21	6	11	9	11	2	11	5	13	30971,00	17
-1- 3p																			
sfr- miR																			
b- 279d	2735	7941	14419	6601	48	98	10	55	73	78	54	97	107	112	148	47	295	32918,00	17
-3p																			
sfr- miR																			
b- 279b	2983	7557	19176	7387	9	8	1	2	10	2	1	3	4	2	5	1	4	37155,00	17
-2- 3p																			
sfr- miR																			
b-	9	95	110	27	49	36	2	269	342	4177	3347	14741	18797	3220	2847	3	8	48079,00	17

2779
-1-
5p e
sfr-
miR
b-
2779
-2-
5p
sfr-
miR
b-
306a
-5p
sfr-
miR
b-
11-
3p
sfr-
miR
b-
2766
-3p
sfr-
miR
b-
10-
1-5p

5942	12557	28513	13258	31	82	14	55	60	25	17	18	30	21	30	1	13	60667,00	17
2950	18339	27372	6405	10530	3972	1869	8	18	18	16	8	2	4	7	1	8	71527,00	17
2401	10376	17317	4355	22829	20676	4161	1780	2095	282	250	140	177	306	337	146	1453	89081,00	17
303767	4111083	1346432	513258	2070	637	587	50	27	24	21	39	53	14	26	1	5	6278094,00	17

3 CONCLUSÃO

A biotecnologia oferece alternativas inovadoras que podem ser empregadas no controle de pragas de interesse agrônomo. Utilizando o PhD, selecionamos sete clones capazes de se ligar a proteínas intestinais de *S. frugiperda*. O clone SfF3, quando usado em combinação com o *B. thuringiensis* GF 07, promoveu um aumento da mortalidade em lagartas neonatas. Além disso, demonstrou similaridade com as proteínas da cassetes de ligação de ATP subfamília C2 e citocromo P450, as quais são importantes para a sobrevivência desta praga. Já para entender como os miRNAs podem ser usados como uma ferramenta no controle da praga, foram preditos 350 precursores de miRNA no genoma de *S. frugiperda*, sendo 60 específicos para o gênero *Spodoptera*. Além disso, o refinamento de bibliotecas de RNA-seq na busca de possíveis moléculas bioinseticidas forneceu 91 sequências ainda não descritas, evidenciando novas alternativas na modulação de transcritos para o controle biológico do inseto. Nossos resultados validam essas tecnologias como uma abordagem agrobiotecnológica para o controle de pragas e identifica novas moléculas com possível atividades bioinseticidas.

ANEXO A - Normas para revista Plos one

Style and Format

File format	<p>Manuscript files can be in the following formats: DOC, DOCX, or RTF. Microsoft Word documents should not be locked or protected.</p> <p>LaTeX manuscripts must be submitted as PDFs.</p>
Length	<p>Manuscripts can be any length. There are no restrictions on word count, number of figures, or amount of supporting information.</p> <p>We encourage you to present and discuss your findings concisely.</p>
Font	<p>Use a standard font size and any standard font, except for the font named “Symbol”. To add symbols to the manuscript, use the Insert → Symbol function in your word processor or paste in the appropriate Unicode character.</p>
Headings	<p>Limit manuscript sections and sub-sections to 3 heading levels. Make sure heading levels are clearly indicated in the manuscript text.</p>
Layout and spacing	<p>Manuscript text should be double-spaced.</p> <p>Do not format text in multiple columns.</p>
Page and line numbers	<p>Include page numbers and line numbers in the manuscript file. Use continuous line numbers (do not restart the numbering on each page).</p>
Footnotes	<p>Footnotes are not permitted. If your manuscript contains footnotes, move the information into the main text or the reference list, depending on the content.</p>
Language	<p>Manuscripts must be submitted in English.</p> <p>You may submit translations of the manuscript or abstract as supporting information.</p>

Abbreviations	<p>Define abbreviations upon first appearance in the text.</p> <p>Do not use non-standard abbreviations unless they appear at least three times in the text.</p> <p>Keep abbreviations to a minimum.</p>
Reference style	PLOS uses “Vancouver” style, as outlined in the ICMJE sample references.
Equations	<p>We recommend using MathType for display and inline equations, as it will provide the most reliable outcome. If this is not possible, Equation Editor or Microsoft's Insert→Equation function is acceptable.</p> <p>Avoid using MathType, Equation Editor, or the Insert→Equation function to insert single variables (e.g., “$a^2 + b^2 = c^2$”), Greek or other symbols (e.g., β, Δ, or ' [prime]), or mathematical operators (e.g., \times, \geq, or \pm) in running text. Wherever possible, insert single symbols as normal text with the correct Unicode (hex) values.</p> <p>Do not use MathType, Equation Editor, or the Insert→Equation function for only a portion of an equation. Rather, ensure that the entire equation is included. Equations should not contain a mix of different equation tools. Avoid “hybrid” inline or display equations, in which part is text and part is MathType, or part is MathType and part is Equation Editor.</p>
Nomenclature	<p>Use correct and established nomenclature wherever possible.</p> <p><i>Units of measurement</i> Use SI units. If you do not use these exclusively, provide the SI value in parentheses after each value.</p> <p><i>Drugs</i> Provide the Recommended International Non-Proprietary Name (rINN).</p> <p><i>Species names</i> Write in italics (e.g., <i>Homo sapiens</i>). Write out in full the genus and species, both in the title of the manuscript and at the first mention of an organism in a paper. After first mention, the first letter of the</p>

	genus name followed by the full species name may be used (e.g., <i>H. sapiens</i>).
<i>Genes, mutations, genotypes, and alleles</i>	Write in italics. Use the recommended name by consulting the appropriate genetic nomenclature database (e.g., HUGO for human genes). It is sometimes advisable to indicate the synonyms for the gene the first time it appears in the text. Gene prefixes such as those used for oncogenes or cellular localization should be shown in roman typeface (e.g., v-fes, c-MYC).
<i>Allergens</i>	The systematic allergen nomenclature of the World Health Organization/International Union of Immunological Societies (WHO/IUIS) Allergen Nomenclature Sub-committee should be used for manuscripts that include the description or use of allergenic proteins. For manuscripts describing new allergens, the systematic name of the allergen should be approved by the WHO/IUIS Allergen Nomenclature Sub-Committee prior to manuscript publication. Examples of the systematic allergen nomenclature can be found at the WHO/IUIS Allergen Nomenclature site.

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Prior to submission, authors who believe their manuscripts would benefit from professional editing are encouraged to use language-editing and copyediting services. Obtaining this service is the responsibility of the author, and should be done before initial submission. These services can be found on the web using search terms like “scientific editing service” or “manuscript editing service.”

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Middle section	<p><i>The following elements can be renamed as needed and presented in any order:</i></p> <ul style="list-style-type: none"> • Materials and Methods • Results • Discussion • Conclusions (optional)
Ending section	<p><i>The following elements are required, in order:</i></p> <ul style="list-style-type: none"> • Acknowledgments • References • Supporting information captions (if applicable)
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Symbol Legend		
Symbol	Name	Definition
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&	Ampersand	2nd set of equal contributors
*	Asterisk	Corresponding author(s)
#a	Pound/number sign	First Current address
#b	Pound/number sign	Second Current address
†	Dagger/Cross	Deceased
^	Caret	Consortium/Group Authorship

This is the article title

John Doe^{1¶}, Antonie Data^{1¶}, Johannes van Stats^{1,#a}, Marie Testperson^{2*}, David Ribosome Jr. 3,5, Gregory H.T. McBio 4,#b, Angela Reviewerson^{1,2&}, Marina Measure^{1&}, on behalf of The Bunny Genome Sequencing Consortium[^]

¹ Department, Institution, City, State, Country

² Department of Dermatology, Division of Rabbit Health, Section of Veterinary Medicine, St. Hare Hospital, San Francisco, California, United States of America

³ Department of Libraries and Archives, National Contemporary Bunny Museum, Lagomorph, Connecticut, United States of America

⁴ Department of Restoration, National Contemporary Bunny Museum, Lagomorph, Connecticut, United States of America

⁵ Department of Archaeology, Bunny University, Lagomorph, Connecticut, United States of America

#a Current Address: Department of Carrot Science, Bunny University, Lagomorph, Connecticut, United States of America

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¶ These authors contributed equally to this work.

& These authors also contributed equally to this work.

^ Membership of the Bunny Genome Sequencing Consortium is provided in the Acknowledgments.

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1 Abstract

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17 Introduction

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31 **Fig 1. This is the Fig 1 Title.** This is the Fig 1 legend.

32 **Fig 2. This is the Fig 2 Title.** This is the Fig 2 legend.

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Level 1 Heading

- Use Level 1 heading for all major sections (Abstract, Introduction, Materials and methods, Results, Discussion, etc.).
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- Use bold type for the figure titles.

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$$p^2 + 2pq + q^2 = 1 \quad (1)$$

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NOTE: This document is presented in single-space paragraph format for ease of use. Please submit your manuscript in double-space paragraph format.

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Lorem ipsum dolor sit amet, consectetur adipiscing elit. Vestibulum adipiscing urna ut lectus gravida, et bland Table 1. Donec tincidunt porta sem nec hendrerit. Vestibulum nec pharetra quam, vitae convalli. Fido nemo.

Table 1. This is the Table 1 Title.

	Chemical W	Chemical X	Chemical Y	Chemical Z
Chemical 1	Reaction 1W	Reaction 1X	Reaction 1Y	Reaction 1Z
Chemical 2	Reaction 2W	Reaction 2X	Reaction 2Y	Reaction 2Z
Chemical 3	Reaction 3W ^a	Reaction 3X	Reaction 3Y ^b	Reaction 3Z
Chemical 4	Reaction 4W	Reaction 4X	Reaction 4Y	Reaction 4Z
Chemical 5	Reaction 5W	Reaction 5X	Reaction 5Y	Reaction 5Z

This is the Table 1 legend.

^aTable footnotes belong here.

^bFootnotes should have corresponding symbols in the table.

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- Tables must be cell-based in Microsoft Word or embedded with Microsoft Excel.
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Conclusions

Lorem ipsum dolor sit amet, consectetur adipiscing [1-5]. Vestibulum adipiscing urna ut lectus gravida, vitae blandit tortor interdum. Donec tincidunt porta sem nec hendrerit. Vestibulum nec pharetra quam, vitae convallis nunc. Mauris in mattis sapien. Fusce sodales vulputate auctor S1 Fig. Dolor sit amet S1 and S2 Tables.

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83 Acknowledgments

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89 References

- 90 ○ Doe J, Data A, van Stats J, Testperson M, Ribosome D Jr,
 91 McBio GHT, et al. This is the article title. PLoS ONE.
 92 2017;12(12):e000000. doi: 10.1371/journal.pone.0000000
- 93 ○ Doe J, Data A, van Stats J, Testperson M, Ribosome D Jr,
 94 McBio GHT, et al. Bunny dynamics in cartoon landscapes.
 95 PLoS ONE. Forthcoming 2017.

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98 Supporting information

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102 **S2 Table. This is the S2 Table Title.** This is the S2 Table legend.

103 **S1 File. This is the S1 File Title.** This is the S1 File legend.

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Short title	100 characters	State the topic of the study	Cigarette smoke exposure and innate immunity SODIS and childhood diarrhoea

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Source	Format
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Books	Bates B. Bargaining for life: A social history of tuberculosis. 1st ed. Philadelphia: University of Pennsylvania Press; 1992.

Source	Format
Book chapters	Hansen B. New York City epidemics and history for the public. In: Harden VA, Risse GB, editors. AIDS and the historian. Bethesda: National Institutes of Health; 1991. pp. 21-28.
Deposited articles (preprint s, e-prints, or arXiv)	Krick T, Shub DA, Verstraete N, Ferreiro DU, Alonso LG, Shub M, et al. Amino acid metabolism conflicts with protein diversity; 1991. Preprint. Available from: arXiv:1403.3301v1. Cited 17 March 2014.
Published media (print or online newspapers and magazine articles)	Fountain H. For Already Vulnerable Penguins, Study Finds Climate Change Is Another Danger. The New York Times. 29 Jan 2014. Available from: http://www.nytimes.com/2014/01/30/science/earth/climate-change-taking-toll-on-penguins-study-finds.html Cited 17 March 2014.
New media (blogs, web sites, or other written works)	Allen L. Announcing PLOS Blogs. 2010 Sep 1 [cited 17 March 2014]. In: PLOS Blogs [Internet]. San Francisco: PLOS 2006 - . [about 2 screens]. Available from: http://blogs.plos.org/plos/2010/09/announcing-plos-blogs/ .
Masters' theses or doctoral dissertations	Wells A. Exploring the development of the independent, electronic, scholarly journal. M.Sc. Thesis, The University of Sheffield. 1999. Available from: http://cuminCAD.scix.net/cgi-bin/works/Show?2e09
Databases and repositories (Figshare, arXiv)	Roberts SB. QPX Genome Browser Feature Tracks; 2013 [cited 2013 Oct 5]. Database: figshare [Internet]. Available from: http://figshare.com/articles/QPX_Genome_Browser_Feature_Tracks/701214
Multimedia (videos, movies, or TV shows)	Hitchcock A, producer and director. Rear Window [Film]; 1954. Los Angeles: MGM.

Supporting Information

Authors can submit essential supporting files and multimedia files along with their manuscripts. All supporting information will be subject to peer review. All file types can be submitted, but files must be smaller than 10 MB in size.

Authors may use almost any description as the item name for a supporting information file as long as it contains an “S” and number. For example, “S1 Appendix” and “S2 Appendix,” “S1 Table” and “S2 Table,” and so forth.

Supporting information files are published exactly as provided, and are not copyedited.

Supporting information captions

List supporting information captions at the end of the manuscript file. Do not submit captions in a separate file.

The file number and name are required in a caption, and we highly recommend including a one-line title as well. You may also include a legend in your caption, but it is not required.

Example caption

S1 Text. Title is strongly recommended. Legend is optional.

In-text citations

We recommend that you cite supporting information in the manuscript text, but this is not a requirement. If you cite supporting information in the text, citations do not need to be in numerical order.

Figures and Tables

Figures

Do not include figures in the main manuscript file. Each figure must be prepared and submitted as an individual file.

Cite figures in ascending numeric order at first appearance in the manuscript file.

Figure captions

Figure captions must be inserted in the text of the manuscript, immediately following the paragraph in which the figure is first cited (read order). Do not include captions as part of the figure files themselves or submit them in a separate document.

At a minimum, include the following in your figure captions:

A figure label with Arabic numerals, and “Figure” abbreviated to “Fig” (e.g. Fig 1, Fig 2, Fig 3, etc). Match the label of your figure with the name of the file uploaded at submission (e.g. a figure citation of “Fig 1” must refer to a figure file named “Fig1.tif”).

A concise, descriptive title

The caption may also include a legend as needed.

Tables

Cite tables in ascending numeric order upon first appearance in the manuscript file.

Place each table in your manuscript file directly after the paragraph in which it is first cited (read order). Do not submit your tables in separate files.

Tables require a label (e.g., “Table 1”) and brief descriptive title to be placed above the table. Place legends, footnotes, and other text below the table.

Data reporting

All data and related metadata underlying the findings reported in a submitted manuscript should be deposited in an appropriate public repository, unless already provided as part of the submitted article.

Repositories may be either subject-specific (where these exist) and accept specific types of structured data, or generalist repositories that accept multiple data types. We recommend that authors select repositories appropriate to their field. Repositories may be subject-specific (e.g., GenBank for sequences and PDB for structures), general, or institutional, as long as DOIs or accession numbers are provided and the data are at least as open as CC BY. Authors are encouraged to select repositories that meet accepted criteria as trustworthy digital repositories, such as criteria of the Centre for Research Libraries or Data Seal of Approval. Large, international databases are more likely to persist than small, local ones.

To support data sharing and author compliance of the PLOS data policy, we have integrated our submission process with a select set of data repositories. The list is neither representative nor exhaustive of the suitable repositories available to authors. Current

repository integration partners include Dryad and FlowRepository. Please contact data@plos.org to make recommendations for further partnerships.

Instructions for PLOS submissions with data deposited in an integration partner repository:

Deposit data in the integrated repository of choice.

Once deposition is final and complete, the repository will provide you with a dataset DOI (provisional) and private URL for reviewers to gain access to the data.

Enter the given data DOI into the full Data Availability Statement, which is requested in the Additional Information section of the PLOS submission form. Then provide the URL passcode in the Attach Files section.

If you have any questions, please email us.

Accession numbers

All appropriate data sets, images, and information should be deposited in an appropriate public repository. See our list of recommended repositories.

Accession numbers (and version numbers, if appropriate) should be provided in the Data Availability Statement. Accession numbers or a citation to the DOI should also be provided when the data set is mentioned within the manuscript.

In some cases authors may not be able to obtain accession numbers of DOIs until the manuscript is accepted; in these cases, the authors must provide these numbers at acceptance. In all other cases, these numbers must be provided at submission.

Identifiers

As much as possible, please provide accession numbers or identifiers for all entities such as genes, proteins, mutants, diseases, etc., for which there is an entry in a public database, for example:

- Ensembl
- Entrez Gene
- FlyBase
- InterPro
- Mouse Genome Database (MGD)
- Online Mendelian Inheritance in Man (OMIM)
- PubChem

Identifiers should be provided in parentheses after the entity on first use.

Striking image

You can choose to upload a “Striking Image” that we may use to represent your article online in places like the journal homepage or in search results.

The striking image must be derived from a figure or supporting information file from the submission, i.e., a cropped portion of an image or the entire image. Striking images should ideally be high resolution, eye-catching, single panel images, and should ideally avoid containing added details such as text, scale bars, and arrows.

If no striking image is uploaded, we will designate a figure from the submission as the striking image.

Striking images should not contain potentially identifying images of people. Read our policy on identifying information.

The PLOS licenses and copyright policy also applies to striking images.

Additional Information Requested at Submission

Financial Disclosure Statement

This information should describe sources of funding that have supported the work. It is important to gather these details prior to submission because your financial disclosure statement cannot be changed after initial submission without journal approval. If your manuscript is published, your statement will appear in the Funding section of the article.

Enter this statement in the Financial Disclosure section of the submission form. Do not include it in your manuscript file.

The statement should include:

Specific grant numbers

Initials of authors who received each award

Full names of commercial companies that funded the study or authors

Initials of authors who received salary or other funding from commercial companies

URLs to sponsors’ websites

Also state whether any sponsors or funders (other than the named authors) played any role in:

Study design

Data collection and analysis

Decision to publish

Preparation of the manuscript

If they had no role in the research, include this sentence: “The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.”

If the study was unfunded, include this sentence as the Financial Disclosure statement: “The author(s) received no specific funding for this work.”

Competing Interests

This information should not be in your manuscript file; you will provide it via our submission system.

All potential competing interests must be declared in full. If the submission is related to any patents, patent applications, or products in development or for market, these details, including patent numbers and titles, must be disclosed in full.

Manuscripts disputing published work

For manuscripts disputing previously published work, it is *PLOS ONE* policy to invite a signed review by the disputed author during the peer review process. This procedure is aimed at ensuring a thorough, transparent, and productive review process.

If the disputed author chooses to submit a review, it must be returned in a timely fashion and contain a full declaration of all competing interests. The Academic Editor will consider any such reviews in light of the competing interest.

Authors submitting manuscripts disputing previous work should explain the relationship between the manuscripts in their cover letter, and will be required to confirm that they accept the conditions of this review policy before the manuscript is considered further.

Related manuscripts

Upon submission, authors must confirm that the manuscript, or any related manuscript, is not currently under consideration or accepted elsewhere. If related work has been submitted to *PLOS ONE* or elsewhere, authors must include a copy with the submitted article. Reviewers will be asked to comment on the overlap between related submissions.

We strongly discourage the unnecessary division of related work into separate manuscripts, and we will not consider manuscripts that are divided into “parts.” Each

submission to *PLOS ONE* must be written as an independent unit and should not rely on any work that has not already been accepted for publication. If related manuscripts are submitted to *PLOS ONE*, the authors may be advised to combine them into a single manuscript at the editor's discretion.

Preprints

PLOS encourages authors to post preprints as a way to accelerate the dissemination of research and supports authors who wish to share their work early and receive feedback before formal peer review. Deposition of manuscripts with preprint servers does not impact consideration of the manuscript at any PLOS journal.

Authors posting on bioRxiv may concurrently submit directly to PLOS journals through [bioRxiv's direct transfer to journal service](#).

Authors submitting manuscripts in the life sciences to *PLOS ONE* may opt-in to post their work on bioRxiv during the *PLOS ONE* initial submission process.

Guidelines for Specific Study Types

Human subjects research

All research involving human participants must have been approved by the authors' Institutional Review Board (IRB) or by equivalent ethics committee(s), and must have been conducted according to the principles expressed in the Declaration of Helsinki. Authors should be able to submit, upon request, a statement from the IRB or ethics committee indicating approval of the research. We reserve the right to reject work that we believe has not been conducted to a high ethical standard, even when formal approval has been obtained.

Subjects must have been properly instructed and have indicated that they consent to participate by signing the appropriate informed consent paperwork. Authors may be asked to submit a blank, sample copy of a subject consent form. If consent was verbal instead of written, or if consent could not be obtained, the authors must explain the reason in the manuscript, and the use of verbal consent or the lack of consent must have been approved by the IRB or ethics committee.

All efforts should be made to protect patient privacy and anonymity. Identifying information, including photos, should not be included in the manuscript unless the information is crucial and the individual has provided written consent by completing the Consent Form for Publication in a PLOS Journal (PDF). Download additional translations of the form from the

Downloads and Translations page. More information about patient privacy, anonymity, and informed consent can be found in the International Committee of Medical Journal Editors (ICMJE) Privacy and Confidentiality guidelines.

Manuscripts should conform to the following reporting guidelines:

- Studies of diagnostic accuracy: STARD
- Observational studies: STROBE
- Microarray experiments: MIAME
- Other types of health-related research: Consult the EQUATOR web site for

appropriate reporting guidelines

Methods sections of papers on research using human subjects or samples must include ethics statements that specify:

The name of the approving institutional review board or equivalent committee(s).

If approval was not obtained, the authors must provide a detailed statement explaining why it was not needed

Whether informed consent was written or oral. If informed consent was oral, it must be stated in the manuscript:

Why written consent could not be obtained

That the Institutional Review Board (IRB) approved use of oral consent

How oral consent was documented

For studies involving humans categorized by race/ethnicity, age, disease/disabilities, religion, sex/gender, sexual orientation, or other socially constructed groupings, authors should:

Explicitly describe their methods of categorizing human populations

Define categories in as much detail as the study protocol allows

Justify their choices of definitions and categories, including for example whether any rules of human categorization were required by their funding agency

Explain whether (and if so, how) they controlled for confounding variables such as socioeconomic status, nutrition, environmental exposures, or similar factors in their analysis

In addition, outmoded terms and potentially stigmatizing labels should be changed to more current, acceptable terminology. Examples: “Caucasian” should be changed to “white” or “of [Western] European descent” (as appropriate); “cancer victims” should be changed to “patients with cancer.”

For papers that include identifying, or potentially identifying, information, authors must download the Consent Form for Publication in a PLOS Journal, which the individual, parent, or guardian must sign once they have read the paper and been informed about the terms

of PLOS open-access license. The signed consent form should not be submitted with the manuscript, but authors should securely file it in the individual's case notes and the methods section of the manuscript should explicitly state that consent authorization for publication is on file, using wording like:

The individual in this manuscript has given written informed consent (as outlined in PLOS consent form) to publish these case details.

For more information about *PLOS ONE* policies regarding human subjects research, see the Publication Criteria and Editorial Policies.

Clinical trials

Clinical trials are subject to all policies regarding human research. *PLOS ONE* follows the World Health Organization's (WHO) definition of a clinical trial:

A clinical trial is any research study that prospectively assigns human participants or groups of humans to one or more health-related interventions to evaluate the effects on health outcomes [...] Interventions include but are not restricted to drugs, cells and other biological products, surgical procedures, radiologic procedures, devices, behavioural treatments, process-of-care changes, preventive care, etc.

All clinical trials must be registered in one of the publicly-accessible registries approved by the WHO or ICMJE (International Committee of Medical Journal Editors). Authors must provide the trial registration number. Prior disclosure of results on a clinical trial registry site will not affect consideration for publication. We reserve the right to inform authors' institutions or ethics committees, and to reject the manuscript, if we become aware of unregistered trials.

PLOS ONE supports prospective trial registration (i.e. before participant recruitment has begun) as recommended by the ICMJE's clinical trial registration policy. **Where trials were not publicly registered before participant recruitment began**, authors must:

Register all related clinical trials and confirm they have done so in the Methods section

Explain in the Methods the reason for failing to register before participant recruitment

Clinical trials must be reported according to the relevant reporting guidelines, i.e. CONSORT for randomized controlled trials, TREND for non-randomized trials, and other specialized guidelines as appropriate. The intervention should be described according to the requirements of the TIDieR checklist and guide. Submissions must also include the study protocol as supporting information, which will be published with the manuscript if accepted.

Authors of manuscripts describing the results of clinical trials must adhere to the CONSORT reporting guidelines appropriate to their trial design, available on the CONSORT Statement web site. Before the paper can enter peer review, authors must:

Provide the registry name and number in the methods section of the manuscript

Provide a copy of the trial protocol as approved by the ethics committee and a completed CONSORT checklist as supporting information (which will be published alongside the paper, if accepted). This should be named S1 CONSORT Checklist.

Include the CONSORT flow diagram as the manuscript's "Fig 1"

Any deviation from the trial protocol must be explained in the paper. Authors must explicitly discuss informed consent in their paper, and we reserve the right to ask for a copy of the patient consent form.

The methods section must include the name of the registry, the registry number, and the URL of your trial in the registry database for each location in which the trial is registered.

Animal research

All research involving vertebrates or cephalopods must have approval from the authors' Institutional Animal Care and Use Committee (IACUC) or equivalent ethics committee(s), and must have been conducted according to applicable national and international guidelines. Approval must be received prior to beginning research.

Manuscripts reporting animal research must state in the Methods section:

The full name of the relevant ethics committee that approved the work, and the associated permit number(s).

Where ethical approval is not required, the manuscript should include a clear statement of this and the reason why. Provide any relevant regulations under which the study is exempt from the requirement for approval.

Relevant details of steps taken to ameliorate animal suffering.

Example ethics statement

This study was carried out in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. The protocol was approved by the Committee on the Ethics of Animal Experiments of the University of Minnesota (Protocol Number: 27-2956). All surgery was performed under sodium pentobarbital anesthesia, and all efforts were made to minimize suffering.

Authors should always state the organism(s) studied in the Abstract. Where the study may be confused as pertaining to clinical research, authors should also state the animal model in the title.

To maximize reproducibility and potential for re-use of data, we encourage authors to follow the Animal Research: Reporting of *In Vivo* Experiments (ARRIVE) guidelines for all submissions describing laboratory-based animal research and to upload a completed ARRIVE Guidelines Checklist to be published as supporting information.

Non-human primates

Manuscripts describing research involving non-human primates must report details of husbandry and animal welfare in accordance with the recommendations of the Weatherall report, The use of non-human primates in research, including:

Information about housing, feeding, and environmental enrichment.

Steps taken to minimize suffering, including use of anesthesia and method of sacrifice, if appropriate.

Random source animals

Manuscripts describing studies that use random source (e.g. Class B dealer-sourced in the USA), shelter, or stray animals will be subject to additional scrutiny and may be rejected if sufficient ethical and scientific justification for the study design is lacking.

Unacceptable euthanasia methods and anesthetic agents

Manuscripts reporting use of a euthanasia method(s) classified as unacceptable by the American Veterinary Medical Association or use of an anesthesia method(s) that is widely prohibited (e.g., chloral hydrate, ether, chloroform) must include at the time of initial submission, scientific justification for use in the specific study design, as well as confirmation of approval for specific use from their animal research ethics committee. These manuscripts may be subject to additional ethics considerations prior to publication.

Humane endpoints

Manuscripts reporting studies in which death of a regulated animal (vertebrate, cephalopod) is a likely outcome or a planned experimental endpoint, must comprehensively report details of study design, rationale for the approach, and methodology, including consideration of humane endpoints. This applies to research that involves, for instance,

assessment of survival, toxicity, longevity, terminal disease, or high rates of incidental mortality.

Definition of a humane endpoint

A humane endpoint is a predefined experimental endpoint at which animals are euthanized when they display early markers associated with death or poor prognosis of quality of life, or specific signs of severe suffering or distress. Humane endpoints are used as an alternative to allowing such conditions to continue or progress to death following the experimental intervention (“death as an endpoint”), or only euthanizing animals at the end of an experiment. Before a study begins, researchers define the practical observations or measurements that will be used during the study to recognize a humane endpoint, based on anticipated clinical, physiological, and behavioral signs. Please see the NC3Rs guidelines for more information. Additional discussion of humane endpoints can be found in this article: Nuno H. Franco, Margarida Correia-Neves, I. Anna S. Olsson (2012) How “Humane” Is Your Endpoint? — Refining the Science-Driven Approach for Termination of Animal Studies of Chronic Infection. *PLoS Pathog* 8(1): e1002399 doi.org/10.1371/journal.ppat.1002399.

Full details of humane endpoints use must be reported for a study to be reproducible and for the results to be accurately interpreted.

For studies in which death of an animal is an outcome or a planned experimental endpoint, authors should include the following information in the Methods section of the manuscript:

The specific criteria (i.e. humane endpoints) used to determine when animals should be euthanized.

The duration of the experiment.

The numbers of animals used, euthanized, and found dead (if any); the cause of death for all animals.

How frequently animal health and behavior were monitored.

All animal welfare considerations taken, including efforts to minimize suffering and distress, use of analgesics or anaesthetics, or special housing conditions.

If humane endpoints were not used, the manuscript should report:

A scientific justification for the study design, including the reasons why humane endpoints could not be used, and discussion of alternatives that were considered.

Whether the institutional animal ethics committee specifically reviewed and approved the anticipated mortality in the study design.

Observational and field studies

Methods sections for submissions reporting on any type of field study must include ethics statements that specify:

Permits and approvals obtained for the work, including the full name of the authority that approved the study; if none were required, authors should explain why

Whether the land accessed is privately owned or protected

Whether any protected species were sampled

Full details of animal husbandry, experimentation, and care/welfare, where relevant

Paleontology and archaeology research

Manuscripts reporting paleontology and archaeology research must include descriptions of methods and specimens in sufficient detail to allow the work to be reproduced. Data sets supporting statistical and phylogenetic analyses should be provided, preferably in a format that allows easy re-use.

Specimen numbers and complete repository information, including museum name and geographic location, are required for publication. Locality information should be provided in the manuscript as legally allowable, or a statement should be included giving details of the availability of such information to qualified researchers.

If permits were required for any aspect of the work, details should be given of all permits that were obtained, including the full name of the issuing authority. This should be accompanied by the following statement:

All necessary permits were obtained for the described study, which complied with all relevant regulations.

If no permits were required, please include the following statement:

No permits were required for the described study, which complied with all relevant regulations.

Manuscripts describing paleontology and archaeology research are subject to the following policies:

- Sharing of data and materials. Any specimen that is erected as a new species, described, or figured must be deposited in an accessible, permanent repository (i.e., public museum or similar institution). If study conclusions depend on specimens that do not fit these criteria, the article will be rejected under PLOS ONE's data availability criterion.
- Ethics. PLOS ONE will not publish research on specimens that were obtained without necessary permission or were illegally exported.

Systematic reviews and meta-analyses

A systematic review paper, as defined by The Cochrane Collaboration, is a review of a clearly formulated question that uses explicit, systematic methods to identify, select, and critically appraise relevant research, and to collect and analyze data from the studies that are included in the review. These reviews differ substantially from narrative-based reviews or synthesis articles. Statistical methods (meta-analysis) may or may not be used to analyze and summarize the results of the included studies.

Reports of systematic reviews and meta-analyses must include a completed PRISMA (Preferred Reporting Items for Systematic Reviews and Meta-Analyses) checklist and flow diagram to accompany the main text. Blank templates are available here:

Checklist: PDF or Word document

Flow diagram: PDF or Word document

Authors must also state in their “Methods” section whether a protocol exists for their systematic review, and if so, provide a copy of the protocol as supporting information and provide the registry number in the abstract.

If your article is a systematic review or a meta-analysis you should:

State this in your cover letter

Select “Research Article” as your article type when submitting

Include the PRISMA flow diagram as Fig 1 (required where applicable)

Include the PRISMA checklist as supporting information

Meta-analysis of genetic association studies

Manuscripts reporting a meta-analysis of genetic association studies must report results of value to the field and should be reported according to the guidelines presented in Systematic Reviews of Genetic Association Studies by Sagoo *et al.*

On submission, authors will be asked to justify the rationale for the meta-analysis and how it contributes to the base of scientific knowledge in the light of previously published results. Authors will also be asked to complete a checklist (DOCX) outlining information about the justification for the study and the methodology employed. Meta-analyses that replicate published studies will be rejected if the authors do not provide adequate justification.

Personal data from third-party sources

For all studies using personal data from internet-based and other third-party sources (e.g., social media, blogs, other internet sources, mobile phone companies), data must be

collected and used according to company/website Terms and Conditions, with appropriate permissions. All data sources must be acknowledged clearly in the Materials and Methods section.

In the Ethics Statement, authors should declare any potential risks to individuals or individual privacy, or affirm that in their assessment, the study posed no such risks. In addition, the following Ethics and Data Protection requirements must be met.

For interventional studies, which impact participants' experiences or data, the study design must have been prospectively approved by an Ethics Committee, and informed consent is required. The Ethics Committee may waive the requirement for approval and/or consent.

For observational studies in which personal experiences and accounts are not manipulated, consultation with an Ethics or Data Protection Committee is recommended. Additional requirements apply in the following circumstances:

If information used could threaten personal privacy or damage the reputation of individuals whose data are used, an Ethics Committee should be consulted and informed consent obtained or specifically addressed.

If authors accessed any personal identifying information, an Ethics or Data Protection Committee should oversee data anonymization. If data were anonymized and/or aggregated before access and analysis, informed consent is generally not required.

Note that Terms of Use contracts do not qualify as informed consent, even if they address the use of personal data for research.

Cell lines

Authors reporting research using cell lines should state when and where they obtained the cells, giving the date and the name of the researcher, cell line repository, or commercial source (company) who provided the cells, as appropriate.

Authors must also include the following information for each cell line:

For *de novo* (new) cell lines, including those given to the researchers as a gift, authors must follow our policies for human subjects research or animal research, as appropriate. The ethics statement must include:

Details of institutional review board or ethics committee approval; AND

For human cells, confirmation of written informed consent from the donor, guardian, or next of kin

For established cell lines, the Methods section should include:

A reference to the published article that first described the cell line; AND/OR

The cell line repository or company the cell line was obtained from, the catalogue number, and whether the cell line was obtained directly from the repository/company or from another laboratory

Authors should check established cell lines using the ICLAC Database of Cross-contaminated or Misidentified Cell Lines to confirm they are not misidentified or contaminated. Cell line authentication is recommended – e.g., by karyotyping, isozyme analysis, or short tandem repeats (STR) analysis – and may be required during peer review or after publication.

Blots and gels

Manuscripts reporting results from blots (including Western blots) and electrophoretic gels should follow these guidelines:

In accordance with our policy on image manipulation, the image should not be adjusted in any way that could affect the scientific information displayed, e.g. by modifying the background or contrast.

All blots and gels that support results reported in the manuscript should be provided.

Original uncropped and unadjusted blots and gels, including molecular size markers, should be provided in either the figures or the supplementary files.

Lanes should not be overcropped around the bands; the image should show most or all of the blot or gel. Any non-specific bands should be shown and an explanation of their nature should be given.

The image should include all relevant controls, and controls should be run on the same blot or gel as the samples.

A figure panel should not include composite images of bands originating from different blots or gels. If the figure shows non-adjacent bands from the same blot or gel, this should be clearly denoted by vertical black lines and the figure legend should provide details of how the figure was made.

Antibodies

Manuscripts reporting experiments using antibodies should include the following information:

The name of each antibody, a description of whether it is monoclonal or polyclonal, and the host species.

The commercial supplier or source laboratory.

The catalogue or clone number and, if known, the batch number.

The antigen(s) used to raise the antibody.

For established antibodies, a stable public identifier from the Antibody Registry.

The manuscript should also report the following experimental details:

The final antibody concentration or dilution.

A reference to the validation study if the antibody was previously validated. If not, provide details of how the authors validated the antibody for the applications and species used.

We encourage authors to consider adding information on new validations to a publicly available database such as Antibodypedia or CiteAb.

Small and macromolecule crystal data

Manuscripts reporting new and unpublished three-dimensional structures must include sufficient supporting data and detailed descriptions of the methodologies used to allow the reproduction and validation of the structures. All novel structures must have been deposited in a community endorsed database prior to submission (please see our list of recommended repositories).

Small molecule single crystal data

Authors reporting X-Ray crystallographic structures of small organic, metal-organic, and inorganic molecules must deposit their data with the Cambridge Crystallographic Data Centre (CCDC), the Inorganic Crystal Structure Database (ICSD), or similar community databases providing a recognized validation functionality. Authors are also required to include the relevant structure reference numbers within the main text (e.g. the CCDC ID number), as well as the crystallographic information files (.cif format) as Supplementary Information, along with the checkCIF validation reports that can be obtained via the International Union of Crystallography (IUCr).

Macromolecular structures

Authors reporting novel macromolecular structures must have deposited their data prior to submission with the Worldwide Protein Data Bank (wwPDB), the Biological Magnetic Resonance Data Bank (BMRB), the Electron Microscopy Data Bank (EMDB), or other community databases providing a recognized validation functionality. Authors must include the structure reference numbers within the main text and submit as Supplementary Information the official validation reports from these databases.

Methods, software, databases, and tools

PLOS ONE will consider submissions that present new methods, software, databases, or tools as the primary focus of the manuscript if they meet the following criteria:

Utility

The tool must be of use to the community and must present a proven advantage over existing alternatives, where applicable. Recapitulation of existing methods, software, or databases is not useful and will not be considered for publication. Combining data and/or functionalities from other sources may be acceptable, but simpler instances (i.e. presenting a subset of an already existing database) may not be considered. For software, databases, and online tools, the long-term utility should also be discussed, as relevant. This discussion may include maintenance, the potential for future growth, and the stability of the hosting, as applicable.

Validation

Submissions presenting methods, software, databases, or tools must demonstrate that the new tool achieves its intended purpose. If similar options already exist, the submitted manuscript must demonstrate that the new tool is an improvement over existing options in some way. This requirement may be met by including a proof-of-principle experiment or analysis; if this is not possible, a discussion of the possible applications and some preliminary analysis may be sufficient.

Availability

If the manuscript's primary purpose is the description of new software or a new software package, this software must be open source, deposited in an appropriate archive, and conform to the Open Source Definition. If the manuscript mainly describes a database, this database must be open-access and hosted somewhere publicly accessible, and any software used to generate a database should also be open source. If relevant, databases should be open for appropriate deposition of additional data. Dependency on commercial software such as Mathematica and MATLAB does not preclude a paper from consideration, although complete open source solutions are preferred. In these cases, authors should provide a direct link to the deposited software or the database hosting site from within the paper. If the primary focus of a manuscript is the presentation of a new tool, such as a newly developed or modified

questionnaire or scale, it should be openly available under a license no more restrictive than CC BY.

Software submissions

Manuscripts whose primary purpose is the description of new software must provide full details of the algorithms designed. Describe any dependencies on commercial products or operating system. Include details of the supplied test data and explain how to install and run the software. A brief description of enhancements made in the major releases of the software may also be given. Authors should provide a direct link to the deposited software from within the paper.

Database submissions

For descriptions of databases, provide details about how the data were curated, as well as plans for long-term database maintenance, growth, and stability. Authors should provide a direct link to the database hosting site from within the paper.

New taxon names

Zoological names

When publishing papers that describe a new zoological taxon name, PLOS aims to comply with the requirements of the International Commission on Zoological Nomenclature (ICZN). Effective 1 January 2012, the ICZN considers an online-only publication to be legitimate if it meets the criteria of archiving and is registered in ZooBank, the ICZN's official registry.

For proper registration of a new zoological taxon, we require two specific statements to be included in your manuscript.

In the **Results** section, the globally unique identifier (GUID), currently in the form of a Life Science Identifier (LSID), should be listed under the new species name, for example:

Anochetus boltoni Fisher *sp. nov.* urn:lsid:zoobank.org:act:B6C072CF-1CA6-40C7-8396-534E91EF7FBB

You will need to contact Zoobank to obtain a GUID (LSID). Please do this as early as possible to avoid delay of publication upon acceptance of your manuscript. It is your responsibility to provide us with this information so we can include it in the final published paper.

Please also insert the following text into the **Methods** section, in a sub-section to be called “Nomenclatural Acts”:

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Solanum aspersum S.Knapp, sp. nov. [urn:lsid:ipni.org:names:77103633-1] Type: Colombia. Putumayo: vertiente oriental de la Cordillera, entre Sachamates y San Francisco de Sibundoy, 1600-1750 m, 30 Dec 1940, J. Cuatrecasas 11471 (holotype, COL; isotypes, F [F-1335119], US [US-1799731]).

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Hymenogaster huthii. Stielow et al. 2010, sp. nov.
[urn:lsid:indexfungorum.org:names:518624]

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ANEXO B - Normas para revista **Journal of Invertebrate Pathology**

Introduction

The *Journal of Invertebrate Pathology* publishes articles on all aspects of original research concerned with the causation and manifestation (including immunologic responses) of infectious and noninfectious diseases of invertebrates, the suppression of such diseases in beneficial species, and the use of these pathogens in controlling undesirable species such as agricultural pests and vectors of pathogens transmissible to other organisms. In addition, this journal publishes the results of biochemical, physiological, morphological, genetic, and ecological studies related to the etiologic agents of diseases of invertebrates. The journal is particularly dedicated to the publication of contributions of a basic and fundamental nature, although it will accept suitable articles pertaining to the applications of invertebrate pathology. The editor-in-chief and members of the Editorial Board will examine contributions from any qualified worker in any country of the world.

Types of paper

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Include keywords

All figures (include relevant captions)

All tables (including titles, description, footnotes)

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